

APPLICATION

FOR

UNITED STATES LETTERS PATENT

METHOD FOR PRODUCING IMMUNOGLOBULINS CONTAINING  
PROTECTION PROTEINS AND THEIR USES

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DESCRIPTIONMETHODS FOR PRODUCING IMMUNOGLOBULINS CONTAINING  
PROTECTION PROTEINS IN PLANTS AND THEIR USECROSS REFERENCE TO RELATED APPLICATIONS

5        This is a continuation of co-pending application  
Serial No. 08/434,000 filed May 4, 1995, which is a  
continuation-in-part of co-pending application Serial  
No. 08/367,395 filed December 30, 1994, each of which is  
hereby incorporated by reference in its entirety  
10   including drawings.

FIELD OF INVENTION

The present invention relates to expression of  
immunoglobulins in plants that contain a protection  
protein as well as to transgenic plants that express  
15   such immunoglobulins. The therapeutic use of these  
immunoglobulins is also contemplated.

BACKGROUND TO THE INVENTION

Monoclonal antibodies have great potential for  
numerous therapeutic purposes. The advantages of  
20   monoclonal antibody therapeutics over conventional  
pharmaceuticals include their exquisite selectivity,  
multiple effector functions, and ease of molecular  
manipulation such as radioisotope labelling and other  
types of conjugation. A wide variety of target antigens  
25   have been used to generate specific monoclonal  
antibodies. See for example Therapeutic Monoclonal  
Antibodies, C. A. K. Borrebaeck and J.W. Larrick eds.,  
Stockton Press, New York, 1990, and The Pharmacology of  
Monoclonal Antibodies, M. Rosenberg and G.P. Moore eds.,  
30   Springer-Verlag, Berlin, 1994.

One therapeutic application of monoclonal  
antibodies is passive immunotherapy in which the  
exogenously produced immunoglobulins are administered

directly to the animal being treated by injection or by ingestion. To be successful, passive immunotherapy must deliver an appropriate amount of an immunoglobulin to the animal, because passive immunotherapy does not rely  
5 on an immune response in the animal being treated. The immunoglobulins administered must be specific for the pathogen or molecule desired to effect treatment. One advantage of passive immunotherapy is the speed at which the antibody can be contacted with the target compared  
10 to a normal immune response. Passive immunotherapy can also be used as a prophylaxis to prevent the onset of diseases or infections.

A major potential use of passive immunotherapy is in combating bacterial infections. Recent emergence of  
15 anti-biotic resistant bacteria make treatment of bacterial infections with passive immunotherapy desirable. Antibiotic treatment targeted to a single pathogen often involves eradication of a large population of normal microbes, and this can have  
20 undesired side effects. An alternative approach has been to utilize the inherent specificity of immunoglobulins to inhibit a specific pathogenic function in very specific microbial populations. In this strategy, purified immunoglobulins of the  
25 appropriate specificity would be administered in order to provide a passive barrier to pathogen invasion.

In addition, the immunoglobulins used for passive immunotherapies for example, for oral administration of immunoglobulins must meet certain requirements. First,  
30 the immunoglobulin must be functional in very harsh environments, such as the gastrointestinal tract. Second, the immunoglobulin must be resistant to the

actions of proteases so that it will not be degraded prior to inactivating the target.

Certain types of cells, including epithelial cells and hepatocytes, are capable of assembling immuno-  
5 globulin molecules which have been specifically adapted to function in harsh environments. These immuno-  
globulins are referred to as secretory immunoglobulins (SIg) and include both secretory IgA (SIgA) and secretory IgM (SIgM). The protection provided by  
10 endogenous secretory immunoglobulins have been demonstrated. Several mechanisms for protection from bacterial infection by secretory immunoglobulins have been proposed, including, but not limited to, direct  
15 killing, agglutination, inhibition of epithelial attachment and invasion, inactivation of enzymes and toxins, opsonization, and complement activation. In an animal, endogenously produced SIgA are exposed to very harsh environments where numerous proteases, such as intestinal and bacterial enzymes are extremely active  
20 and denaturants, such as stomach acid, are also present.

One component of secretory immunoglobulins, the secretory component, helps to protect the immunoglobulin against these inactivating agents thereby increasing the biological effectiveness of secretory immunoglobulin.

25 The mechanism of synthesis and assembly of these secretory immunoglobulins, such as SIgA or SIgM is extremely complex. In animal cells, secretory immunoglobulins are assembled in a process involving different cell types. Each secretory immunoglobulin is  
30 made up of immunoglobulin heavy and light chains, joining chain (J chain) and a secretory component. The immunoglobulin producing B cells make and assemble the immunoglobulin heavy and light chain together with J



chain to produce dimeric or polymeric IgM or IgA. The secretory component is produced by a second type of cell, either epithelial cells or hepatocytes, and secretory immunoglobulin is assembled in and secreted from these cells. The mechanism by which these cells assemble and secrete the secretory immunoglobulin is extremely complex and requires a unique microenvironment provided, for example, by mucosal tissues. The microenvironment places the B cells that produce the polymeric immunoglobulin near the cells that assemble and secrete secretory immunoglobulin onto the mucosal surface of an animal.

The epithelial cells have a receptor, the poly-immunoglobulin receptor (pIgR), that specifically recognizes and binds polymeric immunoglobulin/containing J chain, internalizing it and transporting it through the epithelial cell. Expressed on the basolateral cell surface, the pIgR has an N-terminal signal peptide of 18 amino acids, an extracellular polyimmunoglobulin binding portion of 629 amino acids, a membrane spanning segment of 23 hydrophobic residues, and a cytoplasmic tail of 103 amino acids. The extra-cellular portion contains five immunoglobulin-like domains of 100-111 amino acids each and constitutes the secreted form of the molecule. See for example, Mostov, Ann. Rev. Immunol., 12:63-84 (1994) The site at which the polyimmunoglobulin receptor is cleaved to generate mature secretory component has not been accurately determined.

The polyimmunoglobulin receptor is located on the basolateral surface of epithelial cells in animals. Polymeric, J chain-containing immunoglobulins produced in B cells interact with and are bound by the receptor resulting in vesicularization, transport across the

epithelial cell, and ultimate secretion to the mucosal surface. Transepithelial transport also involves proteolysis and phosphorylation to produce the mature SIg containing the secretory component. The close  
5 association of the required cells found in the mucosal microenvironment, specifically the B lymphocytes and epithelial cells, is required for secretory immunoglobulin assembly.

The targeting of the production of immunoglobulins  
10 in transgenic organisms, such as mice, is extremely difficult and transgenic organisms made from fungus or plants do not contain the proper cell types and mucosal microenvironment to produce secretory immunoglobulins. The production of large amounts of secretory immuno-  
15 globulins in transgenic organisms and cell culture has, before this invention, been impossible. One desiring to produce a secretory immunoglobulin in cell culture or a transgenic organism must express the immunoglobulin heavy chain, the immunoglobulin light chain, and J chain  
20 in a B lymphocyte. To mimic the proper mucosal microenvironment a cell having the pIgR receptor on its surface would also have to be present and be in close association with that B lymphocyte to even attempt to assemble a functional secretory immunoglobulin.

25 This elaborate process required for natural secretory immunoglobulin assembly is extremely difficult to duplicate in cell culture or transgenic organisms. Production of SIg in cell culture or transgenic organisms would require coupling the functions of cells  
30 producing immunoglobulin with the functions of epithelial cells in artificial (*in vitro*) systems. Moreover, if the desired transgenic organism is a fungus, a bacterium, or a plant, the cell types and

pathways of receptor-mediated cellular internalization, transcytosis, and secretion simply are not present. Those organisms lack epithelial cells and the required mucosal microenvironment.

5       To date only the assembly of immunoglobulins having light, heavy and J chain within the same cell has been reported. See Carayannopoulos et al. Proc. Nat Acad. Sci., U.S.A., 91:8348-8352 (1994). However, the assembly of an immunoglobulin having the additional  
10 protein component, secretory component, within a single cell has not been described.

      The present invention discloses a novel method for the assembly of these complex molecules. Rather than assemble the tetrameric complex at the epithelial cell  
15 surface by the interaction of a membrane bound polyimmunoglobulin receptor with immunoglobulin, we have assembled secretory immunoglobulin composed of alpha, J, and kappa immunoglobulin chains associated with a protection protein derived from pIgR. This invention  
20 produces transgenic plants that assemble secretory immunoglobulins with great efficiency. The present invention makes passive immunotherapy economically feasible.

#### 25                   SUMMARY OF THE INVENTION

      The present invention contemplates a new type of immunoglobulin molecule. Immunoglobulins of the present invention contain a protection protein in association with an immunoglobulin derived heavy chain having at  
30 least a portion of an antigen binding domain. In other embodiments, the immunoglobulin of the present invention further comprise an immunoglobulin derived light chain

having at least a portion of an antigen binding domain associated with the immunoglobulin derived heavy chain.

The protection proteins of the present invention give the immunoglobulins containing these protein useful  
5 properties including resistance to chemical and enzymatic degradation and resistance to denaturation. These protection proteins enhanced the resistance of the immunoglobulins to environmental conditions.

The protection proteins of the proteins of the  
10 present invention comprise at least a segment of amino acid residues 1 to 606 of native polyimmunoglobulin receptor (pIgR) of any species. Other useful protection proteins include protection proteins that contain portions of the pIgR molecule. For example, the  
15 protection protein may comprise all or part of: amino acids 1-118 (domain I of rabbit pIgR), amino acids 1 to 223 (domains I and II of rabbit pIgR); amino acids 1 to 332 (domains I, II, III of rabbit pIgR); amino acids 1 to 441 (domains I, II, III, and IV rabbit of pIgR);  
20 amino acids 1 to 552 (domains I, II, III, IV and V of rabbit pIgR); and amino acids 1 to 606 or 1 to 627 of pIgR. Additional amino acids, derived either from the pIgR sequence 653-755, or from other sources, may be included so long as they do not constitute a functional  
25 transmembrane spanning segment.

In other preferred embodiments, the immunoglobulins of the present invention have a protection protein which has a first amino acid sequence which substantially  
corresponds to at least a portion of the amino acid  
30 residues 1 to 606 or 1 to 627 of the rabbit polyimmuno-  
globulin receptor and has a second amino acid residue sequence contiguous with said first amino acid sequence, wherein said second amino acid residue sequence does not

have an amino acid residue sequence corresponding to the transmembrane segment of the rabbit polyimmunoglobulin receptor.

In more preferred embodiments, the second amino acid residue sequence has at least a portion of an amino acid sequence which corresponds to amino acid residues 655 to 755 of a polyimmunoglobulin receptor. In other preferred embodiments, the second amino acid residue is at least a portion of one or more of the following: an intracellular domain of a polyimmunoglobulin molecule, a domain of a member of the immunoglobulin gene superfamily, an enzyme, a toxin, or a linker.

The present invention contemplates protection proteins which do not have an amino acid residue corresponding to the transmembrane segment of rabbit polyimmunoglobulin receptor but may have amino acid residues corresponding to the intracellular domain of the rabbit polyimmunoglobulin receptor and this are deletion mutants of the receptor.

The present invention also contemplates immunoglobulins containing protection proteins which have an amino acid sequence which does not contain amino acid residues of a polyimmunoglobulin receptor from a species which are analogous to amino acid residues 288 to 755 of the rabbit immunoglobulin receptor, but does contain at least a portion of the amino acid residues or the domains from a polyimmunoglobulin receptor of a species which are analogous to one or more of these amino acid segments: Amino acids corresponding to amino acid residues 20-45 of the rabbit polyimmunoglobulin receptor; amino acids corresponding to or analogous to amino acid residues 1 to 120 of the rabbit polyimmunoglobulin receptor; amino acids corresponding to or

analogous to amino acid residues numbers 120 - 230 of the rabbit immunoglobulin receptor; amino acids corresponding to or analogous to amino acid residues numbers 230 - 340 of the rabbit polyimmunoglobulin receptor; amino acids corresponding to or analogous to amino acid residues 340 - 456 of the rabbit polyimmunoglobulin receptor; amino acids corresponding to or analogous to amino acid residues numbers 450 - 550 to 570 of the rabbit polyimmunoglobulin receptors; amino acids corresponding to or analogous to amino acid residues 550 to 570 - 606 to 627 of the rabbit polyimmunoglobulin receptor.

The protection proteins of the present invention may be derived from many species and include protection proteins derived from mammals, rodents, humans, bovine, porcine, ovine, fowl, caprine, mouse, rat, guinea pig, chicken or other bird and rabbit.

In preferred embodiments, the immunoglobulins of the present invention contain two or four immunoglobulin derived heavy chains having at least a portion of an antigen binding domain associated with the protection protein and two or four immunoglobulin derived light chains having at least a portion of an antigen binding domain bound to the each of the immunoglobulin derived heavy chains.

In other preferred embodiments, the immunoglobulins of the present invention further comprise immunoglobulin J chain bound to at least one of the immunoglobulin derived heavy chains. In preferred embodiments, the component parts of the immunoglobulins of the present invention are bound together by hydrogen bonds, disulfide bonds, covalent bonds, ionic interactions or combinations of said bonds. In other preferred

embodiments, the immunoglobulin of the present invention contain protection proteins and/or immunoglobulin derived heavy, light or J chains that are free from N-linked and/or O-linked oligosaccharides.

5       The immunoglobulins of the present invention may be used as therapeutic immunoglobulins against, for example, mucosal pathogen antigens. In preferred embodiments, the immunoglobulins of the present invention are capable of preventing dental caries by  
10 binding to an antigen from S. mutans serotypes c, e and f; and *S. sobrinus* stereotype d and g, using older nomenclature *S. mutans* a, c, d, e, f, g and h.

      The present invention also contemplates a eukaryotic cell, including a plant cell, containing an  
15 immunoglobulin of the present invention. Eukaryotic cells, including plant cells, containing a nucleotide sequence encoding a protection protein and a nucleotide sequence encoding an immunoglobulin derived heavy chain having at least a portion of an antigen binding domain  
20 is also contemplated. Eukaryotic cells, including plant cells, that additionally contain a nucleotide sequence encoding an immunoglobulin derived light chain having at least a portion of an antigen binding domain is also contemplated. In preferred embodiments, the eukaryotic  
25 cells, including plant cells, of the present invention contain nucleotide sequences that encode immunoglobulins that have an antigen binding domain is capable of binding an antigen from *S. mutans* serotypes a, c, d, e, f, and g, h (*S. mutans* serotypes c, e and f and *S.*  
30 *sobrinus* serotypes d and g under new nomenclature. The nucleotide sequences include RNA and appropriate DNA molecules arranged for expression.

In preferred embodiments, the plant cells of the present invention are part of a plant such as a whole plant. The present invention contemplates the use of all types of plants, both dicotyledonous and  
5 monocotyledonous including alfalfa, and tobacco.

The present invention also contemplates compositions comprising an immunoglobulin of the present invention and plant macromolecules derived from one of the plants useful in practicing the present invention.  
10 Particularly contemplated are compositions containing ribulose biphosphate carboxylase, light harvesting complex, pigments, secondary metabolites or chlorophyll and an immunoglobulin of the present invention. Preferred compositions have an immunoglobulin concentration of between 0.001% and 99.9% mass excluding water.  
15 In more preferred embodiments, the immunoglobulin concentrations present in the composition is between 0.1% and 99%. Other preferred compositions have plant macromolecules present in a concentration of between 1% and 99% mass excluding water.  
20

The present invention also contemplates methods for making an immunoglobulin of the present invention comprising introducing into a plant cell an expression vector having a nucleotide sequence encoding a  
25 protection protein operably linked to a transcriptional promoter; and introducing into the same plant cell an expression vector containing a nucleotide sequence encoding an immunoglobulin derived heavy chain having at least a portion of an antigen binding domain, operably  
30 linked to a transcriptional promoter. Other methods that further include the step of introducing into the same plant cell an expression vector containing a nucleotide sequence encoding an immunoglobulin derived



light chain having at least a portion of an antigen binding domain, operably linked to a transcriptional promoter. Other preferred methods include also introducing into a plant cell an expression vector  
5 containing a nucleotide sequence encoding an immunoglobulin J chain operably linked to a transcriptional promoter.

The present invention also contemplates methods for producing assembled immunoglobulins having heavy, light  
10 and J chains and a protection protein by introducing into a eukaryotic cell nucleotide sequences operatively linked for expression to encode an immunoglobulin derived heavy chain having at least a portion of an antigen binding domain, an immunoglobulin light chain  
15 having at least a portion of an antigen binding domain, and immunoglobulin J chain, and a protection protein. The method further comprises maintaining the eukaryotic cell under conditions allowing the production and assembly of the immunoglobulin derived heavy and light  
20 chains together with the immunoglobulin J chain and the protection protein to form an immunoglobulin containing a protection protein.

The present invention also contemplates methods of making an immunoglobulin resistant to various environ-  
25 mental conditions (more stable) and harsh conditions by operatively linking a nucleotide sequence encoding at least a portion of a desirable antigen binding domain derived from an immunoglobulin heavy chain to a nucleotide sequence encoding at least one domain derived  
30 from an immunoglobulin  $\mu$  or  $\alpha$  (IgM or IgA) heavy chain (or other immunoglobulin having increased stability in the environment) to form a nucleotide sequence encoding a chimeric immunoglobulin heavy chain and expressing

that nucleotide sequence in a eukaryotic which also contains at least one molecule from the following list: a protection protein, an immunoglobulin derived light chain having at least a portion of an antigen binding domain and an immunoglobulin J chain. The method further comprises allowing the chimeric immunoglobulin heavy chain to assemble with the other molecule present in the same cell to form an immunoglobulin which is resistant to environmental conditions and more stable.

10       The large scale production of immunoglobulins of the present invention is contemplated by growing the plants of the present invention and extracting the immunoglobulins from those plants. In preferred embodiments, the method of producing therapeutic  
15 immunoglobulin compositions containing plant macromolecules includes the step of shearing under pressure a portion of a plant of the present invention to produce a pulp containing a therapeutic immunoglobulin and plant macromolecules in an liquid derived from the apoplast or  
20 symplast of the plant and solid plant derived material. Further processing steps are contemplated which include separating the solid plant derived material from the liquid and using a portion of the plant including a leaf, stem, root, tuber, flower, fruit, seed or entire  
25 plant. The present invention contemplates the use of a mechanical device or enzymatic method which releases liquid from the apoplast or symplast of said plant followed optionally by separating using centrifugation, settling, flocculation or filtration.

30       The present invention contemplates immunoglobulins that are chimeric and thus they contain immunoglobulin domains derived from different immunoglobulin molecules.

Particularly preferred are immunoglobulins containing domains from IgG, IgM and IgA.

The present invention contemplates immunoglobulins where the immunoglobulin derived heavy chain is  
5 comprised of immunoglobulin domains from two different isotopes of immunoglobulin. In preferred embodiments, the immunoglobulin domains used include at least the C<sub>H</sub>1, C<sub>H</sub>2, or C<sub>H</sub>3 domain of mouse IgG, IgG1, IgG2a, IgG2b, IgG3, IgA, IgE, or IgD or the C<sub>var</sub> domain. In other  
10 preferred embodiments, the immunoglobulin heavy chain is comprised of at least the C<sub>μ</sub>1, C<sub>μ</sub>2, C<sub>μ</sub>3 or C<sub>μ</sub>4 domain of mouse IgM.

The present invention also contemplates immunoglobulin derived heavy chains made up of immunoglobulin  
15 domains include at least the C<sub>H</sub>1, C<sub>H</sub>2, or C<sub>H</sub>3 domain of a human IgG, IgG1, IgG2, IgG3, IgG4, IgA1, IgA2, or IgD; or least the C<sub>μ</sub>1, C<sub>μ</sub>2, C<sub>μ</sub>3 or C<sub>μ</sub>4 domain of human IgM; or the C<sub>var</sub> domain. The use of immunoglobulin domains derived from mammals, animals or rodents including any  
20 IgG isotype, any IgA isotype, IgE, IgM or IgD is contemplated.

The present invention also contemplates transgenic organisms which are comprised of cells containing four different transgenes each encoding a  
25 different polypeptide of a multi-peptide molecule wherein at least one of those peptides is associated together to form a multi-peptide molecule. The transgenic organisms contemplated by the present invention include transgenic organisms which contain as one of the four transgenes  
30 present a transgene encoding a protection protein. The protection protein present in the transgenic organism's cells is able to assemble together with immunoglobulin

heavy chains when present to form immunoglobulins which contain the protection protein.

In preferred transgenic organisms, the cells of the organism express four transgenes which encode an immunoglobulin derived heavy chain having at least a portion of an antigen binding domain, an immunoglobulin derived light chain having at least a portion of an antigen binding domain, an immunoglobulin J chain, and a protection protein. In other preferred transgenic organisms, the cells contain a transgene which encodes a chimeric immunoglobulin heavy chain, an immunoglobulin heavy chain derived from an IgA heavy chain, an immunoglobulin derived from an IgM heavy chain or an immunoglobulin derived from some other isotype of heavy chain.

In the most preferred embodiment, the transgenic organisms of the present invention are a plant. Various types and species of plants are contemplated by the present invention. In addition, the present invention also contemplates mammals which are transgenic organisms containing the various molecules of the present invention. Mammalian transgenic organisms are contemplated by the present invention and include mammalian transgenic organisms which contain four transgenes encoding different polypeptides.

#### BRIEF DESCRIPTION OF THE DRAWINGS

The drawings will first briefly be described.

FIGURE 1 illustrates synthetic oligonucleotides J1-J5 (restriction enzyme sites are underlined) that were used to amplify DNA fragments for Guy's 13 and alpha chain domains in the construction of hybrid IgG/A heavy chains. The relative positions of the areas encoded by

each oligonucleotide are shown diagrammatically The resulting recombinant heavy chains produced by combining various DNA fragments expressed in plants are also shown.

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## DETAILED DESCRIPTION OF THE INVENTION

### A. DEFINITIONS

Dicotyledon (dicot): A flowering plant whose embryos have two seed halves or cotyledons. Examples of  
10 dicots are: tobacco; tomato; the legumes including alfalfa; oaks; maples; roses; mints; squashes; daisies; walnuts; cacti; violets; and buttercups.

Monocotyledon (monocot): A flowering plant whose embryos have one cotyledon or seed leaf. Examples of  
15 monocots are: lilies; grasses; corn; grains, including oats, wheat and barley; orchids; irises; onions and palms.

Lower plant: Any non-flowering plant including ferns, gymnosperms, conifers, horsetails, club mosses,  
20 liver warts, hornworts, mosses, red algae, brown algae, gametophytes, sporophytes of pteridophytes, and green algae.

Eukaryotic hybrid vector: A DNA by means of which a DNA coding for a polypeptide (insert) can be  
25 introduced into a eukaryotic cell.

Extrachromosomal ribosomal DNA (rDNA): A DNA found in unicellular eukaryotes outside the chromosomes, carrying one or more genes coding for ribosomal RNA and replicating autonomously (independent of the replication  
30 of the chromosomes).

Palindromic DNA: A DNA sequence with one or more centers of symmetry.

DNA: Deoxyribonucleic acid.

T-DNA: A segment of transferred DNA.

rDNA: Ribosomal DNA.

RNA: Ribonucleic acid.

rRNA: Ribosomal RNA.

5     Ti-plasmid: Tumor-inducing plasmid.

Ti-DNA: A segment of DNA from Ti-plasmid.

Insert: A DNA sequence foreign to the rDNA,  
consisting of a structural gene and optionally  
additional DNA sequences.

10     Structural gene: A gene coding for a polypeptide  
and being equipped with a suitable promoter, termination  
sequence and optionally other regulatory DNA sequences,  
and having a correct reading frame.

15     Signal Sequence: A DNA sequence coding for an  
amino acid sequence attached to the polypeptide which  
binds the polypeptide to the endoplasmic reticulum and  
is essential for protein secretion.

20     (Selective) Genetic marker: A DNA sequence coding  
for a phenotypical trait by means of which transformed  
cells can be selected from untransformed cells.

25     Promoter: A recognition site on a DNA sequence or  
group of DNA sequences that provide an expression  
control element for a gene and to which RNA polymerase  
specifically binds and initiates RNA synthesis  
(transcription) of that gene.

30     Inducible promoter: A promoter where the rate of  
RNA polymerase binding and initiation is modulated by  
external stimuli. Such stimuli include light, heat,  
anaerobic stress, alteration in nutrient conditions,  
presence or absence of a metabolite, presence of a  
ligand, microbial attack, wounding and the like.

Viral promoter: A promoter with a DNA sequence  
substantially similar to the promoter found at the 5'

end of a viral gene. A typical viral promoter is found at the 5' end of the gene coding for the p21 protein of MMTV described by Huang et al., Cell, 27:245 (1981).

Other examples include the promoters found in the 35S transcript of the cauliflower mosaic virus as described by Benfey et al., Science, 250:959 (1990).

Synthetic promoter: A promoter that was chemically synthesized rather than biologically derived. Usually synthetic promoters incorporate sequence changes that optimize the efficiency of RNA polymerase initiation.

Constitutive promoter: A promoter where the rate of RNA polymerase binding and initiation is approximately constant and relatively independent of external stimuli. Examples of constitutive promoters include the cauliflower mosaic virus 35S and 19S promoters described by Poszkowski et al., EMBO J., 3:2719 (1989) and Odell et al., Nature, 313:810 (1985).

Regulated promoter: A promoter where the rate of RNA polymerase binding and initiation is modulated at a specific time during development, or in a specific structure of an organism or both of these types of modulation. Examples of regulated promoters are given in Chua et al., Science, 244:174-181 (1989).

Single-chain antigen-binding protein: A polypeptide composed of an immunoglobulin light-chain variable region amino acid sequence ( $V_L$ ) tethered to an immunoglobulin heavy-chain variable region amino acid sequence ( $V_H$ ) by a peptide that links the carboxyl terminus of the  $V_L$  sequence to the amino terminus of the  $V_H$  sequence.

Generally any combination of the heavy chain and light chain antigen binding domains into the same polypeptide using a linker polypeptide to allow the binding domains to assume a useful conformation. Such combinations

include  $V_H$ -Linker- $V_L$ ,  $V_H$ -Linear-Light chain, or  $V_L$ -Linear-Fd.

Single-chain antigen-binding protein-coding gene: A recombinant gene coding for a single-chain antigen-binding protein.

Polypeptide and peptide: A linear series of amino acid residues connected one to the other by peptide bonds between the alpha-amino and carboxy groups of adjacent residues.

Protein: A linear series of greater than about 50 amino acid residues connected one to the other as in a polypeptide.

Immunoglobulin product: A polypeptide, protein or protein containing at least the immunologically active portion of an immunoglobulin heavy chain and is thus capable of specifically combining with an antigen. Exemplary immunoglobulin products are an immunoglobulin heavy chain, immunoglobulin molecules, substantially intact immunoglobulin molecules, any portion of an immunoglobulin that contains the paratope, including those portions known in the art as Fab fragments, Fab' fragment,  $F(ab')_2$  fragment and Fv fragment.

Immunoglobulin molecule: A protein containing the immunologically active portions of an immunoglobulin heavy chain and immunoglobulin light chain covalently coupled together and capable of specifically combining with antigen.

Immunoglobulin derived heavy chain: A polypeptide that contains at least a portion of the antigen binding domain of an immunoglobulin and at least a portion of a variable region of an immunoglobulin heavy chain or at least a portion of a constant region of an immunoglobulin heavy chain. Thus, the immunoglobulin derived heavy



chain has significant regions of amino acid sequence homology with a member of the immunoglobulin gene superfamily. For example, the heavy chain in an Fab fragment is an immunoglobulin derived heavy chain.

5       Immunoglobulin derived light chain: A polypeptide that contains at least a portion of the antigen binding domain of an immunoglobulin and at least a portion of the variable region or at least a portion of a constant region of an immunoglobulin light chain. Thus, the  
10 immunoglobulin derived light chain has significant regions of amino acid homology with a member of the immunoglobulin gene superfamily.

Antigen binding domain: The portion of an immunoglobulin polypeptide that specifically binds to the  
15 antigen. This antigen is typically bound by antigen binding domains of the immunoglobulin heavy and light chain. However, antigen binding domains may be present on a single polypeptide.

J chain: Is a polypeptide that is involved in the  
20 polymerization of immunoglobulins and transport of polymerized immunoglobulins through epithelial cells. See, The Immunoglobulin Helper: The J Chain in Immunoglobulin Genes, at pg. 345, Academic Press (1989). J chain is found in pentameric IgM and dimeric IgA and  
25 typically attached via disulphide bonds. J chain has been studied in both mouse and human.

Fab fragment: A protein consisting of the portion of an immunoglobulin molecule containing the immunologically active portions of an immunoglobulin heavy  
30 chain and an immunoglobulin light chain covalently coupled together and capable of specifically combining with antigen. Fab fragments are typically prepared by proteolytic digestion of substantially intact

immunoglobulin molecules with papain using methods that are well known in the art. However an Fab fragment may also be prepared by expressing in a suitable host cell the desired portions of immunoglobulin heavy chain and  
5 immunoglobulin light chain using methods well known in the art.

F<sub>v</sub> fragment: A protein consisting of the immunologically active portions of an immunoglobulin heavy chain variable region and an immunoglobulin light  
10 chain variable region covalently coupled together and capable of specifically combining with antigen. F<sub>v</sub> fragments are typically prepared by expressing in suitable host cell the desired portions of immunoglobulin heavy chain variable region and immunoglobulin light  
15 chain variable region using methods well known in the art.

Asexual propagation: Producing progeny by regenerating an entire plant from leaf cuttings, stem cuttings, root cuttings, single plant cells (protoplasts)  
20 or callus.

Self-pollination: The transfer of pollen from male flower parts to female flower parts on the same plant. This process typically produces seed.

Cross-pollination: The transfer of pollen from the  
25 male flower parts of one plant to the female flower parts of another plant. This process typically produces seed from which viable progeny can be grown.

Epitope: A portion of a molecule that is specifically recognized by an immunoglobulin product. It  
30 is also referred to as the determinant or antigenic determinant.

Chimeric immunoglobulin heavy chain: An immunoglobulin derived heavy chain having at least a portion of

its amino acid sequence derived from an immunoglobulin heavy chain of a different isotype or subtype or some other peptide, polypeptide or protein. Typically, a chimeric immunoglobulin heavy chain has its amino acid residue sequence derived from at least two different isotypes or subtypes of immunoglobulin heavy chain.

Transgene: A gene that has been introduced into the germ line of an animal. The gene may be introduced into the animal at an early developmental stage. However, the gene could be introduced into the cells of an animal at a later stage by, for example, a retroviral vector.

Multiple molecule: A molecule comprised of more than one peptide or polypeptide associated together by any means including chemical bonds.

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#### B. IMMUNOGLOBULINS CONTAINING PROTECTION PROTEINS

The present invention provides novel methods for producing immunoglobulin molecules containing protection proteins. The immunoglobulins contain a protection protein in association with an immunoglobulin derived heavy chain that has at least a portion of an antigen binding domain.

The protection proteins of the present invention have an amino acid sequence substantially corresponding to or analogous to at least a portion of residues 1 to 627 of the amino acid residue sequence of the rabbit polyimmunoglobulin receptor and is derived from a precursor protein that does not contain the amino acid residue sequence greater than amino acid residue 627 or analogous to amino acid residue 627 of the rabbit polyimmunoglobulin receptor. The nucleotide sequence and the amino acid sequence of the rabbit polyimmunoglobulin receptor are now and have been described by the Mostov et

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al., Nature, 308:37 (1984) and EMBL/Gene Bank K01291.  
The nucleotide sequence of the polyimmunoglobulin  
receptor is SEQ ID NO. 1 and the corresponding amino acid  
residue sequence is SEQ ID NO. 2.

5       The polyimmunoglobulin receptors from any species  
may be used as a protection protein and these protection  
proteins do not contain and are derived from a precursor  
protein that does not contain amino acids having numbers  
greater than the amino acid number analogous to amino  
10   acids 1-627 of the rabbit immunoglobulin sequence. In  
preferred embodiments, the protection protein is derived  
from any species and precursor protein that contains  
amino acids analogous to at least a portion of amino  
acids 1-606 of the rabbit polyimmunoglobulin receptor and  
15   does not contain amino acid residues analogous to  
residues 607-755 of the rabbit polyimmunoglobulin  
receptor.

      The human polyimmunoglobulin receptor sequence has  
been determined and reported by Krajci et al., Eur. J.  
20   Immunol., 22:2309-2315 (1992) and Krajci et al., Biochem.  
Biophys. Res. Comm., 158:783-789 (1989) and EMBL/Gene  
Bank Accession No. X73079. The nucleotide sequence of  
the human polyimmunoglobulin receptor is SEQ ID NO. 3 and  
the corresponding amino acid residue sequence is SEQ ID  
25   NO. 4. The human polyimmunoglobulin receptor shows  
extensive sequence homology and has an analogous domain  
structure to that of the rabbit polyimmunoglobulin  
receptor. See, Kraehenbuhl et al., Trends in Cell Biol.,  
2:170 (1992). The portions of the human polyimmuno-  
30   globulin receptor which are analogous to the domains  
and/or amino acid residues sequence of the rabbit  
polyimmunoglobulin receptor are shown in Table 1.

The rat polyimmunoglobulin receptor sequence has been determined and reported by Banting et al., FEBS Lett., 254:177-183 (1989) and EMBL/Gene Bank Accession No. X15741. The nucleotide of the rat polyimmunoglobulin receptor nucleotide sequence is SEQ ID NO. 9 and the corresponding amino acid residue sequence is SEQ ID NO 10. The rat polyimmunoglobulin receptor shows extensive sequence homology and has an analogous domain structure to that of the rabbit and human polyimmunoglobulin receptor. See, Kraehenbuhl et al., T. Cell Biol., 2:170 (1992). The portions of the rat polyimmunoglobulin receptor which are analogous to the domains and/or amino acid residue sequence of the rabbit polyimmunoglobulin receptor are shown in Table 1.

The bovine polyimmunoglobulin receptor sequence has been determined and reported in EMBL/Gene Bank Accession No. X81371. The bovine polyimmunoglobulin receptor nucleotide sequence is SEQ ID NO.5 and the corresponding amino acid residue sequence is SEQ ID NO. 6. The bovine polyimmunoglobulin receptor shows extensive sequence homology and has an analogous domain structure to that of the rabbit and human polyimmunoglobulin receptor. The portions of the bovine polyimmunoglobulin receptor which are analogous to the domains and/or amino acid residues sequence of the rabbit polyimmunoglobulin receptor are shown in Table 1.

The mouse polyimmunoglobulin receptor sequence has been determined and reported by Piskurich et al., J. Immunol., 150:38 (1993) and EMBL/Gene Bank U06431. The mouse polyimmunoglobulin receptor nucleotide is SEQ ID NO. 7 and the corresponding amino acid residue sequence is SEQ ID NO. 8. The mouse polyimmunoglobulin receptor shows extensive sequence homology and has an analogous

domain structure to that of the rabbit and human polyimmunoglobulin receptor. The portions of the mouse polyimmunoglobulin receptor which are analogous to the domains and/or amino acid residue sequence of the rabbit polyimmunoglobulin receptor are shown in Table 1.

In addition to the above-identified nucleic acid and corresponding amino acid residue sequences of the polyimmunoglobulin receptor from a variety of species, the present invention contemplates the use of a portion of a polyimmunoglobulin receptor from any species. The conserved domain structure of the polyimmunoglobulin receptor between species allows the selection of analogous amino acid residue sequences within each polyimmunoglobulin receptor from different species. The present invention contemplates the use of such analogous amino acid residue sequences from any polyimmunoglobulin receptor. The analogous sequences from several polyimmunoglobulin receptor amino acid sequences is as shown in Table 1.

Table 1 Analogous Regions of the Amino Acid Residue Sequence  
of The Polymunoglobulin Receptor of Several  
Species. The nucleotide sequence coordinates  
approximately define the boundaries of the domains of  
molecules.

5

<u>Rat</u>	<u>Mouse</u>	<u>Rabbit</u>	<u>Bovine</u>	<u>Human</u>
(SEQ ID		(SEQ ID	(SEQ ID	(SEQ ID
10		(SEQ ID		

The protection proteins of the present invention may contain substantially less than the entire amino acid residue sequence of the polyimmunoglobulin receptor. In preferred embodiments the protection protein contains at least a portion of the amino acid residues 1 to 606 of the native polyimmunoglobulin receptor of rabbit. Unlike the native polyimmunoglobulin receptor, the protection proteins of the present invention are derived from precursor proteins that do not contain the entire amino acid residue sequence greater than the amino acid residue 627 derived from the native polyimmunoglobulin receptor and thus may contain more amino acids or fewer amino acids than secretory components. In preferred embodiments, the protection proteins of the present invention do not contain the entire amino acid residue sequence greater than amino acid residue 606 of the native polyimmunoglobulin receptor of rabbit. The present invention contemplates using only portions of the native polyimmunoglobulin receptor sequence as a protection protein. In other embodiments, it is contemplated that the protection protein may end at any amino acid between amino acid residue 606 to 627, including every amino acid position between 606 and 627, such as 607, 608, 609, 610, 611, 612, 613, 614, 615, 616, 617, 618, 619, 620, 621, 622, 623, 624, 625, 626.

In preferred embodiments, a protection protein of the present invention has an amino acid sequence which corresponds to one or more of the following amino acid segments:

- 1) amino acids (AA) corresponding to AA 21-43 of domain I of the rabbit polyimmunoglobulin receptor;
- 2) amino acids (AA) corresponding to AA 1 - 118 of domain I of the rabbit polyimmunoglobulin receptor;



3) amino acids (AA) corresponding to AA 119 - 223 of domain II of the rabbit polyimmunoglobulin receptor;

4) amino acids (AA) corresponding to AA 224 - 332 of domain III of the rabbit polyimmunoglobulin receptor;

5) amino acids (AA) corresponding to AA 333 - 441 of domain IV of the rabbit polyimmunoglobulin receptor;

6) amino acids (AA) corresponding to AA 442 - 552 of domain V of the rabbit polyimmunoglobulin receptor;

7) amino acids (AA) corresponding to AA of 553 to 606 or 553 to 627 of domain VI of the rabbit polyimmunoglobulin receptor; and does not contain amino acid residues corresponding to AA residues 607 to 755 or 628 to 755 of the rabbit polyimmunoglobulin receptor.

It should be noted the exact boundary of a domain may vary within approximately 20 amino acids. However, the domain structure and boundaries will be understood by one skilled in the art.

In addition, the present invention contemplates protection protein ending at the following amino acid residues of the rabbit polyimmunoglobulin receptor or at an amino acid residue which corresponds to the following residues but is in the polyimmunoglobulin receptor of another species: 580 - 605.

In other preferred embodiments, a protection protein has an amino acid sequence which corresponds to the amino acid sequence of a polyimmunoglobulin receptor for a particular species and which is analogous to the following amino acid segments:

i) amino acids (AA) corresponding to AA 21 - 43 of domain I of the rabbit polyimmunoglobulin receptor;

2) amino acids (AA) corresponding to AA 1 - 118 of domain I of the rabbit polyimmunoglobulin receptor;

3) amino acids (AA) corresponding to AA 119 - 223 of domain II of the rabbit polyimmunoglobulin receptor;

4) amino acids (AA) corresponding to AA 224 - 332 of domain III of the rabbit polyimmunoglobulin receptor;

5) amino acids (AA) corresponding to AA 333 - 441 of domain IV of the rabbit polyimmunoglobulin receptor;

6) amino acids (AA) corresponding to AA 442 - 552 of domain V of the rabbit polyimmunoglobulin receptor;

7) amino acids (AA) corresponding to AA of 553 - 606 or 553 - 627 of domain VI of the rabbit polyimmunoglobulin receptor; and does not contain amino acid residues analogous to amino acid residues 607 - 755 or 630 - 755 of the rabbit polyimmunoglobulin receptor.

In other preferred embodiments, the protection protein comprises domains I, IV, V and AA 550 - 606 or 550 - 627 of domain VI of the rabbit polyimmunoglobulin receptor or the amino acid sequence from analogous domains and regions of a polyimmunoglobulin receptor from a different species.

In other embodiments, a protection protein of the present invention has an amino acid residue sequence which substantially corresponds to at least a portion of the amino acid residues from the polyimmunoglobulin receptor of a species which are analogous to amino acid residues 1-627 of the rabbit polyimmunoglobulin receptor. This portion of the amino acid sequence would correspond to at least a portion of the extracellular domains of the receptor of that species.

In preferred embodiments, a protection protein of the present invention has an amino acid sequence which substantially corresponds to at least a portion of the amino acid residues from the polyimmunoglobulin receptor

of a species which are analogous to amino acid residues 1-606 of the rabbit polyimmunoglobulin receptor.

In other preferred embodiments, a protection protein of the present invention has an amino acid residue  
5 sequence which substantially corresponds to or is analogous to (if from a species other than rabbit) at least a portion of the following amino acid residue sequences:

- 1) amino acids (AA) corresponding to AA 21 - 43 of  
10 domain I of the rabbit polyimmunoglobulin receptor;
- 2) amino acids (AA) corresponding to AA 1 - 118 to of domain I of the rabbit polyimmunoglobulin receptor;
- 3) amino acids (AA) corresponding to AA 119 - 223 of domain II of the rabbit polyimmunoglobulin receptor;
- 15 4) amino acids (AA) corresponding to AA 224 - 332 of domain III of the rabbit polyimmunoglobulin receptor;
- 5) amino acids (AA) corresponding to AA 333 - 441 of domain IV of the rabbit polyimmunoglobulin receptor;
- 6) amino acids (AA) corresponding to AA 442 - 552  
20 of domain V of the rabbit polyimmunoglobulin receptor;
- 7) amino acids (AA) corresponding to AA of 553 - 606 or 553 - 627 of domain VI of the rabbit polyimmuno-  
globulin receptor; and does not contain amino acid  
residues corresponding to AA 628 to 755 of the rabbit  
25 polyimmunoglobulin receptor.

In other preferred embodiments, the immunoglobulins of the present invention have a protection protein which has a first amino acid sequence which substantially  
30 residues 1 to 606 or 1 to 627 of the rabbit polyimmuno-  
globulin receptor and has a second amino acid residue sequence contiguous with said first amino acid sequence, wherein said second amino acid residue sequence does not

have an amino acid residue sequence corresponding to the transmembrane segment of the rabbit polyimmunoglobulin receptor.

5 In more preferred embodiments, the second amino acid residue sequence has at least a portion of an amino acid sequence which corresponds to amino acid residues 655 to 755 of a polyimmunoglobulin receptor. In other preferred embodiments, the second amino acid residue is at least a portion of one or more of the following: an intracellular  
10 domain of a polyimmunoglobulin molecule, a domain of a member of the immunoglobulin gene superfamily, an enzyme, a toxin, or a linker.

The present invention contemplates protection proteins which do not have an amino acid residue  
15 corresponding to the transmembrane segment of rabbit polyimmunoglobulin receptor but may have amino acid residues corresponding to the intracellular domain of the rabbit polyimmunoglobulin receptor and this are deletion mutants of the receptor.

20 In other embodiments, protection proteins of the present invention have an amino acid sequence which substantially corresponds to at least one of the extracellular domains of polyimmunoglobulin receptor of a particular species. The protection protein may have an  
25 amino acid sequence of which a segment of that amino acid sequence which substantially corresponds to an extracellular domain of the polyimmunoglobulin receptor of one species, and a different segment of that amino acid sequence may be from a second species and  
30 substantially correspond to an extracellular domain from a different species. This invention contemplates embodiments in which a protection protein has an amino acid sequence which has one amino acid sequence segment

which corresponds to the amino acid sequence of the polyimmunoglobulin receptor from one species and has a second amino acid sequence within the same domain which corresponds to the amino acid sequence of the polyimmunoglobulin receptor of a different species.

Thus, the protection protein may have individual domains or portions of a particular domain that are comprised of amino acid sequences which correspond to the polyimmunoglobulin receptor from different species.

Other embodiments are contemplated in which protection protein has portions of its amino acid sequence derived from a molecule which is a member of the immunoglobulin superfamily. See, Williams and Barclay, "The Immunoglobulin Superfamily." In Immunoglobulin Genes, p. 361, Academic Press (Honjo Alt and Rabbits Eds. 1989). These derived portions may include amino acid sequences encoding peptides, domains or multiple domains from an immunoglobulin superfamily molecule.

The present invention also contemplates a nucleotide sequence encoding a protection protein which has a first nucleotide sequence encoding at least a portion of amino acids 1-606 or 1-627 of the rabbit polyimmunoglobulin receptor nucleotide sequence and which does not have a nucleotide sequence which encodes a functional trans-membrane segment 3' of the first nucleotide sequence. Further preferred embodiments include a second nucleotide sequence located 3' of the first nucleotide sequence which encodes the amino acids 1-606 or 1-627 of the rabbit polyimmunoglobulin receptor sequence. This second nucleotide sequence may encode a variety of molecules including portions of the intracellular domain of rabbit polyimmunoglobulin receptor or another polyimmunoglobulin receptor or a portion of an immunoglobulin superfamily

molecule. In addition, embodiments are contemplated in which this second nucleotide sequence encodes various effector molecules, enzymes, toxins and the like. Preferred embodiments include a second nucleotide  
5 sequence which encodes amino acid residues which correspond to amino acid residues 655 to 775 of the rabbit polyimmunoglobulin receptor or polyimmunoglobulin receptor from another species.

The present invention also contemplates expression  
10 vectors containing a nucleotide sequence encoding a protection protein which has been operatively linked to for expression. These expression vectors place the nucleotide sequence to be expressed in a particular cell 3' of a promoter sequence which causes the nucleotide  
15 sequence to be transcribed and expressed. The expression vector may also contain various enhancer sequences which improve the efficiency of this transcription. In addition, such sequences as terminators, polydenylation (poly A) sites and other 3' end processing signals may be  
20 included to enhance the amount of nucleotide sequence transcribed within a particular cell.

In preferred embodiments, the protection protein is part of an immunoglobulin that is in association with an immunoglobulin derived heavy chain having at least a  
25 portion of an antigen binding domain. Immunoglobulin derived heavy chains containing at least a portion of an antigen binding domain are well known in the art and have been described, for example, by Huse et al., Science, 246:1275 (1989), and by Lerner and Sorge, PCT Application  
30 WO 90/14430, published November 29, 1990. The disclosure of these documents are hereby incorporated by reference.

In other embodiments, the immunoglobulins of the present invention contain a protection protein and

immunoglobulin derived heavy chain and immunoglobulin derived light chain that contain at least a portion of an antigen binding site in association with the immunoglobulin derived heavy chain. Immunoglobulin light chains having at least a portion of an antigen binding domain are well known in the art and are described in available sources. See, for example, Early and Hood, Genetic Engineering, Setlow & Hollaender, (eds.), Vol. 3, Plenum Publishing Corp., New York (1981), pages 157-188; and Kabat et al., Sequences of Immunologic Interest, National Institutes of Health, Bethesda, Maryland (1987). The disclosures of all references cited herein are hereby incorporated by reference.

The immunoglobulin components of the complex (alpha, J, kappa or lambda) can contain all or part of the full length polypeptide. Parts of these chains may be used to substitute for the whole chain. For instance, the entire immunoglobulin alpha heavy chain may be replaced by the variable region and only a portion of the alpha constant region sufficient to enable assembly with the other components. Likewise, a truncated kappa or lambda chain, containing only a small section of constant region can replace the full length kappa or lambda chains. The prerequisite of any complex is the ability to bind the protection protein.

In addition to truncated components, the present invention contemplates the combination of different types of immunoglobulins. For example, a heavy chain constant region comprising the C<sub>H</sub>1 and C<sub>H</sub>2 regions of IgG followed by the C<sub>H</sub>2 and C<sub>H</sub>3 regions derived from an IgA will form a stable complex containing the protection protein. This is specifically described as an example.

The immunoglobulins containing the protection proteins of the present invention preferably contain at least a portion of an IgM or IgA heavy chain which allows that immunoglobulin heavy chain to bind to immunoglobulin J chain and thereby bind to the protection protein. It is contemplated that the immunoglobulin heavy chain of the present invention may be comprised of individual domains selected from the IgA heavy chain or the IgM heavy chain or from some other isotype of heavy chain. It is also contemplated that an immunoglobulin domain derived from an immunoglobulin heavy chain other than IgA or IgM may be molecularly engineered to bind immunoglobulin J chain and thus may be used to produce immunoglobulins of the present invention.

One skilled in the art will understand that immunoglobulins consist of domains which are approximately 100-110 amino acid residues. These various domains are well known in the art and have known boundaries. The removal of a single domain and its replacement with a domain of another antibody molecule is easily achieved with modern molecular biology. The domains are globular structures which are stabilized by intrachain disulfide bonds. This confers a discrete shape and makes the domains a self-contained unit that can be replaced or interchanged with other similarly shaped domains. The heavy chain constant region domains of the immunoglobulins confer various properties known as antibody effector functions on a particular molecule containing that domain. Example effector functions include complement fixation, placental transfer, binding to staphylococcal protein, binding to streptococcal protein G, binding to mononuclear cells, neutrophils or mast cells and basophils. The association of particular domains and particular immunoglobulins



isotopes with these effector functions is well known and for example, described in Immunology, Roitt et al., Mosby St. Louis, Missouri (1993 3rd Ed.)

The immunoglobulins of the present invention may, in addition to the protection protein, contain immuno-  
5 globulin heavy chains, immunoglobulin light chains, or immunoglobulin J chain bound to the immunoglobulin derived heavy chains. In preferred embodiments, the immunoglobulin of the present invention comprises two or  
10 four immunoglobulin derived heavy chains, together with two or four immunoglobulin light chains and an immunoglobulin J chain bound to at least one of the immunoglobulin derived heavy chains. The immunoglobulin J chain is described and known in the art. See, for  
15 example, M. Koshland, The Immunoglobulin Helper: The J Chain, in Immunoglobulin Genes, Academic Press, London, Pg. 345, (1989) and Matsuuchi et al., Proc. Natl. Acad. Sci. U.S.A., 83:456-460 (1986). The sequence of the immunoglobulin J chain is available on various data bases  
20 in the United States.

The immunoglobulin of the present invention has a protection protein associated with at least an immunoglobulin derived heavy chain. This association may occur by hydrogen bonds, disulfide bonds, covalent bonds,  
25 ionic interactions or combinations of these various bonds. Typically, immunoglobulin molecules are held together by disulfide bonds between the immunoglobulin heavy chains and immunoglobulin light chains. The interaction of the protection protein with the  
30 immunoglobulin is by non-covalent or disulfide bonding.

The immunoglobulins of the present invention containing the protection protein, the immunoglobulin derived heavy chain and optionally an immunoglobulin

derived light chain, and J chain are typically bonded together by one of the following: hydrogen bonds, disulfide bonds, covalent bonds, ionic interactions or combinations of these bonds. The present invention  
5 contemplates molecules in which the required portions of the immunoglobulin heavy, light and/or J chain have been placed into a single polypeptide and function to bind antigen and protection protein. Examples of such proteins are single-chain antigen-binding proteins.

10 The present invention contemplates a method of assembling a multimeric immunoglobulin comprising the steps of: introducing into an organism a DNA segment encoding all or part of an immunoglobulin J chain, and a DNA segment encoding all or part of an immunoglobulin  
15 alpha chain, and a DNA segment encoding all or part of either an immunoglobulin kappa chain or an immunoglobulin lambda chain; and introducing into the same organism a protection protein, said protection protein comprising at least a segment of the amino acid residues 1 to residue  
20 606 of the rabbit polyimmunoglobulin receptor (pIgR) amino acid residue sequence or analogous amino acid residues from other species such that the segment is derived from a precursor protein that does not contain the amino acid residues comprising a functional membrane  
25 spanning region nor is the segment derived from a precursor protein in which the sequence of amino acid residues from the beginning of the membrane spanning region (approximately residue 630 of rabbit polyimmuno-  
globulin receptor) to the carboxyl end of the protein  
30 (approximately residue 755 of the rabbit polyimmuno-  
globulin receptor) are fully intact. In preferred embodiments the precursor protein does not contain amino acid residues greater than 606 of the rabbit

polyimmunoglobulin receptor or analogous amino acid residues from other species.

As is understood by those of ordinary skill in the art, a membrane spanning region or functional transmembrane segment consists of a contiguous section of amino acid residues containing from about 20 to about 30 amino acids in which none of the residues is charged, virtually all of the residues are hydrophobic or non-polar, and the segment forms an alpha helix. A functional transmembrane segment is capable of spanning a biomembrane. Membrane spanning regions can be bounded by charged residues. An example of a membrane spanning region of pIgR is residues 630 to 653 of the polyimmunoglobulin receptor amino acid residue sequence of rabbit.

The chains that comprise the immunoglobulin containing the protection protein may be derived from precursors containing a signal sequence at the amino terminal of the protein. Each component can thereby be synthesized into an endomembrane system where assembly occurs. In addition to a signal sequence, the various components of the complex may or may not contain additional signals for N terminal glycosylation or for various other modifications which can affect the structure of the complex. In one embodiment of the invention, the signals for glycosylation (i.e. asparagine-X-serine or threonine or the signals for O-linked glycosylation) are not present or present in more or less places within the nucleotide sequence. The resulting antibody therefore would contain no carbohydrate, which may be advantageous for applications in which carbohydrates elicit an immune response.

In preferred embodiments, the immunoglobulin of the present invention contains a protection protein

associated with an immunoglobulin derived heavy chain and the protection protein is free from N-linked and/or O-linked oligosaccharides. One skilled in the art will understand that a gene coding for a polypeptide having within its amino acid residue sequence the N-linked glycosylation signal asparagine-X-serine/threonine where X can be any amino acid residue except possibly proline and aspartic acid, when introduced into a plant cell would be glycosylated via oligosaccharides linked to the asparagine residue of the sequence (N-linked). See, Marshall, Ann. Rev. Biochem., 41:673 (1972) and Marshall, Biochem. Soc. Symp., 40:17 (1974) for a general review of the polypeptide sequences that function as glycosylation signals. These signals are recognized in both mammalian and in plant cells. One skilled in the art will understand that the N-linked glycosylation signal may be easily removed using common mutagenesis procedures to change the DNA sequence encoding the protection protein of the present invention. This mutagenesis typically involves the synthesis of oligonucleotide having the N-linked glycosylation signal deleted and then preparing a DNA strand with that oligonucleotide sequence incorporated into it. Such mutagenesis procedures and reagents are commercially available from many sources such as Stratagene (La Jolla, CA.).

Assembly of the individual polypeptides that form a multi-peptide molecule (for example immunoglobulin) may be obtained by expressing in a single cell by directly introducing all the transgenes encoding the individual polypeptides into that cell either sequentially or all at once. The transgenes encoding the polypeptides may be present on individual constructs or DNA segments or may

be contained in a DNA segment or construct together with one or more other transgenes.

Assembly of these components can be by cross pollination as originally described by Mendel to produce  
5 a population of segregants expressing all chains. Previous disclosures have demonstrated this to be an adequate method for the assembly and co-segregation of multimeric glycoconjugates. The disclosure of U.S. Patent No. 5,202,422 is hereby incorporated by reference  
10 and describes these methods. In a preferred embodiment of the present invention, the antibody molecules contain a reduced number of glycans and antibody molecules with no glycans are contemplated.

The immunoglobulins of the present invention  
15 containing the protection protein, the immunoglobulin derived heavy chain and optionally an immunoglobulin derived light chain, and J chain may contain a protection protein that is free from N-linked oligosaccharides.

The immunoglobulins of the present invention that  
20 contain the protection protein are preferably therapeutic immunoglobulins that are useful in preventing a disease in an animal. In preferred embodiments, the immunoglobulins of the present invention are therapeutic immunoglobulins which are capable of binding to mucosal  
25 pathogen antigens. In other preferred embodiments, the therapeutic immunoglobulins of the present invention are capable of preventing dental caries. In the most preferred embodiment, the immunoglobulin of the present invention containing the protection protein contains an  
30 antigen binding domain that is capable of binding to an antigen from S. mutans serotypes a, c, d, e, f, g and h (*S. mutans* c, e and f and *S. sobrinus* serotypes d and g under new nomenclature). Such antigen binding domains

are known in the art and include, for example, the binding domains described in U.S. Patent 5,352,446, J. K-C. Ma et al., Clin. Exp. Immunol. 77:331 (1989); and J. K-C. Ma et al., Eur. J. Immunol. 24:131-138 (1994); U.S. Patent 5,352,446; U.S. Patent 4,594,244; and European Patent Publication 371 017 B1. The disclosures of these documents are hereby incorporated by reference. In preferred embodiments, the immunoglobulins of the present invention are part of a composition that has a therapeutic activity on either animals or humans. Examples of therapeutic immunoglobulins are numerous, however, we envision the most appropriate therapeutic effect to be prophylaxis for mucosal and enteric pathogens by direct oral administration of the composition derived from an edible plant.

Administration of the therapeutic composition can be before or after extraction from the plant or other transgenic organism. Once extracted the immunoglobulins may also be further purified by conventional techniques such as size exclusion, ion exchange, or affinity chromatography. In the preferred embodiment, the transgenic organism is an edible plant and administration of the complex is by ingestion after partial purification. Plant molecules may be co-administered with the complex.

The present invention also contemplates that the relative proportion of plant-derived molecules and animal-derived molecules can vary. Quantities of specific plant proteins, such as RuBisCo, or chlorophyll may be as little as 1% of the mass or as much as 99.9% of the mass of the extract, excluding water.

The present invention also contemplates the use of the therapeutic plant extract containing immunoglobulins having a protection protein directly without any further

purification of the specific therapeutic component, e.g. the antibody. Administration may be by topical application, oral ingestion or any other method appropriate for delivering the antibody to the mucosal target pathogen. This form of administration is distinct from parenteral applications involving direct injection or commingling of the therapeutic plant extract with the blood stream.

The present invention also contemplates the use of the therapeutic plant extract containing immunoglobulins having a protection protein after manipulating the taste or texture of the extract. Appropriate quantities of gelling substances or flavorings could be added to enhance the contact of the antibody with the target pathogen in, for example, direct oral applications.

In preferred embodiments, the immunoglobulins of the present invention are used to passively immunize an animal against a preselected ligand by contacting a composition comprising an immunoglobulin containing a protection protein of the present invention that is capable of binding a preselected ligand with a mucosal surface of an animal. Passive immunization requires large amounts of antibody and for wide-spread use this antibody must be inexpensive.

Immunoglobulin molecules containing protection proteins that are capable of binding a preselected antigen can be efficiently and economically produced in plant cells. In preferred embodiments, the immunoglobulin molecule is either IgA, IgM, secretory IgM or secretory IgA or an immunoglobulin having a chimeric immunoglobulin heavy or light chain.

The immunoglobulins containing protection proteins are more resistant to proteolysis and denaturation and

therefore are desirable for use in harsh environments. Contemplated harsh environments include acidic environments, protease containing environments, high temperature environments, and other harsh environments.

5 For example, the gastrointestinal tract of an animal is a harsh environment where both proteases and acid are present. See, Kobayashi et al., Immunochimistry, 10:73 (1973).

Passive immunization of the animal using these more  
10 resistant immunoglobulins of the present invention is produced by contacting the immunoglobulin containing the protection protein with a mucosal surface of the animal. Animals have various mucosal surfaces including the lungs, the digestive tract, the nasopharyngeal cavity,  
15 the urogenital system, and the like. Typically, these mucosal surfaces contain cells that produce various secretions including saliva, lacrimal fluid, nasal fluid, tracheobronchial fluid, intestinal fluid, bile, cervical fluid, and the like.

20 In preferred embodiments the immunoglobulins that contain the protection protein are immunospecific for a preselected antigen. Typically, this antigen is present on a pathogen that causes a disease that is associated with the mucosal surface such as necrotizing  
25 enterocolitis, diarrheal disease, ulcers, and cancer caused by carcinogen absorption in the intestine. See e.g., McNabb and Tomasi, Ann. Revl. Microbiol., 35:477 (1981) and Lawrence et al., Science, 243:1462 (1989). Typical pathogens that cause diseases associated with a  
30 mucosal surface include both bacterial and viral pathogens, such as E. coli., S. typhimurium, V. cholera, H. pylori, and S. mutans. See also, European Patent Application 484, 148 A1, published 5/6/92 and hereby



incorporated by reference. The immunoglobulins of the present invention are capable of binding to these pathogens and preventing them from causing mucosal associated diseases.

5        Immunoglobulins capable of binding to S. mutans and preventing dental caries have been described in European Patent Specification 371,017 which is hereby incorporated by reference. The disclosure of U.S. Patent No. 5,352,440 is also hereby incorporated by reference.

10        Therapeutic immunoglobulins of the present invention that contain protection proteins that would be effective against bacterial infection or carcinomas are contemplated. Monoclonal antibodies with therapeutic activity have been described in U.S. Patents 4,652,448,  
15        4,443,549 and 5,183,756 which are hereby incorporated by reference.

          In preferred embodiments, the immunoglobulin of the invention are part of a composition which is contacted with the animal mucosal surface comprises plant material  
20        and an immunoglobulin of the present invention that is capable of binding a preselected ligand. The plant material present may be plant cell walls, plant organelles, plant cytoplasm, intact plant cells, viable plants, and the like. This plant cell material is  
25        present in a ratio from about 10,000 grams of plant material to about 100 nanograms of immunoglobulin to about 100 nanograms of plant material for each 10 grams of immunoglobulin present. In more preferred  
          embodiments, the plant material is present in a ratio  
30        from about 10,000 grams of plant material for each 1 gram of immunoglobulin present to about a ratio of 100 nanograms of plant material present for each gram of immunoglobulin present. In other preferred embodiments,

the plant material is present in a ratio from about 10,000 grams of plant material for each milligram of immunoglobulin present to about 1 milligram of plant material present for each 500 milligram of immunoglobulin present.

5 In preferred embodiments, the composition containing the immunoglobulins of the present invention is a therapeutic composition. The preparation of therapeutic compositions which contain polypeptides or proteins as active ingredients is well understood in the art. Therapeutic compositions may be liquid solutions or suspensions, solid forms suitable for solution in, or suspension in a liquid prior to ingestion may also be prepared. The therapeutic may also be emulsified. The active therapeutic ingredient is typically mixed with inorganic and/or organic carriers which are pharmaceutically acceptable and compatible with the active ingredient. The carriers are typically physiologically acceptable excipients comprising more or less inert substances when added to the therapeutic composition to confer suitable consistencies and form to the composition. Suitable carriers are for example, water, saline, dextrose, glycerol, and the like and combinations thereof. In addition, if desired the composition can contain minor amounts of auxiliary substances such as wetting or emulsifying agents and pH buffering agents which enhance the effectiveness of the active ingredient. Therapeutic compositions containing carriers that have nutritional value are also contemplated.

30 In embodiments in which a composition containing an immunoglobulin having a protection protein of the present invention is applied to the tooth or mouth of a mammal, any convenient method may be used. Methods for applying

such a composition to the teeth are well known and utilize various materials for a variety of purposes. For example, the composition may be directly applied to the tooth by painting the surface of the tooth with that composition. Alternatively, the composition of the present invention may be included in a toothpaste, mouthwash, chewing gum, lozenge or gel that will result in it being applied to the teeth. In some formulations, it may be desirable to provide for a formulation that prolongs the contact of the composition and therefore the immunoglobulin having the protection protein with the tooth surface. Formulations for this purpose are well known and include such formulations that may be placed in various dental trays that are used to cover the tooth and other dental apparatuses that are used in adjusting various conditions with the teeth.

The exact amount of a composition that must be applied to the teeth during any particular application is not critical because such treatment may be easily repeated at a given interval. For example, compositions present in toothpaste would be applied to the teeth each time that toothpaste is used, typically twice per day. For example, the order of 10 to 100 micrograms of an immunoglobulin having a protection protein can be applied to each tooth on each occasion the composition is applied to the teeth. However, this in no way should be taken as a limitation on a range that may be applied during any particular application as applications of a composition having more or less immunoglobulin of the present invention may be used without detrimental effect. The use of much lower concentrations of an immunoglobulin of the present invention would result in, at some point, a reduction in the protection provided by such formulation.

The exact formulation for the composition of the present invention may vary and will depend on the method of application to be used and the frequency of that application. In general, it may be any formulation which has an appropriate pH and which is free of material which would render the immunoglobulin having the protection protein of the present invention ineffective. For example, the compositions of the present invention may be applied as a simple aqueous solution in which the composition is disbursed at anywhere from 0.1 to 10 milligrams of immunoglobulin per 100 microliters of that solution. Generally, such a solution would be applied during dental surgery at a rate of approximately 1 to 10 microliters of the solution per tooth.

The formulations of the compositions of the present invention which are designed to be self-administered may vary and will be formulated taking in to account the frequency of application of the particular product in which is it used.

In preferred embodiments, a composition containing an immunoglobulin of the present invention comprises an immunoglobulin molecule that is immunospecific for a pathogen antigen. Pathogens are any organism that causes a disease in another organism. Particularly preferred are immunoglobulins that are immunospecific for a mucosal pathogen antigen. A mucosal pathogen antigen is present on a pathogen that invades an organism through mucosal tissue or causes mucosal associated diseases. Mucosal pathogens include lung pathogens, nasal pathogens, intestinal pathogens, oral pathogens, and the like. For a general discussion of pathogens, including mucosal pathogens, see, Davis et al., Microbiology, 3rd ed., Harper and Row, Hagerstown, MD (1980).

Antibodies immunospecific for a pathogen may be produced using standard monoclonal antibody production techniques. See, Antibodies: A Laboratory Manual, Harlow et al., eds., Cold Spring Harbor, NY (1988). The genes  
5 coding for the light chain and heavy chain variable regions can then be isolated using the polymerase chain reaction and appropriately selected primers. See, Orlandi et al., Proc. Natl. Acad. Sci., U.S.A., 86:3833 (1989) and Huse et al., Science, 246:1275 (1989). The  
10 variable regions are then inserted into plant expression vectors, such as the expression vectors described by Hiatt et al., Nature, 342:76-78 (1989).

In a preferred embodiment, the immunoglobulin of the present invention is immunospecific for an intestinal  
15 pathogen antigen. Particularly preferred are immunoglobulins immunospecific for intestinal pathogens such as bacteria, viruses, and parasites that cause disease in the gastrointestinal tract, such as E. coli, Salmonellae, Vibrio cholerae, Salmonellae typhimurium,  
20 Shigella and H. pylori.

In other preferred embodiments, the immunoglobulin containing the protection protein present in the composition is an immunoglobulin molecule that is immuno-  
specific for a dental pathogen such as Streptococcus  
25 mutans and the like. Particularly preferred are immunoglobulins immunospecific for a Streptococcus mutans antigen such as the immunoglobulin produced by hybridoma 15B2 (ATCC No. HB 8510); the hybridoma deposited as European Collection of Animal cells Deposit No. 86031901;  
30 and the Guy's 13 monoclonal antibody described by Ma et al., Eur. J. Immunol., 24:131 (1994) and Smith and Lehner, Oral Micro. Immunol., 4:153 (1989).

The present invention contemplates producing passive immunity in an animal, such as vertebrate. In preferred embodiments, passive immunity is produced in fish, birds, reptiles, amphibians, or insects. In other preferred  
5   embodiments passive is produced in an mammal, such as a human, a domestic animal, such as a ruminant, a cow, a pig, a horse, a dog, a cat, and the like. In particularly preferred embodiments, passive immunity is produced in an adult or child mammal.

10       In preferred embodiments, passive immunity is produced in an animal, such as a mammal that is weaned and therefore no longer nurses to obtain milk from its mother. Passive immunity is produced in such an animal by administering to the animal a sufficient amount of  
15   composition containing an immunoglobulin containing a protection protein immunospecific for a preselected ligand to produce a prophylactic concentration of the immunoglobulin within the animal. A prophylactic concentration of an immunoglobulin is an amount  
20   sufficient to bind to a pathogen present and prevent that pathogen from causing detectable disease within the animal. The amount of composition containing the immunoglobulin of the present invention required to produce a prophylactic concentrations will vary as is  
25   well known in the art with the size of the animal, the amount of pathogen present, the affinity of the particular immunoglobulin for the pathogen, the efficiency with which the particular immunoglobulin is delivered to its active location within the animal, and  
30   the like.

C. EUKARYOTIC CELLS CONTAINING IMMUNOGLOBULINS HAVING A PROTECTION PROTEIN

The present invention contemplates eukaryotic cells,  
5 including plant cells, containing immunoglobulins of the present invention. The present invention also contemplates plant cells that contain nucleotide sequences encoding the various components of the immunoglobulins of the present invention. One skilled in  
10 the art will understand that the nucleotide sequences that encode the protection protein and the various immunoglobulin heavy and light chains and J chain will typically be operably linked to a promoter and present as part of an expression vector or cassette.

15 After the immunoglobulin heavy and light chain genes, and J chain genes are isolated, they are typically operatively linked to a transcriptional promoter in an expression vector.

Expression of the components in the organism of  
20 choice can be derived from an independently replicating plasmid, or from a permanent component of the chromosome, or from any piece of DNA which may transiently give rise to transcripts encoding the components. Organisms suitable for transformation can be either prokaryotic or  
25 eukaryotic. Introduction of the components of the complex can be by direct DNA transformation, by ballistic delivery into the organism, or mediated by another organism as for example by the action of recombinant *Agrobacteria* on plant cells. Expression of proteins in  
30 transgenic organisms usually requires co-introduction of an appropriate promoter element and polyadenylation signal. In one embodiment of the invention, the promoter element potentially results in the constitutive expression of the components in all of the cells of a

plant. Constitutive expression occurring in most or all of the cells will ensure that precursors can occupy the same cellular endomembrane system as might be required for assembly to occur.

5        Expression vectors compatible with the host cells, preferably those compatible with plant cells are used to express the genes of the present invention. Typical expression vectors useful for expression of genes in plants are well known in the art and include vectors  
10        derived from the tumor-inducing (Ti) plasmid of Agrobacterium tumefaciens described by Rogers et al., Meth. in Enzymol., 153:253-277 (1987). However, several other expression vector systems are known to function in plants. See for example, Verma et al., PCT Publication  
15        No. WO87/00551; and Cocking and Davey, Science, 236:1259-1262 (1987).

         The expression vectors described above contain expression control elements including the promoter. The genes to be expressed are operatively linked to the  
20        expression vector to allow the promoter sequence to direct RNA polymerase binding and synthesis of the desired polypeptide coding gene. Useful in expressing the genes are promoters which are inducible, viral, synthetic, constitutive, and regulated. The choice of  
25        which expression vector and ultimately to which promoter a nucleotide sequence encoding part of the immunoglobulin of the present invention is operatively linked depends directly, as is well known in the art, on the functional properties desired, e.g. the location and timing of  
30        protein expression, and the host cell to be transformed, these being limitations inherent in the art of constructing recombinant DNA molecules. However, an expression vector useful in practicing the present



invention is at least capable of directing the replication, and preferably also the expression of the polypeptide coding gene included in the DNA segment to which it is operatively linked.

5 In preferred embodiments, the expression vector used to express the genes includes a selection marker that is effective in a plant cell, preferably a drug resistance selection marker. A preferred drug resistance marker is the gene whose expression results in kanamycin  
10 resistance, i.e., the chimeric gene containing the nopaline synthase promoter, Tn5 neomycin phosphotransferase II and nopaline synthase 3' nontranslated region described by Rogers et al., in Methods For Plant Molecular Biology, a Weissbach and H. Weissbach, eds.,  
15 Academic Press Inc., San Diego, CA (1988). A useful plant expression vector is commercially available from Pharmacia, Piscataway, NJ.

Expression vectors and promoters for expressing foreign proteins in plants have been described in U.S.  
20 Patent Nos. 5,188,642; 5,349,124; 5,352,605, and 5,034,322 which are hereby incorporated by reference.

A variety of methods have been developed to operatively link DNA to vectors via complementary cohesive termini. For instance, complementary  
25 homopolymer tracks can be added to the DNA segment to be inserted and to the vector DNA. The vector and DNA segment are then joined by hydrogen bonding between the complementary homopolymeric tails to form recombinant DNA molecules.

30 Alternatively, synthetic linkers containing one or more restriction endonuclease sites can be used to join the DNA segment to the expression vector. The synthetic linkers are attached to blunt-ended DNA segments by

incubating the blunt-ended DNA segments with a large excess of synthetic linker molecules in the presence of an enzyme that is able to catalyze the ligation of blunt-ended DNA molecules, such as bacteria phage T4 DNA  
5 ligase. Thus, the products of the reaction are DNA segments carrying synthetic linker sequences at their ends. These DNA segments are then cleaved with the appropriate restriction endonuclease and ligated into an expression vector that has been cleaved with an enzyme  
10 that produces termini compatible with those of the synthetic linker. Synthetic linkers containing a variety of restriction endonuclease sites are commercially available from a number of sources including New England BioLabs, Beverly, MA.

15 The nucleotide sequences encoding the protection protein and any other of the immunoglobulins of the present invention are introduced into the same plant cell either directly or by introducing each of the components into a plant cell and regenerating a plant and cross-  
20 hybridizing the various components to produce the final plant cell containing all the required components.

Any method may be used to introduce the nucleotide sequences encoding the components of the immunoglobulins of the present invention into a eukaryotic cell.. For  
25 example, methods for introducing genes into plants include Agrobacterium-mediated plant transformation, protoplast transformation, gene transfer into pollen, injection into reproductive organs and injection into immature embryos. Each of these methods has distinct  
30 advantages and disadvantages. Thus, one particular method of introducing genes into a particular eukaryotic cell or plant species may not necessarily be the most effective for another eukaryotic cell or plant species.

Agrobacterium tumefaciens-mediated transfer is a widely applicable system for introducing genes into plant cells because the DNA can be introduced into whole plant tissues, bypassing the need for regeneration of an intact  
5 plant from a protoplast. The use of Agrobacterium-mediated expression vectors to introduce DNA into plant cells is well known in the art. See, for example, the methods described by Fraley et al., Biotechnology, 3:629 (1985) and Rogers et al., Methods in Enzymology, 153:253-  
10 277 (1987). Further, the integration of the Ti-DNA is a relatively precise process resulting in few rearrangements. The region of DNA to be transferred is defined by the border sequences and intervening DNA is usually inserted into the plant genome as described by  
15 Spielmann et al., Mol. Gen. Genet., 205:34 (1986) and Jorgensen et al., Mol. Gen. Genet., 207:471 (1987). Modern Agrobacterium transformation vectors are capable of replication in Escherichia coli as well as Agrobacterium, allowing for convenient manipulations as  
20 described by Klee et al., in Plant DNA Infectious Agents, T. Hohn and J. Schell, eds., Springer-Verlag, New York (1985) pp. 179-203. Further recent technological advances in vectors for Agrobacterium-mediated gene transfer have improved the arrangement of genes and  
25 restriction sites in the vectors to facilitate construction of vectors capable of expressing various polypeptide coding genes. The vectors described by Rogers et al., Methods in Enzymology, 153:253 (1987), have convenient multi-linker regions flanked by a  
30 promoter and a polyadenylation site for direct expression of inserted polypeptide coding genes and are suitable for present purposes.

Agrobacterium-mediated transformation of leaf disks and other tissues appears to be limited to plant species that Agrobacterium tumefaciens naturally infects. Thus, Agrobacterium-mediated transformation is most efficient  
5 in dicotyledonous plants. However, the transformation of Asparagus using Agrobacterium can also be achieved. See, for example, Bytebier, et al., Proc. Natl. Acad. Sci., 84:5345 (1987).

In those plant species where Agrobacterium-mediated  
10 transformation is efficient, it is the method of choice because of the facile and defined nature of the gene transfer. However, few monocots appear to be natural hosts for Agrobacterium, although transgenic plants have been produced in asparagus using Agrobacterium vectors as  
15 described by Bytebier et al., Proc. Natl. Acad. Sci. U.S.A., 84:5345 (1987). Therefore, commercially important cereal grains such as rice, corn, and wheat must be transformed using alternative methods. Transformation of plant protoplasts can be achieved using  
20 methods based on calcium phosphate precipitation, polyethylene glycol treatment, electroporation, and combinations of these treatments. See, for example, Potrykus et al., Mol. Gen. Genet., 199:183 (1985); Lorz et al., Mol. Gen. Genet., 199:178 (1985); Fromm et al.,  
25 Nature, 319:791 (1986); Uchimiya et al., Mol. Gen. Genet., 204:204 (1986); Callis et al., Genes and Development, 1:1183 (1987); and Marcotte et al., Nature, 335:454 (1988).

Application of these systems to different plant  
30 species depends upon the ability to regenerate that particular plant species from protoplasts. Illustrative methods for the regeneration of cereals from protoplasts are described in Fujimura et al., Plant Tissue Culture

Letters, 2:74 (1985); Toriyama et al., Theor Appl. Genet., 73:16 (1986); Yamada et al., Plant Cell Rep., 4:85 (1986); Abdullah et al., Biotechnology, 4:1087 (1986).

5       To transform plant species that cannot be successfully regenerated from protoplast, other ways to introduce DNA into intact cells or tissues can be utilized. For example, regeneration of cereals from immature embryos or explants can be effected as described  
10 by Vasil, Biotechnology, 6:397 (1988). In addition, "particle gun" or high-velocity microprojectile technology can be utilized as well. Using such technology, DNA is carried through the cell wall and into the cytoplasm on the surface of small (0.525 um) metal  
15 particles that have been accelerated to speeds of one to several hundred meters per second as described in Klein et al., Nature, 327:70 (1987); Klein et al., Proc. Natl. Acad. Sci. U.S.A., 85:8502 (1988); and McCabe et al., Biotechnology, 6:923 (1988). The metal particles  
20 penetrate through several layers of cells and thus allow the transformation of cells within tissue explants. Metal particles have been used to successfully transform corn cells and to produce fertile, stably transformed tobacco and soybean plants. Transformation of tissue  
25 explants eliminates the need for passage through a protoplast stage and thus speeds the production of transgenic plants.

DNA can be introduced into plants also by direct DNA transfer into pollen as described by Zhou et al., Methods  
30 in Enzymology, 101:433 (1983); D. Hess, Intern Rev. Cytol., 107:367 (1987); Luo et al., Plant Mol. Biol. Reporter, 6:165 (1988). Expression of polypeptide coding genes can be obtained by injection of the DNA into

reproductive organs of a plant as described by Pena et al., Nature, 325:274 (1987). DNA can also be injected directly into the cells of immature embryos and the rehydration of desiccated embryos as described by Neuhaus et al., Theor. Apl. Genet., 75:30 (1987); and Benbrook et al., in Proceedings Bio Expo 1986, Butterworth, Stoneham, MA, pp. 27-54 (1986).

The regeneration of plants from either single plant protoplasts or various explants is well known in the art. See, for example, Methods for Plant Molecular Biology, A. Weissbach and H. Weissbach, eds., Academic Press, Inc., San Diego, CA (1988). This regeneration and growth process includes the steps of selection of transformant cells and shoots, rooting the transformant shoots and growth of the plantlets in soil.

The regeneration of plants containing the foreign gene introduced by Agrobacterium tumefaciens from leaf explants can be achieved as described by Horsch et al., Science, 227:1229-1231 (1985). In this procedure, transformants are grown in the presence of a selection agent and in a medium that induces the regeneration of shoots in the plant species being transformed as described by Fraley et al., Proc. Natl. Acad. Sci. U.S.A., 80:4803 (1983). This procedure typically produces shoots within two to four weeks and these transformant shoots are then transferred to an appropriate root-inducing medium containing the selective agent and an antibiotic to prevent bacterial growth. Transformant shoots that rooted in the presence of the selective agent to form plantlets are then transplanted to soil to allow the production of roots. These procedures will vary depending upon the particular plant

species employed, such variations being well known in the art.

The immunoglobulins of the present invention may be produced in any plant cell including plant cells derived  
5 from plants that are dicotyledonous or monocotyledonous, solanaceous, alfalfa, legumes, or tobacco.

Transgenic plants of the present invention can be produced from any sexually crossable plant species that can be transformed using any method known to those  
10 skilled in the art. Useful plant species are dicotyledons including tobacco, tomato, the legumes, alfalfa, oaks, and maples; monocotyledons including grasses, corn, grains, oats, wheat, and barley; and lower plants including gymnosperms, conifers, horsetails, club  
15 mosses, liver warts, horn warts, mosses, algae, gametophytes, sporophytes of pteridophytes.

The plant cells of the present invention may in addition to the protection protein and the immunoglobulin derived heavy chain also contains a nucleotide sequence  
20 encoding an immunoglobulin derived light chain having at least a portion of an antigen binding domain.

The plant cells of the present invention may have an antigen binding domain that is capable of binding an antigen from S. mutans serotypes a, c, d, e, f, g, and h  
25 (*S. mutans* serotypes c, e, and f; and *S. sobrinus* serotypes d and g under new nomenclature) on the immunoglobulin derived heavy and light chains. The antigen binding domain present in these plant cells also can be able to bind to the responsible mucosal pathogens and  
30 prevent dental caries.

The plant cells of the present invention may be part of a plant and make up one of the following types of

plants: dicotyledonous, monocotyledonous, solanaceous,  
alfalfa, tobacco or other type of plant.

5 D. COMPOSITIONS CONTAINING IMMUNOGLOBULINS HAVING  
PROTECTION PROTEINS.

The present invention contemplates compositions of  
matter that comprise immunoglobulins of the present  
invention and plant macromolecules. Typically these  
10 plant macromolecules are derived from any plant useful in  
the present invention. The plant macromolecules are  
present together with an immunoglobulin of the present  
invention for example, in a plant cell, in an extract of  
a plant cell, or in a plant. Typical plant macro-  
15 molecules associated with the immunoglobulins of the  
present invention in a composition are ribulose  
biphosphate carboxylase, light harvesting complex, (LH6)  
pigments, secondary metabolites or chlorophyll. The  
compositions of the present invention have an  
20 immunoglobulin of the present invention present in a  
concentration of between 1% and 99% mass excluding water.  
Other preferred compositions include compositions having  
the immunoglobulins of the present invention present at a  
concentration of between 1% and 50% mass excluding water.  
25 Other preferred compositions include immunoglobulins at a  
concentration of 1% to 25% mass excluding water.

The compositions of the present invention contain  
plant macromolecules at a concentration of between 1% and  
99% mass excluding water.. Typically the mass present in  
30 the composition will consist of plant macromolecules and  
immunoglobulins of the present invention. When the  
immunoglobulins of the present invention are present at a  
higher or lower concentration the concentration of plant  
macromolecules present in the composition will vary



inversely. In preferred embodiments the composition of plant macromolecules are present in a concentration of between 50% and 99% mass excluding water. In the most preferred compositions, the plant macromolecules are  
5 present in a concentration of between 75% and 99% mass excluding water.

The present invention contemplates a composition of matter comprising all or part of the following: an IgA heavy chain, a kappa or lambda chain, a J chain. These  
10 components form a complex and are attached to the protection protein as defined earlier. The composition also contains molecules derived from a plant. This composition may also be obtained after an extraction process yielding functional antibody and plant-derived  
15 molecules.

The extraction method comprises the steps of applying a force to a plant containing the complex whereby the apoplastic compartment of the plant is ruptured releasing said complex. The force involves  
20 shear, in dyn/cm<sup>2</sup>, as the primary method of releasing the apoplastic liquid.

The whole plant or plant extract contains an admixture of antibody and various other macromolecules of the plant. Among the macromolecules contained in the  
25 admixture is ribulose biphosphate carboxylase (RuBisCo) or fragments of RuBisCo. Another macromolecule is LHCP. Another molecule is chlorophyll.

Shear force is a useful component of the overall force applied to the plant for disruption of apoplastic  
30 spaces. Other types of force may also be included to optimize the effects of shear. Direct pressure, for example, measured in lbs/in<sup>2</sup>, may enhance the effects of the apparatus used to apply shear. Commonly used

homogenization techniques which are not appropriate for antibody extraction involve the use of high speed blades or cylinders which explosively destroy all plant structures.

5       The compositions of the present invention may contain an immunoglobulin of the present invention and plant molecules that are derived from a dicotyledonous, monocotyledonous, solanaceous, alfalfa, tobacco or other plant. The plant molecules present in the compositions  
10 of the present invention can be ribulose biphosphate carboxylase, light harvesting complex, pigments, secondary metabolites, chlorophyll or other plant molecules.

Other useful methods for preparing composition  
15 containing immunoglobulins having protection protein include extraction with various solvents and application of vacuum to the plant material. The compositions of the present invention may contain immunoglobulins of the present in a concentration of between 1% and 99% mass  
20 excluding water. The compositions of the present invention may contain plant macromolecules in a concentration of between 1% and 99% mass excluding water.

Therapeutic compositions containing immunoglobulins of the present invention and plant macromolecules may be  
25 produced by processing a plant of the present invention by shearing under pressure a portion of that plant to produce a pulp containing the therapeutic immunoglobulin and plant macromolecules in a liquid derived from the apoplast or symplast of the plant which also contains the  
30 solid plant derived material. Further processing may be accomplished by separating the solid plant derived material from the plant derived liquid containing the immunoglobulins of the present invention. The starting

material for such a process may include plant leaves, stem, roots, tubers, seeds, fruit or the entire plant. Typically, this processing is accomplished by a mechanical device which releases liquid from the apoplast  
5 or symplast of the plant. Additional processing steps may include separation of the solid plant derived material from the liquid using centrifugation settling flocculation or filtration. One skilled in the art will understand that these separation methods result in  
10 removing the solid plant derived material from the liquid including the immunoglobulins of the present invention. The methods of the present invention may produce immunoglobulins containing a protection protein and an immunoglobulin derived heavy chain that is comprised of  
15 domains or portions of immunoglobulin alpha chain and immunoglobulin gamma chain. The methods of the present invention may produce immunoglobulins containing a protection protein and an immunoglobulin derived light chain that is comprised of domains or portions of  
20 immunoglobulin kappa or lambda chain.

The methods of the present invention are operable on plant cells or part of a plant. The methods of the present invention may also included methods that further comprise growing the plant. The methods of the present  
25 invention may be applied to any plant including dicotyledonous, monocotyledonous, solanaceous, leguminous, alfalfa or tobacco plant. The methods of the present invention may be used to extract immunoglobulins from a portion of the plant such as a leaf, stem, root,  
30 tuber, seeds, fruit or entire plant. The methods of the present invention may use a mechanical device to shear the plants to release liquid from the apoplast or symplast of the plant. The plant pulp of the present

invention may be separated to remove the solid plant material using one of the following methods: centrifugation, settling, flocculation or filtration.

5 E. METHODS OF PRODUCING IMMUNOGLOBULINS CONTAINING PROTECTION PROTEINS

The present invention contemplates methods of producing an immunoglobulin containing a protection  
10 protein comprising the steps of:

- (a) Introducing into the plant cell an expression vector containing a nucleotide sequence encoding a protection protein operatively linked to a transcriptional  
15 promoter; and
- (b) Introducing into the same plant cell an expression vector containing a nucleotide sequence encoding an immunoglobulin derived heavy chain having at least a portion of an  
20 antigen binding domain operatively linked to a transcriptional promoter.

The methods of the present invention optionally include introducing into the plant cell containing the expression vector with the nucleotide sequences for the  
25 protection protein and the immunoglobulin derived heavy chain a nucleotide sequence encoding an immunoglobulin derived light chain at least having a portion of an antigen binding domain operatively linked to a transcriptional promoter. Methods are also contemplated that  
30 introduce into a cell that already contains nucleotide sequences and promoters operatively linked to encode a protection protein and an immunoglobulin heavy chain and an immunoglobulin light chain, a promoter operatively linked to a nucleotide sequence encoding J chain. This

results in a cell containing the nucleotide sequences operatively linked to promoters for an immunoglobulin heavy chain and an immunoglobulin light chain, J chain and a protection protein.

5       The plant cells of the present invention may be present as part of a plant that is capable of growth. Particularly useful plants for this invention include dicotyledonous, monocotyledonous, solanaceous, legumes, alfalfa, tomato, and tobacco plants.

10       The methods of the present invention include producing an assembled immunoglobulin having heavy, light and J chains and a protection protein within a eukaryotic cell. This eukaryotic cell is produced by introducing into that cell nucleotide sequences operatively linked  
15 for expression encoding an immunoglobulin derived heavy chain having at least a portion of an antigen binding domain, an immunoglobulin derived light chain having at least a portion of an antigen binding domain, an immunoglobulin J chain, and a protection protein. These  
20 nucleotide sequences are operatively linked for expression by attaching appropriate promoters to each individual nucleotide sequence or to more than one nucleotide sequence thereby placing two nucleotide sequences encoding various molecules in tandem.

25       The eukaryotic cell produced by the present methods which contains these nucleotide sequences encoding the immunoglobulin heavy, light and J chains and the protection protein is maintained under conditions which allow those molecules to reproduce and assemble into an  
30 immunoglobulin which contains the protection proteins of the present invention.

      The present invention also contemplates methods for making a particular immunoglobulin or antigen binding

domain or domains of an immunoglobulin resistant to environmental conditions and more stable by operatively linking a nucleotide sequence encoding at least a portion of an antigen binding domain derived from an immunoglobulin heavy chain to a nucleotide sequence encoding at least one domain derived from an immunoglobulin  $\alpha$  or  $\mu$  heavy chain to form a nucleotide sequence encoding a chimeric immunoglobulin heavy chain. That nucleotide sequence encoding the chimeric immunoglobulin heavy chain is expressed in a eukaryotic cell which also contains at least one other molecule such as a protection protein, an immunoglobulin derived light chain having at least a portion of an antigen binding domain and an immunoglobulin J chain. In preferred embodiments, the cell contains all of the molecules including an immunoglobulin derived light chain having an antigen binding domain which is complementary to the antigen binding domain present on the immunoglobulin derived heavy chain. This method allows the chimeric immunoglobulin heavy chain to assemble with at least one other molecule, for example, the immunoglobulin derived light chain having the complementary antigen binding domain and an immunoglobulin J chain and the protection protein to form an immunoglobulin containing the protection protein which is resistant to environmental conditions.

These immunoglobulins are resistant to environmental conditions and thus more stable when subjected to elevated or reduced temperatures, high or low pH, high ionic or low ionic concentrations proteolytic enzymes and other harsh conditions. Such harsh conditions are typically found in the environment within natural water sources, within the human body, for example within the

gut and on mucosal surfaces, and on the surface of an animal such as a mammal.

5 F. CHIMERIC IMMUNOGLOBULINS CONTAINING PROTECTION  
PROTEINS

The present invention contemplates immunoglobulins containing a protection protein in which the immuno-  
globulin domains comprising the heavy and light chain are  
10 derived from different isotopes of either heavy or light  
chain immunoglobulins. One skilled in the art will  
understand that using molecular techniques these domains  
can be substituted for a similar domain and thus produce  
an immunoglobulin that is a hybrid between two different  
15 immunoglobulin molecules. These chimeric immunoglobulins  
allow immunoglobulins containing protection proteins to  
be constructed that contain a variety of different and  
desirable properties that are conferred by different  
immunoglobulin domains.

20 The present invention also contemplates chimeric  
immunoglobulins, including heavy, light and J chain which  
contain less than an entire domain derived from a  
different molecule. The same molecular techniques may be  
employed to produce such chimeric immunoglobulins.

25 In preferred embodiments, the immunoglobulins of the  
present invention contain at least the  $C_H1$ ,  $C_H2$ ,  $C_H3$ ,  
domain of mouse IgG, IgG1, IgG2A, IgG2B, IgG3, IgA, IgE,  
or IgD. Other preferred embodiments of the present  
invention contain immunoglobulin domains that include at  
30 least the  $C_{\mu}1$ ,  $C_{\mu}2$ ,  $C_{\mu}3$ , or  $C_{\mu}4$  domain of mouse IGM.  
Preferred immunoglobulins include immunoglobulins that  
contain the domains of  $C_{\epsilon}2$ ,  $C_{\epsilon}3$ , and  $C_{\epsilon}4$  of mouse  
immunoglobulin IGE.

The present invention also contemplates chimeric immunoglobulins derived from human immunoglobulins. These chimeric immunoglobulins contain domains from two different isotopes of human immunoglobulin. Preferred immunoglobulins include immunoglobulins that contain immunoglobulin domains including at least the C<sub>H</sub>1, C<sub>H</sub>2, or C<sub>H</sub>3 of human IgG, IgG1, IgG2, IgG3, IgG4, IgA1, IgA2, IgE, or IgD. Other preferred immunoglobulins include immunoglobulins that contain domains from at least the C<sub>H</sub>1, C<sub>H</sub>2, C<sub>H</sub>3, or C<sub>H</sub>4 domain of human IgM or IgE. The present invention also contemplates immunoglobulins that contain immunoglobulin domains derived from at least two different isotopes of mammalian immunoglobulins. Generally, any of the mammalian immunoglobulins can be used in the preferred embodiments, such as the following isotopes: any isotype of IgG, any isotype of IgA, IgE, IgD or IgM. The immunoglobulins of the present invention contained at least one of the constant region domains from two different isotopes of mammalian immunoglobulin.

The present invention also contemplates immunoglobulins that contain immunoglobulin domains derived from two different isotopes of rodent immunoglobulin. The isotopes of rodent immunoglobulin are well known in the art. The immunoglobulins of the present invention may contain immunoglobulin derived heavy chains that include at least one of the following immunoglobulin domains: the C<sub>H</sub>1, C<sub>H</sub>2, or C<sub>H</sub>3 domain of a mouse IgG, IgG1, IgG2a, IgG2b, IgG3, IgA, IgE, or IgD; the C<sub>H</sub>1, C<sub>H</sub>2, C<sub>H</sub>3, C<sub>H</sub>4 domain of mouse IgE or IgM; the C<sub>H</sub>1, C<sub>H</sub>2, or C<sub>H</sub>3 domain of a human IgG, IgG1, IgG2, IgG3, IgG4, IgA1, IgA2, or IgD; the C<sub>H</sub>1, C<sub>H</sub>2, C<sub>H</sub>3, C<sub>H</sub>4 domain of human IgM or IgE; the C<sub>H</sub>1, C<sub>H</sub>2, or C<sub>H</sub>3 domain of an isotype of mammalian IgG, an isotype of IgA, IgE, or IgD; the C<sub>H</sub>1, C<sub>H</sub>2, C<sub>H</sub>3, C<sub>H</sub>4 domain



of a mammalian IgE or IgM; the C<sub>H</sub>1, C<sub>H</sub>2, or C<sub>H</sub>3 domain of  
an isotype of rodent IgG, IgA, IgE, or IgD; the C<sub>H</sub>1, C<sub>H</sub>2,  
C<sub>H</sub>3, C<sub>H</sub>4 domain of a rodent IgE or IgM; the C<sub>H</sub>1, C<sub>H</sub>2, or  
C<sub>H</sub>3 domain of an isotype of animal IgG, an isotype of IgA,  
5 IgE, or IgD; and the C<sub>H</sub>1, C<sub>H</sub>2, C<sub>H</sub>3, C<sub>H</sub>4 domain of an animal  
IgE or IgM. The present invention also contemplates the  
replacement or addition of protein domains derived from  
molecules that are members of the immunoglobulin super-  
family. The molecules that belong to the immunoglobulin  
10 superfamily have amino acid residue sequence and nucleic  
acid sequence homology to immunoglobulins. The molecules  
that are part of the immunoglobulin superfamily can be  
identified by amino acid or nucleic acid sequence  
homology. See, for example, p. 361 of Immunoglobulin  
15 Genes, Academic Press (1989).

#### Tetratransgenic Organisms:

The present invention also contemplates a tetra-  
transgenic organism which is comprised of cells having  
20 incorporated into the nucleic acid of that cell or plant  
within the cell four different transgenes, each encoding  
a different polypeptide. These transgenes are different  
in that the messenger RNA and polypeptides produced from  
that transgene are different from the messenger RNA and  
25 polypeptides produced from the other of the four  
transgenes. Thus, the number of transgenes referred to  
in the present invention does not include multiple copies  
of the same transgene as is commonly found in transgenic  
organisms. The present invention is directed to  
30 transgenic organisms having four transgenes which are not  
identical copies of other transgenes. The present  
invention does not exclude the possibility that each of  
the four different transgenes may be present in multiple

copies. However, at least four separate transgenes that are different are present within the cells of the transgenic organism.

In addition, the present invention contemplates that  
5 four different transgenes are related in that the transgenes encode a polypeptide that is part of a multipolypeptide molecule. Therefore, the present invention contemplates that each individual polypeptide chain of a multi-peptide molecule would be present on a  
10 transgene within a cell of the transgenic organism. The expression of each individual different polypeptide of the multi-peptide molecule allows the different polypeptides to associate together to form the multi-peptide molecule within the transgenic animal's cells. Thus, the  
15 present invention does not include within the four different transgenes in each individual cell, transgenes which encode polypeptides which do not associate together to perform a multi-peptide molecule. Examples of such transgenes encoding molecules that do not associate  
20 together are polypeptides for antibiotic resistance such as kanamycin or neomycin or thymidine kinase.

In preferred embodiments, the transgenes present within a transgenic organism of the present invention encode the following four different polypeptides: a  
25 protection protein; an immunoglobulin derived heavy chain having at least a portion of an antigen binding domain; an immunoglobulin derived light chain having at least a portion of an antigen binding domain; and an immunoglobulin J chain. In other preferred embodiments, one of  
30 the transgenes present in the transgenic organism encodes a chimeric immunoglobulin heavy, light or J chain. In other preferred embodiments, a transgene of the transgenic organisms of the present invention encode

either an immunoglobulin heavy chain derived at least in part from an IgA or a IgM immunoglobulin. Other preferred embodiments include transgenic organisms containing transgenes which encode at least a portion of the amino acid sequence derived from an immunoglobulin heavy chain derived from either an IgA or IgM immunoglobulin heavy chain.

The present invention contemplates transgenic organisms including mammals, plants, rodents, reptiles, insects, amphibians, fishes or other organisms. In preferred embodiments, the transgenic organism of the present invention is a plant or a mammal. Methods of producing such organisms are well known. See, i.e., U.S. Patents 4,736,866; 4,607,388; 4,870,009 and 4,873,191 which are hereby incorporated by reference.

The present invention also contemplates immunoglobulin that contain immunoglobulin derived heavy or immunoglobulin derived light chains that contain immunoglobulin domains which have been engineered to make those domains less immunogenic in a particular species. Typically, the immunoglobulin molecule is engineered as to be "humanized" in that it appears to be a human immunoglobulin even though derived from various other species.

25

#### EXAMPLES

The following examples illustrate the disclosed invention. These examples in no way limit the scope of the claimed invention.

1. Construction of DNA Vectors For Expression of  
Antibodies in Plants.

5 a. Isolation of the Nucleotide Sequences  
Encoding the Guy's 13 Immunoglobulin

Molecular cloning of the gamma and kappa chains of the Guy's 13 anti-S. mutans antibody was done by the procedures described in Ma et al., Eur. J. Immunol.,  
10 24:131 (1994). Briefly, mRNA was extracted from the Guy's 13 hybridoma cell line and converted to the cDNA by standard procedures. The cDNA was then amplified with the use of a pair of oligonucleotides specifically complementary to either the gamma or kappa cDNA.  
15 Amplification was catalyzed by Taq 1 polymerase using a thermal cycler as described. The amplified cDNAs were then digested with the appropriate restriction endonucleases and ligated into the corresponding restriction site in a standard plant expression vector.  
20 Numerous examples of such vectors have been reported in the literature and are generally available. An example of one vector that may be used is pBIN19.

In a related series of experiments, the cDNAs were cloned into the bacterial vector bluescript. Using this  
25 construct, the sequence of the gamma and kappa cDNAs was determined using the methods of Maxam and Gilbert.

Procedures for cloning antibody cDNAs involving PCR techniques or by construction of cDNA libraries followed by ligation of the obtained cDNAs into appropriate  
30 vectors are commonplace techniques which are familiar to one of ordinary skill in the art.

- b) Hybrid cDNAs encoding the Guy's 13 heavy chain variable region, a part of the gamma chain constant region and a part of an alpha chain constant region.

5

These constructs were synthesized as described in Ma et al., Eur. J. Immunol., 24:131 (1994) and ligated into the appropriate plant expression vectors as described above. The final construct had the structure: Guy's 13  
10 variable region - (IgG1 C<sub>H</sub>1) - (IgG1 C<sub>H</sub>2) - (IgA C<sub>H</sub>2) - (IgA C<sub>H</sub>3), referred to as IgG2A heavy chain, and Guy's 13 variable region - (IgG1C<sub>H</sub>1) - (IgACH2) - (IgACH3).

15

- c) The Protection Protein and J chain.

The cloned rabbit polyimmunoglobulin receptor (pIgR) cDNA was described by Mostov, Nature, 308:37 (1984) and shown in Figure 8. The protection protein portion was obtained by PCR amplification of a portion of the  
20 nucleotide sequence coding for the (pIgR) and ligation into appropriate plant expression vectors as described above. The protection protein portion of the pIgR used in these constructs included the codon for amino acid number 1 to the codon for amino acid number 606. The  
25 method to accomplish this construction are well known in the art and the oligonucleotides can be selected using the pIgR nucleic acid sequence.

30

- d) cDNAs encoding aglycosylated derivatives of heavy-chain constant regions.

Mutagenesis procedures were performed either according to Stratagene protocols. In each case (i.e. alpha constant region, or protection protein) the codon  
35 for the asparagine utilized as the attachment site for carbohydrates, was changed to a codon for histidine.

2. Production of Transgenic Plants Expressing  
Therapeutic Antibodies.

5       Plants and plant cells containing immunoglobulins  
having a protection protein were produced in the  
following manner.

10           a) Transfer of vectors to Agrobacterium  
tumefaciens

Plant transformation was accomplished by using  
Agrobacterium tumefaciens. E. coli DH5 $\alpha$  bearing the  
recombinant pMON530 plant expression vector were mated  
15 with Agrobacterium in the presence of a helper strain  
(pRK2013) to provide transfer functions. Alternatively,  
pMON530 plasmid DNA was introduced into Agrobacteria by  
direct transformation. In this procedure, the  
Agrobacterium strain was first grown overnight at 28° C in  
20 YEP medium. 2 ml of the overnight culture was used to  
inoculate 50 ml of YEP and was grown to an OD<sub>600</sub> Of 1.0.  
The cells were then chilled to 4° C, pelleted by  
centrifugation and resuspended in 1 ml of ice cold 20 mM  
CaCl<sub>2</sub>. About 1  $\mu$ g of DNA was added to aliquots of 0.1 ml  
25 of ice cold cells. The cells were then rapidly frozen by  
immersion in liquid nitrogen or in a dry ice ethanol  
bath. The cells were thawed by incubation at 37° C for 5  
minutes followed by the addition of 1 ml YEP medium. The  
cells were allowed to incubate for 2-4 hours with gentle  
30 shaking. Individual colonies carrying the recombinant  
vector were isolated by incubation on YEP agar plates  
containing the appropriate antibiotic.

Agrobacteria containing pMON530 were grown in media  
containing kanamycin, spectinomycin and chloramphenicol.

Small segments of tobacco leaf were then co-cultivated with the *Agrobacterium* for 2 days after which the leaf segments were transferred to plates containing carbenicillin to kill the *Agrobacterium*. Regeneration of transformed leaf cells into whole plants was allowed to proceed in the presence of kanamycin selection until the plants were competent for growth in soil.

10                   b)   Regeneration of transformed tobacco and petunia plants.

Leaves from greenhouse grown tobacco or petunia plants were sterilized in 20% (by volume) Chlorox bleach, 0.1% sodium dodecyl sulfate at room temperature for 8 minutes. The leaves were then briefly rinsed in 70% ethanol and allowed to dry in sterile Petri plates.

Leaf discs of approximately 0.5 cm diameter were removed with a sterile hole puncher and placed on agar plates containing MS10 medium (MS10 medium per liter: 4.4 g Murashige and Skoog basal salts with minimal organics [Sigma #M68991, 30 g sucrose, 0.2 mg naphthalene acetic acid, 2 mg benzylaminopurine, 0.1 mg nicotinic acid, 0.1 mg pyridoxin, 0.1 mg thiamine, 10 g agar, pH 5.7 with KOH).

25           A 2 ml aliquot of a suspension of *Agrobacterium* in LB (approximately  $1 \times 10^8$  *Agrobacteria* per ml) was then added to the leaf pieces. All surfaces of the leaf discs were contacted with *Agrobacteria*, excess liquid was poured off the plate, and the discs were co-cultivated with the bacteria for 2 days at room temperature. The discs were then transferred to agar plates containing MS10 medium, 50  $\mu$ g/ml kanamycin and 250  $\mu$ g/ml carbenicillin (MS10-KC). Regeneration was allowed to proceed with weekly transfer of discs to fresh MS10-KC

plates until regenerating shoots were visible. Shoots were then transferred to agar plates containing MSO-KC medium (MSO-KC per liter: 4.4 g Murashige and Skoog basal salts with minimal organics [Sigma #M68991, 30 g sucrose, 1 mg nicotinic acid, 1 mg pyridoxin, 0.1 mg thiamine, 50  $\mu$ g/ml kanamycin and 250  $\mu$ g/ml carbenicillin, 10 g agar, pH 5.7 with KOH).

After root formation, plantlets were transferred to soil and grown to maturity.

10

c) Regeneration of transformed alfalfa Plants.

Alfalfa trifoliate were cut from a greenhouse grown plant and sterilized in 20% (v/v) Chlorox bleach, 0.1% sodium dodecyl sulfate at room temperature for 8 minutes. The trifoliate were then briefly rinsed in 70% ethanol and allowed to dry in sterile Petri plates.

Leaf pieces of approximately 1 cm X 4 mm were cut with a sterile scalpel and placed on agar plates containing B5H medium (B5H medium per liter: 3.1 g Gamborg's powdered medium (Sigma #G5893), 500 mg KNO<sub>3</sub>, 250 mg MgSO<sub>4</sub> 7H<sub>2</sub>O, 30 g sucrose, 500 mg proline, 1 mg 2,4-dichlorophenoxyacetic acid, 100  $\mu$ g kinetin, 100 mg inositol, 1 mg nicotinic acid, 1 mg pyridoxin, 10 mg thiamine, 10 g agar, 30 ml stock amino acids, pH 5.7 with KOH; stock amino acids consist of 26.6 g L-glutamine, 3.32 g serine, 16.8 mg adenine, 333 mg glutathione per liter and are added after autoclaving when the medium is approximately 50° C).

To the leaf pieces was then added 2 ml of a suspension of Agrobacterium in LB (approximately  $1 \times 10^8$  Agrobacteria per ml). All surfaces of the leaf were contacted with Agrobacteria, excess liquid was poured off



the plate, and the leaves were co-cultivated with the bacteria for 2 days at room temperature. The leaf pieces were then transferred to agar plates containing B5H medium, 25  $\mu\text{g/ml}$  kanamycin and 250  $\mu\text{g/ml}$  carbenicillin (B5H-KC). Regeneration was allowed to proceed with weekly transfer of leaf pieces to fresh B5H-KC plates until somatic embryos were visible. Embryos were then transferred to agar plates containing BI02Y-KC medium (BI02Y-KC per liter: 25 ml macronutrients, 10 ml micronutrients, 25 ml iron, 1 ml vitamins, 1 ml aminos, 2 g yeast extract, 100 mg myo-inositol, 30 g sucrose, 10 g agar, 25 mg kanamycin, 250 mg carbenicillin, pH 5.9 with KOH; macronutrients consist of 40 g  $\text{KNO}_3$ , 40 g  $\text{NH}_4\text{NO}_3$ , 13.88 g  $\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ , 1.4 g  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ , 2.6 g  $\text{KCl}$ , 12 g  $\text{K}_2\text{HPO}_4$  per liter yielding a 40X stock; vitamins consist of 100 mg thiamine HCl, 500 mg nicotinic acid, 100 mg pyridoxin-HCl per liter yielding a 1000X stock; aminos consists of 2 g per liter glycine yielding a 1000X stock; micronutrients consist of 580 mg  $\text{MnSO}_4 \cdot 4\text{H}_2\text{O}$ , 1550 mg  $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ , 160 mg  $\text{H}_3\text{BO}_3$ , 80 mg  $\text{KI}$  per liter yielding a 100X stock; iron consists of 1.28 g  $\text{NaFeEDTA}$  per liter yielding a 40X stock).

After root formation, plantlets were transferred to soil and grown to maturity.

d) Regeneration of Transformed Tomato Plants.

Cotyledons from 7 day old tomato seedlings were sterilized in 20% (v/v) Chlorox bleach, 0.1% sodium dodecyl sulfate at room temperature for 8 minutes. The leaves were then briefly rinsed in 70% ethanol and allowed to dry in sterile Petri plates.

Cotyledon pieces of approximately 0.5 cm diameter were cut with a sterile scalpel and placed on agar plates

containing MS4 medium (MS4 medium per liter: 4.4 g Murashige and Skoog basal salts with minimal organics [Sigma #M68991, 30 g sucrose, 2 mg zeatin riboside, 5 mg nicotinic acid, 0.5 mg pyridoxin, 0.5 mg thiamine, 1 mM acetosyringone, 10 g agar, pH 5.7 with KOH).

To the leaf pieces was then added 2 ml of a suspension of *Agrobacterium* in LB (approximately  $1 \times 10^8$  *Agrobacteria* per ml). All surfaces of the leaf discs were contacted with *Agrobacteria*, excess liquid was poured off the plate, and the discs were co-cultivated with the bacteria for 2 days at room temperature. The discs were then transferred to agar plates containing MS4 medium minus acetosyringone containing 50  $\mu$ g/ml kanamycin and 250  $\mu$ g/ml carbenicillin (MS4-KC). Regeneration was allowed to proceed with weekly transfer of discs to fresh MS4-KC plates until regenerating shoots were visible. Shoots were then transferred to agar plates containing MSO-KC medium (MSO-KC per liter: 4.4 g Murashige and Skoog basal salts with minimal organics [Sigma #M68991, 30 g sucrose, 1 mg nicotinic acid, 1 mg pyridoxin, 10 mg thiamine, 50  $\mu$ g/ml kanamycin and 250  $\mu$ g/ml carbenicillin, 10 g agar, pH 5.7 with KOH).

After root formation, plantlets were transferred to soil and grown to maturity.

e) Regeneration of Transformed *Arabidopsis* Plants.

Intact roots derived from *Arabidopsis thaliana* plants grown in sterile culture were first pretreated on callus inducing medium (CIM) for 3 days at 28° C in the dark (CIM medium per liter: 3.1 g Gamborg's powdered medium (Sigma #G5893), 30 g sucrose, 1 mg 2,4-dichlorophenoxyacetic acid, 100  $\mu$ g kinetin, 1 mg

inositol, 0.1 mg nicotinic acid, 0.1 mg pyridoxin, 0.1 mg thiamine, 8 g agar, pH 5.7 with KOH).

To the intact roots was then added 2 ml of a suspension of Agrobacterium in LB (approximately  $1 \times 10^8$  Agrobacteria per ml). All surfaces of the roots were contacted with Agrobacteria and excess liquid was poured off the plate. The intact roots were then cut into 5 mm segments and were co-cultivated with the Agrobacteria for 2 days at 28° C on CIM plates. The root pieces were then transferred to agar plates containing shoot inducing medium (SIM) containing 50 µg/ml kanamycin and 250 µg/ml carbenicillin (SIM medium per liter: 3.1 g Gamborg's powdered medium (Sigma #G5893), 30 g sucrose, 5 mg N<sup>6</sup>-(2-isopentenyl) adenine, 150 µg indole-3-acetic acid, 1 mg inositol, 0.1 mg nicotinic acid, 0.1 mg pyridoxin, 0.1 mg thiamine, 8 g agar, pH 5.7 with KOH).

Regeneration was allowed to proceed with weekly transfer of root pieces to fresh SIM plates until green regenerating shoots were visible. Shoots were then transferred to agar plates containing EM medium (MSO-KC per liter: 4.4 g Murashige and Skoog basal salts with minimal organics [Sigma #M6899], 10 g sucrose, 1 mg indole-3-butyric acid 1 mg nicotinic acid, 0.1 mg pyridoxin, 0.1 mg thiamine, 250 µg/ml carbenicillin, 8 g agar, pH 5.7 with KOH).

After root formation, plantlets were transferred to soil and grown to maturity.

### 3. Identification of Transgenic Plants.

Kanamycin resistant transformants expressing individual immunoglobulin chains were identified by ELISA as described. Further analysis of the transformants included evaluation of RNA by Northern blotting and

evaluation of immunoglobulin polypeptides by Western blotting, both as described in Maniatis et al.

For each immunoglobulin chain, antigenic material, RNA or protein were detected by the respective assays.

5 Transformants identified as having the highest levels of immunoglobulin chains were used in cross pollination protocols.

10 4. Assembly of Antibodies by Cross Pollination of Transformants.

Cross pollinations were performed in order to obtain plants co-expressing the various components of the desired antibodies. These crosses yielded alfalfa,  
15 tomato, tobacco and Arabidopsis plants containing the following assembled components, all of which also contained the Guy's 13 antigen binding domain.

Type of Antibody	Immunoglobulin Components
20	1 G1 heavy chain, kappa light chain
	2 G2/A heavy chain, kappa light chain
	3 G2/A heavy chain, kappa light chain, J chain
	4 G1/A heavy chain, kappa light, J chain, protection protein
25	5 G1/A heavy chain Kappa light chain

30 5. Extraction and Evaluation of Guy's 13 Type 1, 2 and 3 & 4 Antibodies From Transgenic Plants.

a) Extraction and enrichment of antibody contained in leaf.

Leaf pieces were chopped into approximately 1 cm<sup>2</sup>  
35 pieces. The pieces were then added to a cold solution of TBS having 10µg/ml leupeptin (1 ml TBS per gram of leaf)

contained in a chilled porcelain mortar both at approximately 4° C. Plant liquid was extracted by pulverizing the pieces with a cold pestle using a circular motion and hand pressure. Pulverizing was continued  
5 until the pieces became a nearly uniform pulp (approximately 3 minutes of pulverizing). The pulp was centrifuged at 4° C and approximately 50,000 X g to yield a supernatant devoid of solid plant pieces. Alternatively, the pulp was filtered through a plastic  
10 mesh with a pore size of approximately 100 microns.

Depending on the titer of antibody contained in the particular plant, the supernatant was either directly suitable for exposure to antigen or required enrichment to a suitable concentration. Yields of IgG1's or IgG/A's  
15 in the crude extract were routinely less than 10 µg/ml and averaged approximately 5 µg/ml. For applications of a Guy's 13 antibody to mucosal surfaces, enrichment to a concentration of 1 to 4 mg/ml may be required. As a Type 1, 2 or 3 construct, Guy's 13 antibody required a ten to  
20 forty-fold enrichment to yield the desired concentration. This was accomplished either by affinity adsorption (utilizing either Protein A or Protein G), or by lyophilization to remove water. Size exclusion chromatography was also used for enrichment but required  
25 complete fractionation of the crude extract to yield an antibody of the required concentration. By ELISA assay and by polyacrylamide gel electrophoresis, the co-expressed chains assembled into a complex of approximately 180-200 k daltons for types 1 & 2 and  
30 approximately 400 k daltons for type 3. Crude extracts were routinely obtained containing approximately of 5-10 µg/ml.

A dramatic increase in antibody accumulation was observed when the protection protein was crossed into a plant containing Type 3 antibody yielding a plant containing a Type 4 antibody. By ELISA assay and by  
5 polyacrylamide gel electrophoresis, the co-expressed chains assembled into a complex of approximately 470,000 daltons. Crude extracts were routinely obtained containing in excess of 200  $\mu\text{g/ml}$  with an average of approximately 250  $\mu\text{g/ml}$ . Therefore, the SIgA construct  
10 of the Guy's 13 antibody required minimal enrichment to achieve the target concentration. This enrichment could be accomplished by the techniques described above. Alternatively, it was found that the antibody is readily separated from the majority of plant molecules by a one  
15 ultrafiltration step using membrane with a molecular exclusion of 200,000 d.

b) Functionality of the Guy's 13 Type 4 Antibody.

20

Functional antibody studies were carried out by ELISA. All plants expressing antibody light and heavy chains assembled functional antibody that specifically recognized streptococcal antigen (SA I/II). The levels  
25 of binding and titration curves were similar to those of mouse hybridoma cell supernatants. No SA I/II binding was detected with plants expressing only J chain or only protection protein. Likewise, wild-type plants expressing no immunoglobulin showed no detectable levels  
30 of binding.

In a similar set of experiments, binding of antibody to immobilized purified streptococcal antigen or native antigen on the bacterial cell surface was detected using an anti-secretory component antiserum. In these assays,

only the Type 4 antibody binding was detected. The functional Type 1, 2 or 3 antibodies did not bind the anti-secretory component antiserum. These results confirm that the protection protein was assembled with antibody in the plants expressing Type 4 constructs and in a manner which did not interfere with antigen binding.

#### 6. Expression of Chimeric Immunoglobulins.

The genes encoding the heavy and light chains of a murine monoclonal antibody (mAb Guy's 13) have been cloned and expressed in *Nicotiana tabacum*. Transgenic plants have been regenerated that secrete full-length Guy's 13 antibody. By manipulation of the heavy chain gene sequence, constant region domains from an immunoglobulin alpha heavy chain have been introduced, and plants secreting Guy's 13 mAb with chimeric gamma/alpha heavy chains have also been produced. For each plant antibody, light and heavy chains have been detected by Western blot analysis and the fidelity of assembly confirmed by demonstrating that the antibody is fully functional, by antigen binding studies. Furthermore, the plant antibodies retained the ability to aggregate streptococci, which confirms that the bivalent antigen-binding capacity of the full length antibodies is intact.

#### a) Cloning of heavy and light chain genes

Messenger RNA was purified from the Guy's 13 and a murine IgA (MOPC315) hybridoma cell line, using an acid guanidiniumthiocyanate-phenol-chloroform extraction. Complementary DNA was made using Moloney murine leukemia virus reverse transcriptase (Promega, GB). DNA encoding the gamma and kappa chains of Guy's 13 were amplified by

polymerase chain reaction (PCR). The degenerate oligonucleotides used in the PCR were designed to incorporate a 5' terminal XhoI, and a 3'-terminal EcoRI restriction site in the amplified DNA fragments.

5 Following restriction enzyme digestion, the immunoglobulin chain encoding DNA was ligated into a constitutive plant expression vector (pMON 530), which contains a mouse immunoglobulin leader sequence upstream of the cloning site. The recombinant vector was used to  
10 transform *E. coli* (DH5- $\alpha$ , Gibco BRL) and screening was by Southern blotting, using radiolabeled DNA probes derived from the original PCR products. Plasmid DNA was purified from positive transformants and introduced into *Agrobacterium tumefaciens*.

15 A similar approach was used to construct two forms of a hybrid Guy's 13 heavy chain. The synthetic oligonucleotides shown in Fig. 1 were used in PCR to amplify the regions: (a) Guy's 13 signal sequence to the 3' end of C $\tau$ 1 domain (J1-J5), (b) Guy's 13 signal sequence  
20 to the 3' end of C $\tau$ 2 domain (J1-J2), and (c) 5'end of C $\alpha$ 2 domain to the 3' terminus of DNA from the MOPC 315 hybridoma (J3-J4). The fragments were purified (Geneclean II, Bio 101, La Jolla, CA) and digested with HindIII for 1 h at 37°C. The Guy's 13 fragments were  
25 ligated to the MOPC 315 fragment with T4 DNA ligase (Gibco, BRL), at 16°C for 16 h, and an aliquot of the reaction mixture was used as template DNA for a further PCR, using the 5' terminal oligonucleotide for Guy's 13 (J1) and the 3' terminal oligonucleotide for MOPC 315  
30 (J4). Amplified DNA fragments were purified and ligated into the pMON 530 vector as described above. The vector used in this procedure did not have a previously inserted



mouse leader sequence, as in this case, the DNA encoding the native Guy's 13 leader sequence was included in the PCR amplification.

5                   b)   Plant transformation and regeneration

Leaf discs, about 6 mm in diameter, were cut from surface-sterilized tobacco leaves (*Nicotiana tabacum*, var. *xanthii*) and incubated overnight at 28°C, with a culture of the recombinant *A. tumefaciens*, containing immunoglobulin cDNA inserts. The discs were transferred to culture plates containing a medium that induces regeneration of shoots, supplemented with kanamycin (200 mg/l) and carbenicillin (500 mg/l). Shoots developing after this stage were excised and transplanted onto a root-inducing medium, supplemented with kanamycin (200 mg/l). Rooted plantlets were transplanted into soil as soon as possible after the appearance of roots. Plants were screened for expression of immunoglobulin chains as described below. Those that expressed heavy chains were crossed with those expressing light chains, by cross-pollination. The resulting seeds were sown in soil and allowed to germinate. Twenty-two transgenic plants were regenerated from transformations with light or heavy chain constructs, as determined by ELISA. Crossing of light and heavy chain-secreting plants resulted in 3/10 F1 progeny plants expressing kappa and gamma chains together, 4/17 plants expressing both kappa and the plant G1/A heavy chain and 3/8 plants expressing both kappa and the plant G2/A heavy chain together.

30           The three different forms of Guy's 13 monoclonal antibody expressed in plants, therefore, all contain the identical light (kappa) chain, but different heavy chains. These will be abbreviated throughout this report

as follows (Fig. 1): Guy's 13 IgG1 with original gamma heavy chain, plant G13, Guy's 13 with IgG/IgA hybrid heavy chain consisting of var- $\tau$ 1- $\tau$ 2- $\alpha$ 2- $\alpha$ 3 domains, plant G2/A. The Guy's 13 hybridoma cell culture supernatant used as a positive control will be abbreviated to Mouse G13. Negative control plants were those that had been transformed with pMON 530 vector containing an insert that encodes an irrelevant mouse protein.

10                   c)   Antibody chain detection

Production of either gamma, kappa or the gamma/alpha chain hybrids was detected by ELISA. Microtiter wells were coated with a goat anti-mouse heavy or light chain-specific IgG (Fisher, USA; Sigma, GB; Nordic Pharmaceuticals, GB) in 150 mM NaCl, 20 mM Tris-HCl (pH 8) (TBS). Blocking was with 5% non-fat dry milk in TBS at 4°C overnight. Plant leaves were homogenized in TBS with leupeptin (10  $\mu$ g/ml) (Calbiochem, USA). The supernatant was added in serial twofold dilutions to the microtiter plate and incubation was at 4°C overnight. After washing with TBS with 0.05% Tween 20, bound immunoglobulin chains were detected with the appropriate goat anti-mouse heavy or light chain-specific antibody, conjugated with horseradish peroxidase (Fisher; Sigma; Nordic Pharmaceuticals), for 2 h at 37°C. Detection was with 2,2'-azino-di-(3-ethyl-benzthiazoline-sulfonate) (Boehringer, FRG).

A similar assay was used to determine the concentrations of the murine and plant Guy's 13 antibodies. These were compared with a mouse IgG1 mAb (MOPC 21), and a mouse IgA mAb (TEPC 21) used at known concentrations (Sigma). ELISA plates were coated with an anti-mouse

kappa antiserum. After blocking, bound antibody was detected with horseradish peroxidase-labeled anti-mouse gamma or alpha antiserum. Antibody concentration was determined by comparison of binding curves for each  
5 antibody.

ELISA was also used to detect the binding function of the assembled antibody. Binding to SA I/II was detected using microtiter plates that had been coated with purified SA I/II at an optimized concentration of 2  
10  $\mu\text{g/ml}$ . The ELISA procedure was as described above. The ability to bind *S. mutans* or *E. coli* cells was detected using intact cells (strains Guy's c, *S. mutans* and DH5- $\alpha$ , *E. coli*) that had been grown to stationary phase, for 18 h at 37°C and fixed in 10% formalin. All the antibody  
15 solutions were adjusted to an initial concentration of 1.5  $\mu\text{g/ml}$  and used in serial twofold dilutions. Extracts from plants expressing wither Guy's 13 heavy or light chain singly were also included in these assays, to determine if the single immunoglobulin chains exhibited  
20 any antigen-binding activity. Antibodies bound to either cells or purified SA I/II were detected using a horseradish peroxidase-conjugated goat anti-mouse light or heavy chain antiserum (Nordic Pharmaceuticals). The results are expressed as mean  $\pm$  standard deviation of  
25 duplicate results from three separate assays.

Competition ELISA was performed on microtiter plates coated with purified SA I/II as above. The plates were incubated with plant extracts of Guy's 13 hybridoma supernatant at 1.5  $\mu\text{g/ml}$  and serial twofold dilutions at  
30 37°C for 1 h and 4°C overnight. After washing,  $^{125}\text{I}$ -labeled mouse Guy's 13 was added and left to incubate for 2 h at 37°C. The plates were washed again and the bound

radioactivity was counted in a gamma counter (Hydragamma  
16, Innotec, GB). The results are expressed as %  
inhibition of labeled mouse Guy's 13 binding, in which  
100% is the radioactive count from wells to which no  
5 blocking solution had been added.

d) Western blot analysis

Aliquots of 10 $\mu$ l of leaf homogenates were boiled  
with 75 mM Tris-HCl (pH 6.8), 2% SDS, under reducing and  
10 nonreducing conditions. SDS-PAGE in 10% acrylamide was  
performed, and the gels were blotted onto nitrocellulose.  
The blots were incubated for 16 h in TBS with 0.05% Tween  
20 and 1% non-fat dry milk, followed by goat anti-mouse  
IgG1, kappa (Nordic Pharmaceuticals) or alpha chain-  
15 specific antisera (Sigma), and incubated for 2 h at 37°C.  
After washing, the second-layer antibody, an alkaline  
phosphatase-conjugated rabbit anti-goat IgG (Sigma) was  
applied for 2 hours at 37°C. Antibody binding was  
detected by incubation with 300  $\mu$ g/ml nitroblue  
20 tetrazolium and 15p  $\mu$ g/ml 5-bromo-4-chloro-3-idolyl  
phosphate (Promega).

e) DNA sequencing

The DNA sequence of each cloned immunoglobulin gene  
25 insert confirmed that no mutations had occurred during  
PCR amplification or the cloning procedures. The  
introduction of the HindIII site in the  $\lambda/\gamma$  hybrid heavy  
chains resulted in the predicted addition of the leucine  
residue between the Cy2 and C $\alpha$ 2 domains in Plant G2/A and  
30 leucine-lysine between the Cy1 and C $\alpha$ 2 domains in Plant  
G1/A. The additional Cy2 domain in the Plant G2/A  
construct is predicted to increase the length of the

heavy chain by 141 amino acid residues (approximately 12000 Da). The plant G1/A heavy chain is predicted to be slightly larger than the native Guy's 13 heavy chain, by 33 amino acids, approximately 3000 Da.

5 Plasmid DNA that was purified from positive transformants in *E. coli* was sequenced. The immunoglobulin gene inserts were excised and sub-cloned into Bluescript (Stratagene, USA). The DNA sequence was determined by a di-deoxy termination procedure (Sequenase, USB, USA).

10

f) Expression of assembled antibody

Western blot analysis on extracts from three representative F1 progeny plants was performed and reported in Figure 2 of Ma et al., Eur. J. Immunol., 24:131-138  
15 (1994). Samples run under reducing conditions demonstrate the presence of light (kappa) chain at approximately 25 Kd, in the mouse Guy's 13, as well as in the three transgenic plants, but not in the control plant. Guy's 13 heavy (gamma) chain was also detected in  
20 plant G13 at approximately 57 Kd, but not in the control plant extract. A single protein species was detected, unlike the hybridoma producing the Guy's 13 antibody cell culture supernatant, in which a two protein species was a consistent finding. The difference in the molecular size  
25 of the mouse heavy chains is probably due to glycosylation differences, and the result suggests that in plants the two heavy chains may be glycosylated in the same way.

The heavy chains of plant G1/A and G2/A were  
30 detected with an anti-alpha chain antiserum. Compared with the mouse Guy's 13 heavy chain, (approximately 57 Kd), the heavy chain of plant G1/A has a slightly higher relative molecular mass (approximately 60 Kd) and the

plant G2/A heavy chain is much larger (approximately 70 Kd). This is consistent with the molecular weights predicted by sequence analysis. Several other protein species were detected in the transgenic plant extracts.

5 These are likely to be proteolytic fragments of either light/heavy chain complexes, or of the heavy chain, as no bands were detected in the extract from the control transgenic plant. The anti-alpha chain antiserum did not cross-react with the mouse Guy's 13, which only contains  
10 gamma chain domains.

Samples were also run under nonreducing conditions to confirm the assembly of heavy and light chains into an immunoglobulin molecule and reported in Figure 3 of Ma et al., Eur. J. Immunol., 24:131-138 (1994). Detection was  
15 with a labeled anti-kappa antiserum, and all three transgenic plants had assembled immunoglobulin at the correct  $M_r$  of above 150 Kd for full-length antibody. The plant G13 antibody has the same  $M_r$  as the mouse G13, but the plant G2/A and plant G1/A antibodies have higher  $M_r$  as  
20 predicted. A number of smaller proteolytic fragments were also detected, which is consistent with previous findings and the fact that a number of proteases are released by plants during the antibody extraction procedure. That these are antibody fragments, is  
25 confirmed by the absence of any detectable bands in the control plant extract.

#### g) Antigen binding

Ten plants which were producing immunoglobulin were  
30 made in total, and the concentration of immunoglobulin in plant extracts varied between 1 and 10  $\mu\text{g/ml}$  (mean 4.5  $\mu\text{g/ml}$ ). For the murine antibody and the representative plants used in this study, the concentrations estimated

by ELISA were: mouse IgG-15.4  $\mu\text{g/ml}$ , plant IgG-7.7  $\mu\text{g/ml}$ , plant G1/A-1.5  $\mu\text{g/ml}$  and plant G2/A-2.1  $\mu\text{g/ml}$ . The concentrations determined for plant antibodies containing hybrid heavy chains are possibly underestimated, as they  
5 do not carry all of the constant region determinants, as compared with the standard mAb IgA used.

Titration curves for extracts from the three representative transgenic plants binding to SA I/II were generated and reported in Figure 4 of Ma et al., Eur. J. Immunol., 24:131-138 (1994). Specific antibody was  
10 detectable in all three transgenic plant extracts, and the titration curves were similar to that of the murine hybridoma cell culture supernatant, used at the same concentration. The binding of the plant G1/A antibody  
15 appeared to be slightly lower than the other antibodies, although the titration curve followed a similar pattern. No SA I/II binding activity was detected in the negative control plant nor did extracts from plants individually expressing light or heavy chains have binding activity  
20 towards purified SA I/II. These findings demonstrate that the transgenic plants expressing both light and heavy chains have assembled the antibody molecule correctly to form a functional antigen binding site and that single light or heavy chains are not capable of  
25 binding the antigen.

The plant antibodies also recognized native antigen on the surface of streptococcal cells as shown in Figure 5 of Ma et al., Eur. J. Immunol., 24:131-138 (1994) (*S. mutans* serotype c), which further confirms the integrity  
30 of the antigen-binding site in the plant antibodies. There were no significant differences between the binding of the different antibodies. Neither extracts from control plants, nor plants expressing only heavy or light

chains showed any binding to *S. mutans* cells. There was no binding to *E coli* cells by any of the plant extracts, at concentrations of 1.0 and 0.5 µg/ml.

The plant antibodies competed with the original  
5 mouse Guy's 13 mAb for binding to SA I/II. Up to 85% inhibition of <sup>125</sup>I-labeled mouse Guy's 13 mAb binding to SA I/II was demonstrated using the plant antibodies as shown in Figure 6 of Ma et al., Eur. J. Immunol., 24:131-138 (1994). As before, the inhibition titration curves  
10 of the plant antibodies were similar to each other, and comparable to that of the mouse Guy's 13, whereas the control plant extract gave no inhibition.

h) Aggregation of *S. mutans*

15 The action of the immunoglobulin produced in plants having the Guy's 13 antigen binding region on bacteria was determined and reported in Figure 7 of Ma et al., Eur. J. Immunol., 24:131-138 (1994). Plant extracts were sterilized by filtration through a 0.22 µm pore size  
20 filter and diluted tenfold with Todd Hewitt broth. The samples were inoculated with 0.05 vol of an overnight *S. mutans* culture and incubated at 37°C overnight. The samples were Gram stained and examined under oil immersion microscopy. *S. mutans* grown in the presence of  
25 mouse Guy's 13, plant Guy's 13, plant G1/A or plant G2/A became aggregated and cell clumping was evident. However, the control plant extract had no effect on *S. mutans* growth. None of the plant mAb appeared to affect  
30 *S. mutans* rate of growth, as determined by culture of viable organisms at 8, 12 and 16 h. This result demonstrates not only that the plant antibodies have correctly assembled antigen-binding regions, but also that the antibody molecules bind antigen bivalently.



Example 7. PRODUCTION OF IMMUNOGLOBULINS CONTAINING  
PROTECTION PROTEINS

5 Four transgenic *Nicotiana tabacum* plants were generated to express (1) a murine monoclonal immunoglobulin kappa chain having the antigen binding site of the Guy's 13 light chain, (2) a hybrid IgA/G murine immunoglobulin heavy chain containing C $\gamma$  and C $\alpha$   
10 chain domains and the antigen binding site of the Guy's 13 heavy chain, (3) a murine J chain and (4) protection protein comprised of amino acids 1-606 of rabbit polyimmunoglobulin receptor and did not contain amino acids 627-675 of the rabbit polyimmunoglobulin receptor.  
15 See, Example 1. Successive sexual crosses between these plants resulted in simultaneous expression of all four protein chains in the progeny plants. In some cases, back crossing was used to produce homozygous plants. The four recombinant polypeptides were assembled into a  
20 functional, high molecular weight immunoglobulin containing a protection protein of approximately 470,000 Kd. The assembly of the protection protein with the immunoglobulin was dependent on the presence of a J chain, as no association of the protection protein was  
25 detected when plants expressing antibody alone were crossed with those expressing the protection protein. Microscopic evaluation of plants expressing the immunoglobulins containing the protection protein demonstrated co-incident expression of protection protein  
30 and immunoglobulin heavy chains in single cells. Single cells are able to produce immunoglobulin having a protection protein in transgenic plants, whereas two cells are required for natural production of secretory

immunoglobulin in mammals. The results demonstrate that sexual crossing of transgenic plants expressing recombinant sub-units is suitable for large scale production of immunoglobulin containing a protection  
5 protein for passive immunotherapy, as well as for expressing other complex protein molecules.

The immunoglobulin which contains the protection protein has the heavy and light chain antigen binding domains from the Guy's 13 monoclonal antibody that  
10 specifically recognize the cell surface adhesion molecule SA 1/11 of an oral streptococcus as shown by Smith, R. & Lehner, T. *Oral Microbiol. Immunol.* 4, 153-158 (1989). Transgenic immunoglobulin of this type containing only heavy and light chains has been generated in *Nicotiana*  
15 *tabacum* plants as described in Example 6. A mouse J chain construct containing the coding length cDNA was amplified using synthetic oligonucleotide primers corresponding to the N terminus MKTHLL and the C terminus SCYPD of mouse J chain as described by Matsuuchi, L.,  
20 Cann, G. M. & Koshland, M.E. *PNAS* 83, 456-460 (1986). This amplified nucleotide sequence was ligated into a constitutive plant expression vector, pMON 530, that includes the 35S promoter from Cauliflower Mosaic Virus and has been described by Rogers, S. G., Klee, H. J.,  
25 Horsch, R. B. & Fraley, R. T. *Meth. Enzymol.* 153, 253-276 (1987). Tobacco leaf tissue was transformed using agrobacterium containing the recombinant plasmid as described in the previous Examples. Regenerated plants were screened for the production of messenger RNA  
30 encoding J chain and positive transformants were self fertilized in order to generate homozygous progeny. The J chain expressing plants were crossed initially with those expressing the chimeric immunoglobulin heavy chain

and kappa chain. Western blot analysis of the plant extract from plants expressing the chimeric immunoglobulin heavy chain with anti-kappa antiserum under non-reducing conditions, revealed a protein species of approximately 210 Kd, which is consistent with the presence of the extra constant region domains present in the chimeric immunoglobulin heavy chain, as compared with the original IgG1 antibody. The progeny from the cross between the plant expressing the immunoglobulin and a J chain plant resulted in the appearance of a major immunoglobulin band at approximately twice the relative molecular mass of approximately 400 Kd, demonstrating that assembly of the 3 polypeptides had occurred to form dimeric immunoglobulin (dlgA/G).

The protection protein construct consisted of a coding length cDNA amplified using synthetic oligonucleotide primers corresponding to the N terminus MALFLL and AVQSAE at amino acids 601-606 of the C terminus of rabbit polyimmunoglobulin receptor. The nucleotide sequence of the rabbit polyimmunoglobulin receptor was reported by Mostov, K. E., Friedlander, M. & Blobel, G. *Nature* 308, 37-43 (1984). The protection protein was generated in transgenic plants as described above and positive transformants expressing the protection protein were identified by Western blot analysis.

Plants expressing J chain assembled with the immunoglobulin having the IgA/G heavy chains to form dimers were then crossed with a homozygous plant expressing the protection protein. The progeny plants expressing the immunoglobulin having the protection protein contained a higher molecular weight protein species at approximately 470 Kd as determined by Western blot analysis under non-reducing conditions. This

molecular size was consistent with that expected for an immunoglobulin containing a protection protein. This high molecular weight protein contained the protection protein as confirmed by Western blotting, using antiserum that specifically recognized the protection protein. The plant extracts also contained a protein species of approximately 400 Kd corresponding to the dimers of IgA/G and a protein species of approximately 210 Kd corresponding to the immunoglobulin with the chimeric heavy chain, but these were only detected by anti-kappa antiserum and not the anti-protection protein antiserum. In the transgenic plant producing the protection protein alone, there was no evidence that the protection protein assembled with endogenous plant proteins or formed multimers, as no high molecular weight proteins were detected in Western blotting under non-reducing conditions. Western blot analysis demonstrated that extracts from the plants expressing immunoglobulin heavy chain (IgA/G, dimeric IgA/G and the immunoglobulin containing a protection protein), but not the plants containing only the protection protein or J chain or wild-type plants, contained identical immunoglobulin derived heavy and light chains. Furthermore, only the plants containing protection proteins and the plants containing the IgG/A immunoglobulin having the protection protein expressed proteins that were recognized by the antiserum that specifically recognized the protection protein. No cross reacting proteins were detected in extracts from the wildtype control plant.

In mammals, the assembly of secretory component with the immunoglobulin requires the presence of J chain as described by Brandtzaeg, P. & Prydz, H. *Nature* 311, 71-73 (1984). Plants expressing immunoglobulins containing a

chimeric heavy chain (IgA/G) were crossed with plants expressing protection protein. None of the 10 resulting progeny that expressed immunoglobulin and the protection protein without J chain produced assembled complexes as compared with the 10/10 plants that co-expressed J chain dimerized immunoglobulin and the protection protein without J chain, which assembled the  $M_r$  470 Kd immunoglobulin containing the protection protein. This confirms that J chain is required for the protection protein association with immunoglobulin as found in mammals. Only the approximately 210 Kd monomeric form of the immunoglobulin was recognized by anti-kappa antiserum, and the antisera that specifically bound the protection protein, recognized free protection protein, but no immunoglobulin heavy or light chains proteins.

Functional studies were carried out using the immunoglobulin produced in the 5 plant constructs using ELISA. All plants expressing immunoglobulin light and heavy chains, assembled functional immunoglobulin that specifically recognized streptococcal antigen (SA I/II). The levels of binding and titration curves were similar to those of the native mouse hybridoma cell supernatant. No SA I/II binding was detected in plants expressing only J chain or only protection protein or in wildtype plants. Binding of the immunoglobulins to immobilized purified streptococcal antigen or to native antigen on the bacterial cell surface was also detected using the antiserum which specifically binds the protection protein. In these assays, the binding of the immunoglobulin containing the protection protein to the streptococcal antigen was specifically detected. These results confirmed that the protection protein was assembled with the immunoglobulin to produce an

immunoglobulin containing a protection protein in a manner which did not interfere with antigen binding.

The assembly of heavy and light chains into functional immunoglobulin molecules in plants is very efficient as shown by Hiatt, A. C., Cafferkey, R. & Bowdish, K. *Nature* 342, 76-78 (1989). A signal peptide must be present on both heavy and light chain constructs to direct the recombinant proteins to the endoplasmic reticulum antibody for assembly to take place in plants as was previously shown by Hiatt, A. C., Cafferkey, R. & Bowdish, K. *Nature* 342, 76-78 (1989). This study has demonstrated the fidelity of immunoglobulin assembly which includes dimerization of monomeric antibody by J chain in the transgenic plants. These results demonstrated that in plants the dimeric immunoglobulin population represents a major proportion (approx. 57%) of the total antibody. These results also demonstrate the production of an assembled immunoglobulin containing a protection protein which binds the corresponding antigen as well as the parent murine monoclonal antibody, which makes up a major proportion of the total antibody when the protection protein is incorporated (approximately 45%).

Co-expression of dimeric immunoglobulin with the protection protein in plants has led to assembly of a functional immunoglobulin containing a protection protein. All four transgenes for this complex protein were introduced into plants with the identical pMON530 expression cassette and native leader sequences. This vector contains a promoter sequence derived from the 35S transcript of the cauliflower mosaic virus which directs expression of transgenes in a variety of cell types of most plant organs as has been described by Benfey, P. N.

& Chua, N-H. *Science* 250, 959-966 (1990); and Barnes, W. M. PNVAS 87,9183-9187 (1990). Directing expression of all four transgenes with the same promoter maximized the likelihood of coincidental expression in a common plant  
5 cell. Microscopic observation of plants expressing an immunoglobulin containing a protection protein revealed that many cell types of the leaves contain the individual protein components that make up the immunoglobulin. These proteins accumulated at highest concentration in  
10 bundle sheath cells and were confined by the cell walls of these and other cells, but were not found in intercellular spaces. Restriction of the largest immunoglobulin components, the protection protein and the chimeric immunoglobulin heavy chain, within the confines  
15 of a protoplasmic or apoplasmic compartment of individual cells would constrain the assembly of the secretory immunoglobulin to those cells in which all the component molecules are synthesized. The subcellular site(s) and mechanism of assembly remain to be determined, assembly  
20 of IgG heterotetramers in plants requires targeting of both proteins to the endomembrane system as has been previously shown by Hiatt, A. C., Cafferkey, R. & Bowdish, K. *Nature* 342, 76-78 (1989); and Hein, M. B., Tang, Y., McLeod, D. A., Janda, K. D. & Matt, A. C.  
25 *Biotechnol Prog.* 7, 455-461 (1991).

In addition, we have demonstrated that a protection protein derived from mature secretory component devoid of signals for membrane integration, transcytosis or subsequent proteolysis can be assembled with chimeric  
30 immunoglobulin heavy chain containing immunoglobulin gamma and alpha protein domains. These results demonstrate that the inherent functions of IgG constant regions (protein A binding, complement fixation, Fc

receptor activity) may be maintained in a dimeric immunoglobulin, capable of binding to a protective protein. These additional capabilities may be employed to enhance the function of an immunoglobulin used for passive immunotherapy and the development of plants capable of generating a functional immunoglobulin containing a protection protein will have significant implications in passive immunotherapy. The level of expression of the immunoglobulin containing a protection protein is high and the production can be scaled up to agricultural proportions, to allow economical production of monoclonal antibodies.

#### Methods

The following methods were used to prepare and analyze the Immunoglobulin of this Example.

i) Antibody assembly in transgenic *Nicotiana tabacum*.

Leaf segments were homogenized in 150mM NaCl 20mM Tris-HCl (pH8) (TBS), with leupeptin (10µg/ml). The extracts were boiled for 3 minutes, in 75mM Tris-HCl (pH6.8), 2% SDS, under non-reducing conditions and SDS-PAGE in 4% acrylamide was performed. The gels were blotted onto nitrocellulose. The blots were incubated for 2 hrs in TBS with 0.05% Tween 20 and 1% non-fat dry milk, followed by the appropriate antiserum and incubated for 2 hrs at 37°C. After washing, the second layer alkaline phosphatase conjugated antibody was applied for 2 hrs at 37°C. Antibody binding was detected by incubation with 300mg/ml nitroblue tetrazolium and 150mg/ml 5-bromo-4-chloro 3-indolyl phosphate.

These extracts were analyzed using western analysis to determine whether the immunoglobulins were assembled into immunoglobulin molecules by analyzing Western blots



of plant extracts prepared under non-reducing conditions, were with anti-kappa antiserum (Bradsure, UK) and an antiserum which specifically recognizes protection protein. The immunoglobulins produced in the plants were compared to the monoclonal IgG1 Guys 13 immunoglobulin described by Smith, R. & Lehner, T. *Oral Microbiol. Immunol.* 4, 153-158 (1989).

ii) Western Analysis.

Western analysis was performed on each of the plant extracts prepared under reducing conditions to identify individual protein components of the immunoglobulin. Samples of the various plant extracts were prepared as described previously, but with the addition of 5%  $\beta$ -mercaptoethanol. SDS-PAGE in 10% acrylamide was performed and the protein in the gels transferred to nitrocellulose. Individual proteins were detected using anti-mouse  $\gamma$ 1 heavy chain (Sigma, UK); anti-mouse kappa chain (Bradsure, UK); or an antiserum that specifically recognized the protection protein, followed by the appropriate alkaline phosphatase conjugated antibody.

iii) Western Analysis to Show Production of Immunoglobulin Having a Protection Protein

Western analysis of transgenic plant extract was performed as described in ii) above. The plant extracts from plants expressing the immunoglobulin containing the protection protein were subjected to SDS-PAGE under both non-reducing and reducing conditions and the proteins transferred to nitrocellulose. The immunoglobulin components were detected with an anti-kappa antiserum or with a sheep antiserum which specifically recognized the protection protein followed by an appropriate alkaline phosphatase labeled 2° antibody.

iv) Expression of Antigen-Specific Immunoglobulin  
Containing a Protection Protein in transgenic  
*Nicotiana tabacum*.

To demonstrate that the plants were producing  
5 antigen-specific immunoglobulin, plant extract binding to  
purified streptococcal antigen (SA) I/II, detected with  
horseradish peroxidase labeled anti-kappa chain antiserum  
was determined. The presence of a protection protein in  
the antigen-specific immunoglobulin was demonstrated by  
10 plant extract binding to purified streptococcal antigen  
I/II and streptococcal cells detected with a sheep  
antiserum immunospecific for a protection protein,  
followed by alkaline phosphatase labeled donkey anti-  
sheep antiserum. These tests for antigen-specific  
15 immunoglobulin were carried out in microtitre plates that  
were coated with purified SA I/II (2µg/ml) in TBS, or log  
phase growth *Strep. mutans* (NCTC 10449), in bicarbonate  
buffer (pH 9.8). Blocking was with 5% non-fat dry milk  
in TBS at room temperature for 2 hours. Plant leaves  
20 were homogenized in TBS with 10µg/ml leupeptin  
(Calbiochem, USA). Mouse Guy's 13 hybridoma cell culture  
supernatant (IgG) was used as a positive control. The  
supernatants were added in serial two-fold dilutions to  
the microtitre plate and incubation was at room  
25 temperature for 2 hours. After washing with TBS with  
0.05% Tween 20, bound immunoglobulin chains were detected  
with either a goat anti-mouse light chain specific  
antibody, conjugated with horseradish peroxidase (Nordic  
Pharmaceuticals, UK), or a sheep anti-SC antiserum,  
30 followed by an alkaline phosphatase labeled donkey anti-  
sheep antibody for 2 hours at room temperature.  
Detection was with 2,2'-azino-di-[3-ethyl-benzthiazolin-  
sulphonate (Boehringer, W. Germany) for HRPO conjugated

antibody or disodium p-nitrophenyl phosphate (Sigma, UK) for alkaline phosphatase conjugated antibody.

v) Localization of Immunoglobulin Components in Plants

Photomicrographs of transgenic plants expressing  
5 immunoglobulins containing protection proteins and control *Nicotiana tabacum* leaf were prepared using immunogold detection of murine alpha chain. Briefly, leaf blades were cut into 2mm x 10mm segments and fixed in 3% (w/v) paraformaldehyde, 0.5% (w/v) glutaraldehyde,  
10 5% (w/v) sucrose in 100mM sodium phosphate (pH 7.4). After dehydration in anhydrous ethanol, leaf segments were infiltrated with xylene, embedded in paraffin and cut into 3mm sections and mounted on glass slides for immunochemical staining. The leaf sections were  
15 incubated with primary antibodies, affinity purified rabbit anti-mouse alpha chain (which reacts with the A/G hybrid heavy chain) or sheep anti-rabbit SC, and then with secondary antibody; goat anti-rabbit-10nm gold or rabbit anti-sheep-10nm gold. The immunogold signal was  
20 intensified by silver enhancement. The plants were visualized using both Phase contrast and bright field microscopy on the same leaf cross section. Immunolocalization of the protection protein on serial sections was used to show the same cellular localization for heavy  
25 chain as immunoglobulin. The analysis was carried out on the following cells and cell compartments: spongy mesophyll cells, epidermal cells, intercellular spaces, palisade parenchyma cells, and vascular bundles.

Further analysis of the exact localization of  
30 immunoglobulin components was carried out by analyzing serial sections of *Nicotiana tabacum* vascular bundle and control *Nicotiana tabacum* vascular bundle with immunogold detection for each of the components of the

immunoglobulin. Serial sections of a transgenic plant leaves from plants expressing secretory immunoglobulin were incubated with an antibody that specifically recognizes the protection protein or with anti-IgA antibody followed by the appropriate gold-labeled secondary antibody. A control leaf section from a transgenic plant that did not contain any immunoglobulin coding sequences was also incubated with anti-IgA antibody, followed by gold-labeled goat anti-rabbit antiserum, or with the gold-labeled secondary antibodies alone and confirmed the specificity of staining. Both Phase contrast illumination of a minor vascular bundle and Bright field illumination of the same field were used to show immunogold localization of the protection protein. Bright field illumination of a serial leaf cross section of the vascular bundle demonstrated the same immunogold localization of the immunoglobulin heavy chain as was shown for the protection protein.

20 Example 8.        Production of a Useful Plant Extract  
                         Containing Immunoglobulins Having a  
                         Protection Protein

Plant pieces (either leaf, stem, flower, root, or combinations) from plants producing immunoglobulins containing a protection protein were mixed with homogenization buffer (2 milliliter buffer per gram of plant material; homogenization buffer: 150 mM NaCl, 20 mM Tris-Cl, pH 7.5), homogenized into a pulp using a Waring blender and centrifuged at 10,000 X g to remove debris. The supernatant was then extracted with an equal volume of HPLC-grade ethyl acetate by shaking at room temperature, followed by centrifugation at 10,000 X g. The aqueous phase was transferred to another container,

remaining ethyl acetate was removed from the aqueous phase by placing the solution under vacuum. The resulting crude extract consistently contained 100  $\mu$ g immunoglobulin having a protection protein per ml. This method is useful for any plant containing an immunoglobulin having a protection protein.

A number of methods for homogenization have been used including a mortar and pestle or a Polytron and can be performed either in the cold or at room temperature.

The extract may be further purified by delipidation, by extraction with hexane or other organic solvents. Delipidation is not essential for deriving a useful product from the plant extract but is advantageous in cases where the final product is a purified immunoglobulin having a protection protein. In many instances the crude extract will contain a sufficiently high quantity of immunoglobulin having a protection protein (i.e. 100  $\mu$ g/mL) to be useful without any further purification or enrichment. For an oral application, the extract would be mixed with commonly used flavorings and stabilizers. For a dental application, the extract would in addition be mixed with a gelling reagent to maintain contact of the extract with teeth. For a gastric application, the flavored extract could be swallowed directly.

Example 9. Stability of an Immunoglobulin Containing a Protection Protein.

Two sets of crude plant extracts were prepared as described above. The first extract was derived from a plant expressing an IgG1 antibody and the second extract was derived from a plant expressing an immunoglobulin containing a protection protein. Crude plant extracts of

this type from plants are known to contain a variety of proteolytic enzymes. Prolonged incubation of extracts at room temperature or at 37° C therefore constitutes a proteolytic digestion.

5        Using ELISA the quantity of gamma-kappa complexes in the two extracts was determined as a function of time at both room temperature and 37° C. In these assays, an anti-kappa chain antibody was used to coat the plate followed by incubation with the plant extract at 37° C for  
10 1 hour. An anti-gamma chain antibody conjugated to HRPO was used for detection of immunoglobulin derived from the plant. The quantity of immunoglobulin having a protection protein contained in the extract immediately after the extract was prepared was taken to be 100%.  
15 After 3 hours at room temperature, the IgG1 contained 40% and the immunoglobulin containing the protection protein contained >95%. After 6 hours, the remaining IgG1 antibody was 20% and the immunoglobulin containing the protection protein abundance was still >95%. After 12  
20 hours, there was no detectable IgG1 whereas ~90% of the immunoglobulin containing the protection protein remained. A significant decrease (to ~70%) in the abundance of protected antibody was not observed until 48 hours after the extract was prepared.

25

Example 10.        Eukaryotic Tetra-transgenic Cells  
                      Expressing Immunoglobulins Containing  
                      Protection Protein.

30        The four chains comprising the immunoglobulin containing a protection protein can also be expressed in other cell types either in *in vitro* (cell cultures) or *in vivo* (transgenic animals). See, Manipulating the Mouse Embryo; A Laboratory Manual, B. Hogan et al., Cold Spring

Harbor Laboratory (1986). In the case of transgenic animals, purified preparations of appropriate vector DNAs are adjusted to a final concentration of 2 ng/ $\mu$ l in 10 mM Tris, 0.2 mM EDTA, pH 7.4. Pronuclear injections are performed using zygotes prepared from inbred animals. Injected eggs are then transferred to pseudopregnant females using standard techniques. Live born animals are then screened for the presence of transgenes using any of a number of commonly used techniques such as PCR and ELISA. Members of the pedigree expressing different components of the immunoglobulin containing the protection protein are then mated to produce multi-transgene animals. Progeny from these crosses are then screened to identify those that express all four chains. Depending on the type of vector used for zygotic injections various cell types can be identified in the transgenic animals which assemble the complete immunoglobulin containing a protection protein. These vector DNAs can consist of specific promoter elements which allow transcription of the transgene in particular cell types or tissues. Each vector could express a single component of the protected antibody (IgG/A, J chain, protection protein, or kappa chain) or could potentially express more than one component. In this instance, the vector would contain an appropriate number of promoter regions and restriction sites to allow for transcription of each transgene.

Expression of all four chains in a cell culture system can be achieved using a DNA vector from which each component can be individually promoted. This would require four expression cassettes (containing promoter, multiple cloning site, and polyadenylation region) on the same vector DNA. Alternatively, individual cell lines

can be sequentially transfected with individual vectors expressing single chains so long as each vector confers a selective resistance onto the cell line.

Commonly available vectors, such as pMAMneo  
5 (Clontech) can be adapted either for multiple expression or as a series of vectors expressing distinct selectable markers.

Transfection of any eukaryotic cells, such as fibroblasts, is done by conventional techniques.  
10 Briefly, cells are split 1:20 the day before transfection and are transfected at approximately 30% confluency using 125 mM CaCl<sub>2</sub>, 140 mM NaCl, 25 mM Hepes, 0.75 mM NaHPO<sub>4</sub>, pH 7.05, and 5 µg DNA / 10 cm dish. After 16 hours of DNA incubation, cells are shocked by 10% dimethyl  
15 sulfoxide for 3 minutes. Forty eight hours after transfection, cells are subjected to selection by growth in the appropriate medium containing an antibiotic or other cytotoxic reagent.

The resulting cells produce all the components for  
20 the immunoglobulin containing the protection protein. These components are properly assembled to produce a functional immunoglobulin containing a protection protein.

25 Example 11.     Engineering A Protection Protein Fused to A Portion of the Cytoplasmic Domain of the Rabbit Polyimmunoglobulin Receptor.

The construction of DNA segments encoding a protection protein fused to a segment encoding a segment  
30 of the cytoplasmic domain of the rabbit polyimmuno-globulin receptor is produced as follows. Protection protein cDNA encoding from the first amino acid of the signal sequence (MET<sub>-18</sub>) to GLU<sub>606</sub> is ligated into any



plant expression vector, such as the pMON530 vector (digested with Bgl II and Xho I) as a Bgl II - Xho I fragment. This protection protein derivative is obtained by PCR amplification using the appropriate oligo-  
5 nucleotide primers containing either a Bgl II or Xho I recognition sequence which are also complementary to DNA encoding residues -18 to -13 and residues 601 to 606 of the rabbit polyimmunoglobulin receptor respectively. The same procedure is performed in order to obtain a  
10 protection protein cDNA encoding from MET<sub>-18</sub> to ALA<sub>628</sub> except that the oligonucleotide containing an Xho site is also complementary to the protection protein cDNA encoding residues 623 to 628.

The cDNA encoding the rabbit polyimmunoglobulin  
15 receptor cytoplasmic domain fragment is obtained, also by PCR amplification, as a Xho I fragment. The oligonucleotides employed are complementary to DNA encoding from ARG<sub>653</sub> to ALA<sub>755</sub> both containing Xho I recognition sequences. This fragment is then ligated  
20 into the pMON530 vectors which contain the either of the protection protein cDNAs described above. The appropriate orientation of the cytoplasmic domain cDNA is determined by restriction digestions and by sequence analysis of plasmids obtained from transformed bacterial  
25 colonies.

The oligonucleotides employed for PCR amplification contain the appropriate number of nucleotides to ensure that the resulting cDNAs are in frame and capable of being translated as a continuous fusion protein  
30 containing both protection protein and cytoplasmic domain.

The resulting constructs in the appropriate orientation encode a protection protein fused directly to

the polyimmunoglobulin receptor cytoplasmic domain with no functional transmembrane segment, operably linked to a DNA segment (promoter) enabling expression in a plant cell. The constructs encode two additional amino acids  
5 (SER - TRP) which are derived from introduction of the Xho I restriction site and which serve as a linker between the protection protein and the cytoplasmic domain.

These vectors are then used to transform  
10 *Agrobacterium* as previously described which in turn is used to transform plant cells. The same techniques described in the above Examples are used to produce a plant expressing this protein as part of an immunoglobulin.

## SEQUENCE LISTING

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(D) SOFTWARE: Word Perfect 5.1

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Docket No. 212/127

## (viii) ATTORNEY/AGENT INFORMATION:

5 (A) NAME: Guise, Jeffrey W.  
(B) REGISTRATION NUMBER: 34,613  
(C) REFERENCE/DOCKET NUMBER: 242/238

## (ix) TELECOMMUNICATION INFORMATION:

10 (A) TELEPHONE: (619) 552-8400  
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(C) TELEX: 67-3510

## SEQUENCE LISTING

(2) INFORMATION FOR SEQ ID NO: 1:

5 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3517 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear  
 DESCRIPTION: Rabbit polyimmunoglobulin receptor

(ix) FEATURE:

15 (A) NAME/KEY: Coding Sequence  
 (B) LOCATION: 124...2445

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

20 GGCCGGGGTT ACGGGCTGGC CAGCAGGCTG TGCCCCGAG TCCGGTCAGCAGGAGGGGAA 60  
 GAAGTGGCCT AAAATCTCTC CCGCATCGGC AGCCCAGGCC TAGTGCCCTA CCAGCCACCA 120  
 GCC ATG GCT CTC TTC TTG CTC ACC TGC CTG CTG GCT GTC TTT TCA GCG 168  
 25 Met Ala Leu Phe Leu Leu Thr Cys Leu Leu Ala Val Phe Ser Ala  
 1 5 10 15  
 GCC ACG GCA CAA AGC TCC TTA TTG GGT CCC AGC TCC ATA TTT GGT CCC 216  
 30 Ala Thr Ala Gln Ser Ser Leu Leu Gly Pro Ser Ser Ile Phe Gly Pro  
 20 25 30  
 GGG GAG GTG AAT GTT TTG GAA GGC GAC TCG GTG TCC ATC ACA TGC TAC 264  
 Gly Glu Val Asn Val Leu Glu Gly Asp Ser Val Ser Ile Thr Cys Tyr  
 35 35 40 45  
 TAC CCA ACA ACC TCC GTC ACC CGG CAC AGC CGG AAG TTC TGG TGC CGG 312  
 Tyr Pro Thr Thr Ser Val Thr Arg His Ser Arg Lys Phe Trp Cys Arg  
 50 55 60  
 GAA GAG GAG AGC GGC CGC TGC GTG ACG CTT GCC TCG ACC GGC TAC ACG 360  
 Glu Glu Glu Ser Gly Arg Cys Val Thr Leu Ala Ser Thr Gly Tyr Thr  
 65 70 75  
 TCC CAG GAA TAC TCC GGG AGA GGC AAG CTC ACC GAC TTC CCT GAT AAA 408  
 45 Ser Gln Glu Tyr Ser Gly Arg Gly Lys Leu Thr Asp Phe Pro Asp Lys  
 80 85 90 95  
 GGG GAG TTT GTG GTG ACT GTT GAC CAA CTC ACC CAG AAC GAC TCA GGG 456  
 Gly Glu Phe Val Val Thr Val Asp Gln Leu Thr Gln Asn Asp Ser Gly  
 50 100 105 110  
 AGC TAC AAG TGT GGC GTG GGA GTC AAC GGC CGT GGC CTG GAC TTC GGT 504  
 Ser Tyr Lys Cys Gly Val Gly Val Asn Gly Arg Gly Leu Asp Phe Gly  
 115 120 125  
 GTC AAC GTG CTG GTC AGC CAG AAG CCA GAG CCT GAT GAC GTT GTT TAC 552  
 55 Val Asn Val Leu Val Ser Gln Lys Pro Glu Pro Asp Asp Val Val Tyr  
 130 135 140

	AAA CAA TAT GAG AGT TAT ACA GTA ACC ATC ACC TGC CCT TTC ACA TAT	600
	Lys Gln Tyr Glu Ser Tyr Thr Val Thr Ile Thr Cys Pro Phe Thr Tyr	
	145 150 155	
5	GCG ACT AGG CAA CTA AAG AAG TCC TTT TAC AAG GTG GAA GAC GGG GAA	648
	Ala Thr Arg Gln Leu Lys Lys Ser Phe Tyr Lys Val Glu Asp Gly Glu	
	160 165 170 175	
10	CTT GTA CTC ATC ATT GAT TCC AGC AGT AAG GAG GCA AAG GAC CCC AGG	696
	Leu Val Leu Ile Ile Asp Ser Ser Ser Lys Glu Ala Lys Asp Pro Arg	
	180 185 190	
15	TAT AAG GGC AGA ATA ACG TTG CAG ATC CAA AGT ACC ACA GCA AAA GAA	744
	Tyr Lys Gly Arg Ile Thr Leu Gln Ile Gln Ser Thr Thr Ala Lys Glu	
	195 200 205	
20	TTC ACA GTC ACC ATC AAG CAT TTG CAG CTC AAT GAT GCT GGG CAG TAT	792
	Phe Thr Val Thr Ile Lys His Leu Gln Leu Asn Asp Ala Gly Gln Tyr	
	210 215 220	
25	GTC TGC CAG AGT GGA AGC GAC CCC ACT GCT GAA GAA CAG AAC GTT GAC	840
	Val Cys Gln Ser Gly Ser Asp Pro Thr Ala Glu Glu Gln Asn Val Asp	
	225 230 235	
30	CTC CGA CTG CTA ACT CCT GGT CTG CTC TAT GGA AAC CTG GGG GGC TCG	888
	Leu Arg Leu Leu Thr Pro Gly Leu Leu Tyr Gly Asn Leu Gly Gly Ser	
	240 245 250 255	
35	GTG ACC TTT GAA TGT GCC CTG GAC TCT GAA GAC GCA AAC GCG GTA GCA	936
	Val Thr Phe Glu Cys Ala Leu Asp Ser Glu Asp Ala Asn Ala Val Ala	
	260 265 270	
40	TCC TTG CGC CAG GTT AGG GGT GGC AAT GTG GTC ATT GAC AGC CAG GGG	984
	Ser Leu Arg Gln Val Arg Gly Gly Asn Val Val Ile Asp Ser Gln Gly	
	275 280 285	
45	ACA ATA GAT CCA GCC TTC GAG GGC AGG ATC CTG TTC ACC AAG GCT GAG	1032
	Thr Ile Asp Pro Ala Phe Glu Gly Arg Ile Leu Phe Thr Lys Ala Glu	
	290 295 300	
50	AAC GGC CAC TTC AGT GTA GTG ATC GCA GGC CTG AGG AAG GAA GAC ACA	1080
	Asn Gly His Phe Ser Val Val Ile Ala Gly Leu Arg Lys Glu Asp Thr	
	305 310 315	
55	GGG AAC TAT CTG TGC GGA GTC CAG TCC AAT GGT CAG TCT GGG GAT GGG	1128
	Gly Asn Tyr Leu Cys Gly Val Gln Ser Asn Gly Gln Ser Gly Asp Gly	
	320 325 330 335	
60	CCC ACC CAG CTT CGG CAA CTC TTC GTC AAT GAA GAG ATC GAC GTG TCC	1176
	Pro Thr Gln Leu Arg Gln Leu Phe Val Asn Glu Glu Ile Asp Val Ser	
	340 345 350	
65	CGC AGC CCC CCT GTG TTG AAG GGC TTT CCA GGA GGC TCC GTG ACC ATA	1224
	Arg Ser Pro Pro Val Leu Lys Gly Phe Pro Gly Gly Ser Val Thr Ile	
	355 360 365	
70	CGC TGC CCC TAC AAC CCG AAG AGA AGC GAC AGC CAC CTG CAG CTG TAT	1272
	Arg Cys Pro Tyr Asn Pro Lys Arg Ser Asp Ser His Leu Gln Leu Tyr	
	370 375 380	

5	CTC TGG GAA GGG AGT CAA ACC CGC CAT CTG CTG GTG GAC AGC GGC GAG	1320
	Leu Trp Glu Gly Ser Gln Thr Arg His Leu Leu Val Asp Ser Gly Glu	
	385 390 395	
10	GGG CTG GTT CAG AAA GAC TAC ACA GGC AGG CTG GCC CTG TTC GAA GAG	1368
	Gly Leu Val Gln Lys Asp Tyr Thr Gly Arg Leu Ala Leu Phe Glu Glu	
	400 405 410 415	
15	CCT GGC AAT GGC ACC TTC TCA GTC GTC CTC AAC CAG CTC ACT GCC GAG	1416
	Pro Gly Asn Gly Thr Phe Ser Val Val Leu Asn Gln Leu Thr Ala Glu	
	420 425 430	
20	GAT GAA GGC TTC TAC TGG TGT GTC AGC GAT GAC GAT GAG TCC CTG ACG	1464
	Asp Glu Gly Phe Tyr Trp Cys Val Ser Asp Asp Asp Glu Ser Leu Thr	
	435 440 445	
25	ACT TCG GTG AAG CTC CAG ATC GTT GAC GGA GAA CCA AGC CCC ACG ATC	1512
	Thr Ser Val Lys Leu Gln Ile Val Asp Gly Glu Pro Ser Pro Thr Ile	
	450 455 460	
30	GAC AAG TTC ACT GCT GTG CAG GGA GAG CCT GTT GAG ATC ACC TGC CAC	1560
	Asp Lys Phe Thr Ala Val Gln Gly Glu Pro Val Glu Ile Thr Cys His	
	465 470 475	
35	TTC CCA TGC AAA TAC TTC TCC TCC GAG AAG TAC TGG TGC AAG TGG AAT	1608
	Phe Pro Cys Lys Tyr Phe Ser Ser Glu Lys Tyr Trp Cys Lys Trp Asn	
	480 485 490 495	
40	GAC CAT GGC TGC GAG GAC CTG CCC ACT AAG CTC AGC TCC AGC GGC GAC	1656
	Asp His Gly Cys Glu Asp Leu Pro Thr Lys Leu Ser Ser Ser Gly Asp	
	500 505 510	
45	CTT GTG AAA TGC AAC AAC AAC CTG GTC CTC ACC CTG ACC TTG GAC TCG	1704
	Leu Val Lys Cys Asn Asn Asn Leu Val Leu Thr Leu Thr Leu Asp Ser	
	515 520 525	
50	GTC AGC GAA GAT GAC GAG GGC TGG TAC TGG TGT GGC GCG AAA GAC GGC	1752
	Val Ser Glu Asp Asp Glu Gly Trp Tyr Trp Cys Gly Ala Lys Asp Gly	
	530 535 540	
55	CAC GAG TTT GAA GAG GTT GCG GCC GTC AGG GTG GAG CTG ACA GAG CCA	1800
	His Glu Phe Glu Glu Val Ala Ala Val Arg Val Glu Leu Thr Glu Pro	
	545 550 555	
60	GCC AAG GTA GCT GTC GAG CCA GCC AAG GTA CCT GTC GAC CCA GCC AAG	1848
	Ala Lys Val Ala Val Glu Pro Ala Lys Val Pro Val Asp Pro Ala Lys	
	560 565 570 575	
65	GCA GCC CCC GCG CCT GCT GAG GAG AAG GCC AAG GCG CGG TGC CCA GTG	1896
	Ala Ala Pro Ala Pro Ala Glu Glu Lys Ala Lys Ala Arg Cys Pro Val	
	580 585 590	
70	CCC AGG AGA AGG CAG TGG TAC CCA TTG TCA AGG AAG CTG AGA ACA AGT	1944
	Pro Arg Arg Arg Gln Trp Tyr Pro Leu Ser Arg Lys Leu Arg Thr Ser	
	595 600 605	

	TGT CCA GAA CCT CGG CTC CTT GCG GAG GAG GTA GCA GTG CAG AGT GCG	1992
	Cys Pro Glu Pro Arg Leu Leu Ala Glu Glu Val Ala Val Gln Ser Ala	
	610 615 620	
5	GAA GAC CCA GCC AGT GGG AGC AGA GCG TCT GTG GAT GCC AGC AGT GCT	2040
	Glu Asp Pro Ala Ser Gly Ser Arg Ala Ser Val Asp Ala Ser Ser Ala	
	625 630 635	
10	TCG GGA CAA AGC GGG AGT GCC AAA GTA CTG ATC TCC ACC CTG GTG CCC	2088
	Ser Gly Gln Ser Gly Ser Ala Lys Val Leu Ile Ser Thr Leu Val Pro	
	640 645 650 655	
15	TTG GGG CTG GTG CTG GCA GCG GGG GCC ATG GCC GTG GCC ATA GCC AGA	2136
	Leu Gly Leu Val Leu Ala Ala Gly Ala Met Ala Val Ala Ile Ala Arg	
	660 665 670	
20	GCC CGG CAC AGG AGG AAC GTG GAC CGA GTT TCC ATC GGA AGC TAC AGG	2184
	Ala Arg His Arg Arg Asn Val Asp Arg Val Ser Ile Gly Ser Tyr Arg	
	675 680 685	
25	ACA GAC ATT AGC ATG TCA GAC TTG GAG AAC TCC AGG GAG TTC GGA GCC	2232
	Thr Asp Ile Ser Met Ser Asp Leu Glu Asn Ser Arg Glu Phe Gly Ala	
	690 695 700	
30	GGA AAG GAT GAG TTA GCG ACG GCC ACC GAG AGC ACC GTG GAG ATT GAG	2328
	Gly Lys Asp Glu Leu Ala Thr Ala Thr Glu Ser Thr Val Glu Ile Glu	
	720 725 730 735	
35	GAG CCC AAG AAG GCA AAA CGG TCA TCC AAG GAA GAA GCC GAC CTG GCC	2376
	Glu Pro Lys Lys Ala Lys Arg Ser Ser Lys Glu Glu Ala Asp Leu Ala	
	740 745 750	
40	TAC TCA GCT TTC CTG CTC CAA TCC AAC ACC ATA GCT GCT GAG CAC CAA	2424
	Tyr Ser Ala Phe Leu Leu Gln Ser Asn Thr Ile Ala Ala Glu His Gln	
	755 760 765	
45	GAT GGC CCC AAG GAG GCC TAG GCACAGCCGG CCACCGCCGC CGCCGCCACC GCCGC	2480
	Asp Gly Pro Lys Glu Ala	
	770	
50	CGCCGCCGCC ACCTGTGAAA ATCACCTTCC AGAATCACGT TGATCCTCGG GGTCCCCAGA	2540
	GCCGGGGGGCT CAACCGCCCT GCACCCCCCA TGTCCTCCACC ACCTAAACTT CCCTACCTGT	2600
	GCCCAGAGGT GTGCTGGTCC CCTCCTCCAC GGCATCCAGG CCTGGCTCAA TGTTCCCGTT	2660
	GGGGTGGGGG TGTGAGGGGT TCCTACTTGC AGCCCGGTTC TCCGAGAGA AGCTAAGGAT	2720
	CCAGGTCTTG AGGGAGGGGC CTCTCGAAGG CAGACAGACC AGAGAGGGGG GAGGAGCCCT	2780
55	TGGATGGGAG GCCAGAGGCG CTTTCCGGCC ACCCCCTCCC TCCCTGCCCC CACCCTCCTT	2840
	CCTTCATTCA AAAGTCCCAG TGGCTGCTGC CTAGGGTCCA GGCGCTGGCC GCACGCCTCC	2900
	TCGAAGCCGT TGTGCAAACA TCACTGGAGG AAGCCAGGGC TCCTCCCGGG CTGTGTATCC	2960



TCACTCAGGC ATCCTGTCCT CCCAGTATC AGGAGATGTC AAGCGTCTGA AGGCTGTGTG 3020  
 CCCTGGGCGT GTCTGCAAGT CACCCCAGAC ACATGTTCTC GCCATTTTAC AGATGAGAAC 3080  
 5 ACTGAGGTTG TACTCAAGGG CACCCTGCGA GATGGAGCAA CAGCAAATA GATGGGCTTC 3140  
 TGCTGTCTC TTGGCCAGAG GTCTCTCCAC AGGAGCCCCT GCCCTGTAG GAAGCAGAGT 3200  
 10 TTTAGAACAT GGAAGAAGAA GAGGGGGATG GCCCTGGACG CTGACCTCTC CCAAGCCCCC 3260  
 ACGGGGAAAA AGGCCCCCTC CTTTCTGTG ACTCTCGGG ACCTGCGGAG TTGAGCATTC 3320  
 15 GTGCCCCGTG TGTCTGAAGA GTTCCCAGTG GAAAGAAGAA AAGAGGGTGT TTGTCAGTGC 3380  
 CGGGGAGGGC CTGATCCCCA GACAGCTGAA GTTTAAGGTC CTTGTCCCTG TGAGCTTTAA 3440  
 CCAGCACCTC CGGGCTGACC CTTGCTAACA CATCAGAAAT GTGATTTAAT CATTAAACAT 3500  
 20 TGTGATTGCC ACTGGGA 3517

25 (2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

30 (A) LENGTH: 773 amino acids  
 (B) TYPE: amino acid  
 (C) STRANDNESS: single  
 (D) TOPOLOGY: linear  
 DESCRIPTION: Rabbit polyimmunoglobulin receptor

35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met Ala Leu Phe Leu Leu Thr Cys Leu Leu Ala Val Phe Ser Ala Ala  
 1 5 10 15  
 40 Thr Ala Gln Ser Ser Leu Leu Gly Pro Ser Ser Ile Phe Gly Pro Gly  
 20 25 30  
 Glu Val Asn Val Leu Glu Gly Asp Ser Val Ser Ile Thr Cys Tyr Tyr  
 35 40 45  
 45 Pro Thr Thr Ser Val Thr Arg His Ser Arg Lys Phe Trp Cys Arg Glu  
 50 55 60  
 Glu Glu Ser Gly Arg Cys Val Thr Leu Ala Ser Thr Gly Tyr Thr Ser  
 50 65 70 75 80  
 Gln Glu Tyr Ser Gly Arg Gly Lys Leu Thr Asp Phe Pro Asp Lys Gly  
 85 90 95  
 55 Glu Phe Val Val Thr Val Asp Gln Leu Thr Gln Asn Asp Ser Gly Ser  
 100 105 110  
 Tyr Lys Cys Gly Val Gly Val Asn Gly Arg Gly Leu Asp Phe Gly Val  
 115 120 125

	Asn	Val	Leu	Val	Ser	Gln	Lys	Pro	Glu	Pro	Asp	Asp	Val	Val	Tyr	Lys
	130						135					140				
5	Gln	Tyr	Glu	Ser	Tyr	Thr	Val	Thr	Ile	Thr	Cys	Pro	Phe	Thr	Tyr	Ala
	145					150					155					160
	Thr	Arg	Gln	Leu	Lys	Lys	Ser	Phe	Tyr	Lys	Val	Glu	Asp	Gly	Glu	Leu
					165					170					175	
10	Val	Leu	Ile	Ile	Asp	Ser	Ser	Ser	Lys	Glu	Ala	Lys	Asp	Pro	Arg	Tyr
				180					185					190		
	Lys	Gly	Arg	Ile	Thr	Leu	Gln	Ile	Gln	Ser	Thr	Thr	Ala	Lys	Glu	Phe
15			195					200					205			
	Thr	Val	Thr	Ile	Lys	His	Leu	Gln	Leu	Asn	Asp	Ala	Gly	Gln	Tyr	Val
	210						215					220				
20	Cys	Gln	Ser	Gly	Ser	Asp	Pro	Thr	Ala	Glu	Glu	Gln	Asn	Val	Asp	Leu
	225					230					235					240
	Arg	Leu	Leu	Thr	Pro	Gly	Leu	Leu	Tyr	Gly	Asn	Leu	Gly	Gly	Ser	Val
					245					250					255	
25	Thr	Phe	Glu	Cys	Ala	Leu	Asp	Ser	Glu	Asp	Ala	Asn	Ala	Val	Ala	Ser
				260					265					270		
	Leu	Arg	Gln	Val	Arg	Gly	Gly	Asn	Val	Val	Ile	Asp	Ser	Gln	Gly	Thr
30			275					280					285			
	Ile	Asp	Pro	Ala	Phe	Glu	Gly	Arg	Ile	Leu	Phe	Thr	Lys	Ala	Glu	Asn
	290						295					300				
35	Gly	His	Phe	Ser	Val	Val	Ile	Ala	Gly	Leu	Arg	Lys	Glu	Asp	Thr	Gly
	305					310					315					320
	Asn	Tyr	Leu	Cys	Gly	Val	Gln	Ser	Asn	Gly	Gln	Ser	Gly	Asp	Gly	Pro
					325					330					335	
40	Thr	Gln	Leu	Arg	Gln	Leu	Phe	Val	Asn	Glu	Glu	Ile	Asp	Val	Ser	Arg
				340					345					350		
	Ser	Pro	Pro	Val	Leu	Lys	Gly	Phe	Pro	Gly	Gly	Ser	Val	Thr	Ile	Arg
45			355					360					365			
	Cys	Pro	Tyr	Asn	Pro	Lys	Arg	Ser	Asp	Ser	His	Leu	Gln	Leu	Tyr	Leu
	370						375					380				
50	Trp	Glu	Gly	Ser	Gln	Thr	Arg	His	Leu	Leu	Val	Asp	Ser	Gly	Glu	Gly
	385					390					395					400
	Leu	Val	Gln	Lys	Asp	Tyr	Thr	Gly	Arg	Leu	Ala	Leu	Phe	Glu	Glu	Pro
				405						410					415	
55	Gly	Asn	Gly	Thr	Phe	Ser	Val	Val	Leu	Asn	Gln	Leu	Thr	Ala	Glu	Asp
				420					425					430		

Glu Gly Phe Tyr Trp Cys Val Ser Asp Asp Asp Glu Ser Leu Thr Thr  
                   435                                  440                                  445  
 5 Ser Val Lys Leu Gln Ile Val Asp Gly Glu Pro Ser Pro Thr Ile Asp  
                   450                                  455                                  460  
 Lys Phe Thr Ala Val Gln Gly Glu Pro Val Glu Ile Thr Cys His Phe  
                   465                                  470                                  475                                  480  
 10 Pro Cys Lys Tyr Phe Ser Ser Glu Lys Tyr Trp Cys Lys Trp Asn Asp  
                                   485                                  490                                  495  
 His Gly Cys Glu Asp Leu Pro Thr Lys Leu Ser Ser Ser Gly Asp Leu  
                                   500                                  505                                  510  
 15 Val Lys Cys Asn Asn Asn Leu Val Leu Thr Leu Thr Leu Asp Ser Val  
                                   515                                  520                                  525  
 20 Ser Glu Asp Asp Glu Gly Trp Tyr Trp Cys Gly Ala Lys Asp Gly His  
                   530                                  535                                  540  
 Glu Phe Glu Glu Val Ala Ala Val Arg Val Glu Leu Thr Glu Pro Ala  
                   545                                  550                                  555                                  560  
 25 Lys Val Ala Val Glu Pro Ala Lys Val Pro Val Asp Pro Ala Lys Ala  
                                   565                                  570                                  575  
 Ala Pro Ala Pro Ala Glu Glu Lys Ala Lys Ala Arg Cys Pro Val Pro  
                                   580                                  585                                  590  
 30 Arg Arg Arg Gln Trp Tyr Pro Leu Ser Arg Lys Leu Arg Thr Ser Cys  
                                   595                                  600                                  605  
 35 Pro Glu Pro Arg Leu Leu Ala Glu Glu Val Ala Val Gln Ser Ala Glu  
                   610                                  615                                  620  
 Asp Pro Ala Ser Gly Ser Arg Ala Ser Val Asp Ala Ser Ser Ala Ser  
                   625                                  630                                  635                                  640  
 40 Gly Gln Ser Gly Ser Ala Lys Val Leu Ile Ser Thr Leu Val Pro Leu  
                                   645                                  650                                  655  
 Gly Leu Val Leu Ala Ala Gly Ala Met Ala Val Ala Ile Ala Arg Ala  
                                   660                                  665                                  670  
 45 Arg His Arg Arg Asn Val Asp Arg Val Ser Ile Gly Ser Tyr Arg Thr  
                                   675                                  680                                  685  
 50 Asp Ile Ser Met Ser Asp Leu Glu Asn Ser Arg Glu Phe Gly Ala Ile  
                   690                                  695                                  700  
 Asp Asn Pro Ser Ala Cys Pro Asp Ala Arg Glu Thr Ala Leu Gly Gly  
                   705                                  710                                  715                                  720  
 55 Lys Asp Glu Leu Ala Thr Ala Thr Glu Ser Thr Val Glu Ile Glu Glu  
                                   725                                  730                                  735  
 Pro Lys Lys Ala Lys Arg Ser Ser Lys Glu Glu Ala Asp Leu Ala Tyr  
                   740                                  745                                  750

Ser Ala Phe Leu Leu Gln Ser Asn Thr Ile Ala Ala Glu His Gln Asp  
755 760 765

5 Gly Pro Lys Glu Ala  
770

10

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

15

(A) LENGTH: 2919 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear  
DESCRIPTION: Human polyimmunoglobulin Receptor

20

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence  
(B) LOCATION: 235....2472

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

AGAGTTTCAG TTTTGGCAGC AGCGTCCAGT GCCCTGCCAG TAGCTCCTAG AGAGGCAGGG 60  
30 GTTACCAACT GGCCAGCAGG CTGTGTCCCT GAAGTCAGAT CAACGGGAGA GAAGGAAGTG 120  
GCTAAAACAT TGCACAGGAG AAGTCGGCCT GAGTGGTGCG GCGCTCGGGA CCCACCAGCA 180  
ATGCTGCTCT TCGTGCTCAC CTGCCTGCTG GCGGTCTTCC CAGCCATCTC CACG AAG 237  
35 Lys  
1  
AGT CCC ATA TTT GGT CCC GAG GAG GTG AAT AGT GTG GAA GGT AAC TCA 285  
40 Ser Pro Ile Phe Gly Pro Glu Glu Val Asn Ser Val Glu Gly Asn Ser  
5 10 15  
GTG TCC ATC ACG TGC TAC TAC CCA CCC ACC TCT GTC AAC CGG CAC ACC 333  
Val Ser Ile Thr Cys Tyr Tyr Pro Pro Thr Ser Val Asn Arg His Thr  
20 25 30  
45 CGG AAG TAC TGG TGC CGG CAG GGA GCT AGA GGT GGC TGC ATA ACC CTC 381  
Arg Lys Tyr Trp Cys Arg Gln Gly Ala Arg Gly Gly Cys Ile Thr Leu  
35 40 45  
50 ATC TCC TCG GAG GGC TAC GTC TCC AGC AAA TAT GCA GGC AGG GCT AAC 429  
Ile Ser Ser Glu Gly Tyr Val Ser Ser Lys Tyr Ala Gly Arg Ala Asn  
50 55 60 65  
55 CTC ACC AAC TTC CCG GAG AAC GGC ACA TTT GTG GTG AAC ATT GCC CAG 477  
Leu Thr Asn Phe Pro Glu Asn Gly Thr Phe Val Val Asn Ile Ala Gln  
70 75 80

	CTG AGC CAG GAT GAC TCC GGG CGC TAC AAG TGT GGC CTG GGC ATC AAT	525
	Leu Ser Gln Asp Asp Ser Gly Arg Tyr Lys Cys Gly Leu Gly Ile Asn	
	85 90 95	
5	AGC CGA GGC CTG TCC TTT GAT GTC AGC CTG GAG GTC AGC CAG GGT CCT	573
	Ser Arg Gly Leu Ser Phe Asp Val Ser Leu Glu Val Ser Gln Gly Pro	
	100 105 110	
10	GGG CTC CTA AAT GAC ACT AAA GTC TAC ACA GTG GAC CTG GGC AGA ACG	621
	Gly Leu Leu Asn Asp Thr Lys Val Tyr Thr Val Asp Leu Gly Arg Thr	
	115 120 125	
15	GTG ACC ATC AAC TGC CCT TTC AAG ACT GAG AAT GCT CAA AAG AGG AAG	669
	Val Thr Ile Asn Cys Pro Phe Lys Thr Glu Asn Ala Gln Lys Arg Lys	
	130 135 140 145	
20	TCC TTG TAC AAG CAG ATA GGC CTG TAC CCT GTG CTG GTC ATC GAC TCC	717
	Ser Leu Tyr Lys Gln Ile Gly Leu Tyr Pro Val Leu Val Ile Asp Ser	
	150 155 160	
	AGT GGT TAT GTG AAT CCC AAC TAT ACA GGA AGA ATA CGC CTT GAT ATT	765
	Ser Gly Tyr Val Asn Pro Asn Tyr Thr Gly Arg Ile Arg Leu Asp Ile	
	165 170 175	
25	CAG GGT ACT GGC CAG TTA CTG TTC AGC GTT GTC ATC AAC CAA CTC AGG	813
	Gln Gly Thr Gly Gln Leu Leu Phe Ser Val Val Ile Asn Gln Leu Arg	
	180 185 190	
30	CTC AGC GAT GCT GGG CAG TAT CTC TGC CAG GCT GGG GAT GAT TCC AAT	861
	Leu Ser Asp Ala Gly Gln Tyr Leu Cys Gln Ala Gly Asp Asp Ser Asn	
	195 200 205	
35	AGT AAT AAG AAG AAT GCT GAC CTC CAA GTG CTA AAG CCC GAG CCC GAG	909
	Ser Asn Lys Lys Asn Ala Asp Leu Gln Val Leu Lys Pro Glu Pro Glu	
	210 215 220 225	
40	CTG GTT TAT GAA GAC CTG AGG GGC TCA GTG ACC TTC CAC TGT GCC CTG	957
	Leu Val Tyr Glu Asp Leu Arg Gly Ser Val Thr Phe His Cys Ala Leu	
	230 235 240	
	GGC CCT GAG GTG GCA AAC GTG GCC AAA TTT CTG TGC CGA CAG AGC AGT	1005
	Gly Pro Glu Val Ala Asn Val Ala Lys Phe Leu Cys Arg Gln Ser Ser	
	245 250 255	
45	GGG GAA AAC TGT GAC GTG GTC GTC AAC ACC CTG GGG AAG AGG GCC CCA	1053
	Gly Glu Asn Cys Asp Val Val Asn Thr Leu Gly Lys Arg Ala Pro	
	260 265 270	
50	GCC TTT GAG GGC AGG ATC CTG CTC AAC CCC CAG GAC AAG GAT GGC TCA	1101
	Ala Phe Glu Gly Arg Ile Leu Leu Asn Pro Gln Asp Lys Asp Gly Ser	
	275 280 285	
55	TTC AGT GTG GTG ATC ACA GGC CTG AGG AAG GAG GAT GCA GGG CGC TAC	1149
	Phe Ser Val Val Ile Thr Gly Leu Arg Lys Glu Asp Ala Gly Arg Tyr	
	290 295 300 305	
	CTG TGT GGA GCC CAT TCG GAT GGT CAG CTG CAG GAA GGC TCG CCT ATC	1197
	Leu Cys Gly Ala His Ser Asp Gly Gln Leu Gln Glu Gly Ser Pro Ile	
	310 315 320	

5	CAG GCC TGG CAA CTC TTC GTC AAT GAG GAG TCC ACG ATT CCC CGC AGC	1245
	Gln Ala Trp Gln Leu Phe Val Asn Glu Glu Ser Thr Ile Pro Arg Ser	
	325 330 335	
10	CCC ACT GTG GTG AAG GGG GTG GCA GGA AGC TCT GTG GCC GTG CTC TGC	1293
	Pro Thr Val Val Lys Gly Val Ala Gly Ser Ser Val Ala Val Leu Cys	
	340 345 350	
15	CCC TAC AAC CGT AAG GAA AGC AAA AGC ATC AAG TAC TGG TGT CTC TGG	1341
	Pro Tyr Asn Arg Lys Glu Ser Lys Ser Ile Lys Tyr Trp Cys Leu Trp	
	355 360 365	
20	GAA GGG GCC CAG AAT GGC CGC TGC CCC CTG CTG GTG GAC AGC GAG GGG	1389
	Glu Gly Ala Gln Asn Gly Arg Cys Pro Leu Leu Val Asp Ser Glu Gly	
	370 375 380 385	
25	TGG GTT AAG GCC CAG TAC GAG GGC CGC CTC TCC CTG CTG GAG GAG CCA	1437
	Trp Val Lys Ala Gln Tyr Glu Gly Arg Leu Ser Leu Leu Glu Glu Pro	
	390 395 400	
30	GGC AAC GGC ACC TTC ACT GTC ATC CTC AAC CAG CTC ACC AGC CGG GAC	1485
	Gly Asn Gly Thr Phe Thr Val Ile Leu Asn Gln Leu Thr Ser Arg Asp	
	405 410 415	
35	GCC GGC TTC TAC TGG TGT CTG ACC AAC GGC GAT ACT CTC TGG AGG ACC	1533
	Ala Gly Phe Tyr Trp Cys Leu Thr Asn Gly Asp Thr Leu Trp Arg Thr	
	420 425 430	
40	ACC GTG GAG ATC AAG ATT ATC GAA GGA GAA CCA AAC CTC AAG GTA CCA	1581
	Thr Val Glu Ile Lys Ile Ile Glu Gly Glu Pro Asn Leu Lys Val Pro	
	435 440 445	
45	GGG AAT GTC ACG GCT GTG CTG GGA GAG ACT CTC AAG GTC CCC TGT CAC	1629
	Gly Asn Val Thr Ala Val Leu Gly Glu Thr Leu Lys Val Pro Cys His	
	450 455 460 465	
50	TTT CCA TGC AAA TTC TCC TCG TAC GAG AAA TAC TGG TGC AAG TGG AAT	1677
	Phe Pro Cys Lys Phe Ser Ser Tyr Glu Lys Tyr Trp Cys Lys Trp Asn	
	470 475 480	
55	AAC ACG GGC TGC CAG GCC CTG CCC AGC CAA GAC GAA GGC CCC AGC AAG	1725
	Asn Thr Gly Cys Gln Ala Leu Pro Ser Gln Asp Glu Gly Pro Ser Lys	
	485 490 495	
60	GCC TTC GTG AAC TGT GAC GAG AAC AGC CGG CTT GTC TCC CTG ACC CTG	1773
	Ala Phe Val Asn Cys Asp Glu Asn Ser Arg Leu Val Ser Leu Thr Leu	
	500 505 510	
65	AAC CTG GTG ACC AGG GCT GAT GAG GGC TGG TAC TGG TGT GGA GTG AAG	1821
	Asn Leu Val Thr Arg Ala Asp Glu Gly Trp Tyr Trp Cys Gly Val Lys	
	515 520 525	
70	CAG GGC CAC TTC TAT GGA GAG ACT GCA GCC GTC TAT GTG GCA GTT GAA	1869
	Gln Gly His Phe Tyr Gly Glu Thr Ala Ala Val Tyr Val Ala Val Glu	
	530 535 540 545	

	GAG AGG AAG GCA GCG GGG TCC CGC GAT GTC AGC CTA GCG AAG GCA GAC	1917
	Glu Arg Lys Ala Ala Gly Ser Arg Asp Val Ser Leu Ala Lys Ala Asp	
	550 555 560	
5	GCT GCT CCT GAT GAG AAG GTG CTA GAC TCT GGT TTT CGG GAG ATT GAG	1965
	Ala Ala Pro Asp Glu Lys Val Leu Asp Ser Gly Phe Arg Glu Ile Glu	
	565 570 575	
10	AAC AAA GCC ATT CAG GAT CCC AGG CTT TTT GCA GAG GAA AAG GCG GTG	2013
	Asn Lys Ala Ile Gln Asp Pro Arg Leu Phe Ala Glu Glu Lys Ala Val	
	580 585 590	
15	GCA GAT ACA AGA GAT CAA GCC GAT GGG AGC AGA GCA TCT GTG GAT TCC	2061
	Ala Asp Thr Arg Asp Gln Ala Asp Gly Ser Arg Ala Ser Val Asp Ser	
	595 600 605	
20	GGC AGC TCT GAG GAA CAA GGT GGA AGC TCC AGA GCG CTG GTC TCC ACC	2109
	Gly Ser Ser Glu Glu Gln Gly Gly Ser Ser Arg Ala Leu Val Ser Thr	
	610 615 620 625	
25	CTG GTG CCC CTG GGC CTG GTG CTG GCA GTG GGA GCC GTG GCT GTG GGG	2157
	Leu Val Pro Leu Gly Leu Val Leu Ala Val Gly Ala Val Ala Val Gly	
	630 635 640	
30	GTG GCC AGA GCC CGG CAC AGG AAG AAC GTC GAC CGA GTT TCA ATC AGA	2205
	Val Ala Arg Ala Arg His Arg Lys Asn Val Asp Arg Val Ser Ile Arg	
	645 650 655	
35	AGC TAC AGG ACA GAC ATT AGC ATG TCA GAC TTC GAG AAC TCC AGG GAA	2253
	Ser Tyr Arg Thr Asp Ile Ser Met Ser Asp Phe Glu Asn Ser Arg Glu	
	660 665 670	
40	TTT GGA GCC AAT GAC AAC ATG GGA GCC TCT TCG ATC ACT CAG GAG ACA	2301
	Phe Gly Ala Asn Asp Asn Met Gly Ala Ser Ser Ile Thr Gln Glu Thr	
	675 680 685	
45	TCC CTC GGA GGA AAA GAA GAG TTT GTT GCC ACC ACT GAG AGC ACC ACA	2349
	Ser Leu Gly Gly Lys Glu Glu Phe Val Ala Thr Thr Glu Ser Thr Thr	
	690 695 700 705	
50	GAG ACC AAA GAA CCC AAG AAG GCA AAA AGG TCA TCC AAG GAG GAA GCC	2397
	Glu Thr Lys Glu Pro Lys Lys Ala Lys Arg Ser Ser Lys Glu Glu Ala	
	710 715 720	
55	GAG ATG GCC TAC AAA GAC TTC CTG CTC CAG TCC AGC ACC GTG GCC GCC	2445
	Glu Met Ala Tyr Lys Asp Phe Leu Leu Gln Ser Ser Thr Val Ala Ala	
	725 730 735	
50	GAG GCC CAG GAC GGC CCC CAG GAA GCC TAGACGGTGT CGCCGCCTGC TCCCTGCA	2500
	Glu Ala Gln Asp Gly Pro Gln Glu Ala	
	740 745	
55	CCCATGACAA TCACCTTCAG AATCATGTCG ATCCTGGGGG CCCTCAGCTC CTGGGGACCC	2560
	CACTCCCTGC TCTAACACCT GCCTAGGTTT TTCCTACTGT CCTCAGAGGC GTGCTGGTCC	2620
	CCTCCTCAGT GACATCAAAG CCTGGCCTAA TTGTTCTTAT TGGGGATGAG GGTGGCATGA	2680
	GGAGGTCCCA CTTGCAACTT CTTTCTGTTG AGAGAACCTC AGGTACGGAG AAGAATAGAG	2740

GTCCTCATGG GTCCCTTGAA GGAAGAGGGA CCAGGGTGGG AGAGCTGATT GCAGAAAGGA 2800  
 GAGACGTGCA GCGCCCCTCT GCACCCTTAT CATGGGATGT CAACAGAATT TTTTCCCTCC 2860  
 ACTCCATCCC TCCCTCCCGT CCTTCCCCTC TTCTTCTTTC CTTACCATCA AAAGATGTA 2919

10

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

15

(A) LENGTH: 746 amino acids  
 (B) TYPE: amino acid  
 (C) STRANDNESS: single  
 (D) TOPOLOGY: linear

20

DESCRIPTION: Human Polyimmunoglobulin Receptor  
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4

25

Lys Ser Pro Ile Phe Gly Pro Glu Glu Val Asn Ser Val Glu Gly Asn  
 1 5 10 15

25

Ser Val Ser Ile Thr Cys Tyr Tyr Pro Pro Thr Ser Val Asn Arg His  
 20 25 30

30

Thr Arg Lys Tyr Trp Cys Arg Gln Gly Ala Arg Gly Gly Cys Ile Thr  
 35 40 45

Leu Ile Ser Ser Glu Gly Tyr Val Ser Ser Lys Tyr Ala Gly Arg Ala  
 50 55 60

35

Asn Leu Thr Asn Phe Pro Glu Asn Gly Thr Phe Val Val Asn Ile Ala  
 65 70 75 80

Gln Leu Ser Gln Asp Asp Ser Gly Arg Tyr Lys Cys Gly Leu Gly Ile  
 85 90 95

40

Asn Ser Arg Gly Leu Ser Phe Asp Val Ser Leu Glu Val Ser Gln Gly  
 100 105 110

45

Pro Gly Leu Leu Asn Asp Thr Lys Val Tyr Thr Val Asp Leu Gly Arg  
 115 120 125

Thr Val Thr Ile Asn Cys Pro Phe Lys Thr Glu Asn Ala Gln Lys Arg  
 130 135 140

50

Lys Ser Leu Tyr Lys Gln Ile Gly Leu Tyr Pro Val Leu Val Ile Asp  
 145 150 155 160

Ser Ser Gly Tyr Val Asn Pro Asn Tyr Thr Gly Arg Ile Arg Leu Asp  
 165 170 175

55

Ile Gln Gly Thr Gly Gln Leu Leu Phe Ser Val Val Ile Asn Gln Leu  
 180 185 190

Arg Leu Ser Asp Ala Gly Gln Tyr Leu Cys Gln Ala Gly Asp Asp Ser  
 195 200 205



	Asn	Ser	Asn	Lys	Lys	Asn	Ala	Asp	Leu	Gln	Val	Leu	Lys	Pro	Glu	Pro
	210						215					220				
5	Glu	Leu	Val	Tyr	Glu	Asp	Leu	Arg	Gly	Ser	Val	Thr	Phe	His	Cys	Ala
	225					230					235					240
	Leu	Gly	Pro	Glu	Val	Ala	Asn	Val	Ala	Lys	Phe	Leu	Cys	Arg	Gln	Ser
					245					250					255	
10	Ser	Gly	Glu	Asn	Cys	Asp	Val	Val	Val	Asn	Thr	Leu	Gly	Lys	Arg	Ala
				260					265					270		
	Pro	Ala	Phe	Glu	Gly	Arg	Ile	Leu	Leu	Asn	Pro	Gln	Asp	Lys	Asp	Gly
15			275					280					285			
	Ser	Phe	Ser	Val	Val	Ile	Thr	Gly	Leu	Arg	Lys	Glu	Asp	Ala	Gly	Arg
	290						295					300				
20	Tyr	Leu	Cys	Gly	Ala	His	Ser	Asp	Gly	Gln	Leu	Gln	Glu	Gly	Ser	Pro
	305					310					315					320
	Ile	Gln	Ala	Trp	Gln	Leu	Phe	Val	Asn	Glu	Glu	Ser	Thr	Ile	Pro	Arg
					325					330					335	
25	Ser	Pro	Thr	Val	Val	Lys	Gly	Val	Ala	Gly	Ser	Ser	Val	Ala	Val	Leu
				340					345					350		
	Cys	Pro	Tyr	Asn	Arg	Lys	Glu	Ser	Lys	Ser	Ile	Lys	Tyr	Trp	Cys	Leu
30			355					360					365			
	Trp	Glu	Gly	Ala	Gln	Asn	Gly	Arg	Cys	Pro	Leu	Leu	Val	Asp	Ser	Glu
	370						375					380				
35	Gly	Trp	Val	Lys	Ala	Gln	Tyr	Glu	Gly	Arg	Leu	Ser	Leu	Leu	Glu	Glu
	385					390					395					400
	Pro	Gly	Asn	Gly	Thr	Phe	Thr	Val	Ile	Leu	Asn	Gln	Leu	Thr	Ser	Arg
					405					410					415	
40	Asp	Ala	Gly	Phe	Tyr	Trp	Cys	Leu	Thr	Asn	Gly	Asp	Thr	Leu	Trp	Arg
				420					425					430		
	Thr	Thr	Val	Glu	Ile	Lys	Ile	Ile	Glu	Gly	Glu	Pro	Asn	Leu	Lys	Val
45			435				440						445			
	Pro	Gly	Asn	Val	Thr	Ala	Val	Leu	Gly	Glu	Thr	Leu	Lys	Val	Pro	Cys
	450						455					460				
50	His	Phe	Pro	Cys	Lys	Phe	Ser	Ser	Tyr	Glu	Lys	Tyr	Trp	Cys	Lys	Trp
	465					470					475					480
	Asn	Asn	Thr	Gly	Cys	Gln	Ala	Leu	Pro	Ser	Gln	Asp	Glu	Gly	Pro	Ser
					485					490					495	
55	Lys	Ala	Phe	Val	Asn	Cys	Asp	Glu	Asn	Ser	Arg	Leu	Val	Ser	Leu	Thr
				500					505					510		

Leu Asn Leu Val Thr Arg Ala Asp Glu Gly Trp Tyr Trp Cys Gly Val  
 515 520 525  
 5 Lys Gln Gly His Phe Tyr Gly Glu Thr Ala Ala Val Tyr Val Ala Val  
 530 535 540  
 Glu Glu Arg Lys Ala Ala Gly Ser Arg Asp Val Ser Leu Ala Lys Ala  
 545 550 555 560  
 10 Asp Ala Ala Pro Asp Glu Lys Val Leu Asp Ser Gly Phe Arg Glu Ile  
 565 570 575  
 Glu Asn Lys Ala Ile Gln Asp Pro Arg Leu Phe Ala Glu Glu Lys Ala  
 580 585 590  
 15 Val Ala Asp Thr Arg Asp Gln Ala Asp Gly Ser Arg Ala Ser Val Asp  
 595 600 605  
 20 Ser Gly Ser Ser Glu Glu Gln Gly Gly Ser Ser Arg Ala Leu Val Ser  
 610 615 620  
 Thr Leu Val Pro Leu Gly Leu Val Leu Ala Val Gly Ala Val Ala Val  
 625 630 635 640  
 25 Gly Val Ala Arg Ala Arg His Arg Lys Asn Val Asp Arg Val Ser Ile  
 645 650 655  
 Arg Ser Tyr Arg Thr Asp Ile Ser Met Ser Asp Phe Glu Asn Ser Arg  
 660 665 670  
 30 Glu Phe Gly Ala Asn Asp Asn Met Gly Ala Ser Ser Ile Thr Gln Glu  
 675 680 685  
 Thr Ser Leu Gly Gly Lys Glu Glu Phe Val Ala Thr Thr Glu Ser Thr  
 690 695 700  
 Thr Glu Thr Lys Glu Pro Lys Lys Ala Lys Arg Ser Ser Lys Glu Glu  
 705 710 715 720  
 40 Ala Glu Met Ala Tyr Lys Asp Phe Leu Leu Gln Ser Ser Thr Val Ala  
 725 730 735  
 Ala Glu Ala Gln Asp Gly Pro Gln Glu Ala  
 740 745  
 45

(2) INFORMATION FOR SEQ ID NO: 5:

50 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3630 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 55 (D) TOPOLOGY: linear  
 DESCRIPTION: Bovine Polyimmunoglobulin Receptor

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 152....2425

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

5 GATCTCCTCG GAGGGTCGTG CAGCGGCCCT' GGTCCCTGC CGGCACCAGT ACTTGCGCGT 60

GTGCTCCCAA AGCTGACGGG ATAGGAGGAA GGAGCTCAAA CAACCACACA GGACGGTGGC 120

10 TGGCGGCAGA GACCCGCGGG AGCCCCCAGC G ATG TCG CGC CTG TTC CTC GCC 172  
Met Ser Arg Leu Phe Leu Ala  
1 5

15 TGC CTG CTG GCC ATC TTC CCA GTG GTC TCC ATG AAG AGT CCC ATC TTC 220  
Cys Leu Leu Ala Ile Phe Pro Val Val Ser Met Lys Ser Pro Ile Phe  
10 15 20

20 GGT CCC GAG GAG GTG AGC AGC GTG GAA GGC CGC TCA GTG TCC ATC AAG 268  
Gly Pro Glu Glu Val Ser Ser Val Glu Gly Arg Ser Val Ser Ile Lys  
25 30 35

25 TGC TAC TAC CCG CCC ACC TCC GTC AAC CGG CAC ACG CGC AAG TAC TGG 316  
Cys Tyr Tyr Pro Pro Thr Ser Val Asn Arg His Thr Arg Lys Tyr Trp  
40 45 50 55

TGC CGG CAG GGA GCC CAG GGC CGC TGC ACG ACC CTC ATC TCC TCG GAG 364  
Cys Arg Gln Gly Ala Gln Gly Arg Cys Thr Thr Leu Ile Ser Ser Glu  
60 65 70

30 GGC TAC GTC TCC GAC GAC TAC GTG GGC AGA GCC AAC CTC ACC AAC TTC 412  
Gly Tyr Val Ser Asp Asp Tyr Val Gly Arg Ala Asn Leu Thr Asn Phe  
75 80 85

35 CCG GAG AGC GGC ACG TTT GTG GTG GAC ATC AGC CAT CTC ACC CAT AAA 460  
Pro Glu Ser Gly Thr Phe Val Val Asp Ile Ser His Leu Thr His Lys  
90 95 100

40 GAC TCA GGG CGC TAC AAG TGT GGC CTG GGC ATT AGC AGC CGT GGC CTT 508  
Asp Ser Gly Arg Tyr Lys Cys Gly Leu Gly Ile Ser Ser Arg Gly Leu  
105 110 115

AAC TTC GAT GTG AGC CTG GAG GTC AGC CAA GAT CCT GCA CAG GCA AGT 556  
Asn Phe Asp Val Ser Leu Glu Val Ser Gln Asp Pro Ala Gln Ala Ser  
120 125 130 135

45 CAT GCC CAC GTC TAC ACT ATA GAC CTG GGC AGG ACT GTG ACC ATC AAC 604  
His Ala His Val Tyr Thr Ile Asp Leu Gly Arg Thr Val Thr Ile Asn  
140 145 150

50 TGC CCT TTC ACG CGT GCG AAT TCT GAG AAG AGA AAA TCC TTG TGC AAG 652  
Cys Pro Phe Thr Arg Ala Asn Ser Glu Lys Arg Lys Ser Leu Cys Lys  
155 160 165

55 AAG ACA ATC CAG GAC TGT TTC CAA GTT GTC GAC TCC ACC GGG TAT GTG 700  
Lys Thr Ile Gln Asp Cys Phe Gln Val Val Asp Ser Thr Gly Tyr Val  
170 175 180

AGC AAC AGC TAT AAA GAC AGA GCA CAT ATC AGT ATC CTA GGT ACC AAC 748  
Ser Asn Ser Tyr Lys Asp Arg Ala His Ile Ser Ile Leu Gly Thr Asn

	185					190					195						
5	ACA Thr 200	TTA Leu	GTG Val	TTC Phe	AGC Ser	GTT Val 205	GTC Val	ATC Ile	AAC Asn	CGA Arg	GTC Val 210	AAG Lys	CTC Leu	AGT Ser	GAT Asp	GCT Ala 215	796
10	GGG Gly	ATG Met	TAT Tyr	GTC Val	TGC Cys 220	CAG Gln	GCT Ala	GGG Gly	GAC Asp	GAT Asp 225	GCC Ala	AAA Lys	GCC Ala	GAT Asp	AAA Lys	ATC Ile 230	844
15	AAC Asn	ATT Ile	GAC Asp	CTC Leu 235	CAG Gln	GTG Val	CTG Leu	GAG Glu	CCT Pro 240	GAG Glu	CCT Pro	GAG Glu	CTG Leu	GTT Val 245	TAT Tyr	GGA Gly	892
20	GAC Asp	TTG Leu	AGG Arg 250	AGC Ser	TCG Ser	GTG Val	ACC Thr	TTT Phe 255	GAC Asp	TGT Cys	TCC Ser	CTG Leu	GGC Gly 260	CCC Pro	GAG Glu	GTG Val	940
25	GCA Ala 265	AAT Asn	GTG Val	CCC Pro	AAA Lys	TTT Phe	CTG Leu 270	TGC Cys	CAG Gln	AAG Lys	AAG Lys	AAT Asn 275	GGG Gly	GGA Gly	GCT Ala	TGC Cys	988
30	AAT Asn 280	GTA Val	GTC Val	ATC Ile	AAC Asn	ACG Thr 285	TTG Leu	GGG Gly	AAG Lys	AAG Lys	GCT Ala 290	CAG Gln	GAC Asp	TTC Phe	CAG Gln	GGC Gly 295	1036
35	AGG Arg	ATC Ile	GTG Val	TCC Ser	GTG Val 300	CCC Pro	AAG Lys	GAC Asp	AAT Asn	GGT Gly 305	GTC Val	TTC Phe	AGT Ser	GTG Val	CAC His	ATT Ile 310	1084
40	ACC Thr	AGC Ser	CTG Leu	AGG Arg 315	AAA Lys	GAG Glu	GAC Asp	GCA Ala	GGG Gly 320	CGC Arg	TAC Tyr	GTG Val	TGC Cys	GGG Gly 325	GCC Ala	CAG Gln	1132
45	CCT Pro	GAG Glu	GGT Gly 330	GAG Glu	CCC Pro	CAG Gln	GAC Asp	GGC Gly 335	TGG Trp	CCT Pro	GTG Val	CAG Gln	GCC Ala	TGG Trp	CAA Gln	CTC Leu	1180
50	TTC Phe 345	GTC Val	AAT Asn	GAA Glu	GAG Glu	ACG Thr	GCA Ala 350	ATC Ile	CCC Pro	GCA Ala	AGC Ser	CCC Pro	TCC Ser	GTG Val	GTG Val	AAA Lys	1228
55	GGT Gly 360	GTG Val	AGG Arg	GGA Gly	GGC Gly	TCT Ser 365	GTG Val	ACT Thr	GTA Val	TCT Ser	TGC Cys 370	CCC Pro	TAC Tyr	AAC Asn	CCT Pro	AAG Lys 375	1276
60	GAT Asp	GCC Ala	AAC Asn	AGC Ser	GCG Ala 380	AAG Lys	TAC Tyr	TGG Trp	TGT Cys	CAC His 385	TGG Trp	GAA Glu	GAG Glu	GCT Ala	CAA Gln	AAC Asn 390	1324
65	GGC Gly	CGC Arg	TGC Cys	CCG Pro 395	CGG Arg	CTG Leu	GTG Val	GAG Glu	AGC Ser 400	CGG Arg	GGG Gly	CTG Leu	ATG Met	AAG Lys	GAG Glu	CAG Gln	1372
70	TAC Tyr	GAG Glu	GGC Gly 410	AGG Arg	CTG Leu	GTG Val	CTG Leu	CTC Leu 415	ACC Thr	GAG Glu	CCG Pro	GGC Gly 420	AAC Asn	GGC Gly	ACC Thr	TAC Tyr	1420
75	ACC	GTC	ATC	CTC	AAC	CAG	CTC	ACC	GAT	CAG	GAC	GCC	GGC	TTC	TAC	TGG	1468

	Thr	Val	Ile	Leu	Asn	Gln	Leu	Thr	Asp	Gln	Asp	Ala	Gly	Phe	Tyr	Trp	
	425						430					435					
5	TGC	GTG	ACC	GAC	GGC	GAC	ACG	CGC	TGG	ATC	TCC	ACA	GTG	GAG	CTC	AAG	1516
	Cys	Val	Thr	Asp	Gly	Asp	Thr	Arg	Trp	Ile	Ser	Thr	Val	Glu	Leu	Lys	
	440					445					450					455	
10	GTT	GTC	CAA	GGA	GAA	CCA	AGC	CTC	AAG	GTA	CCC	AAG	AAC	GTC	ACG	GCT	1564
	Val	Val	Gln	Gly	Glu	Pro	Ser	Leu	Lys	Val	Pro	Lys	Asn	Val	Thr	Ala	
					460					465					470		
15	TGG	CTG	GGA	GAG	CCC	TTA	AAG	CTC	TCC	TGC	CAC	TTC	CCC	TGC	AAA	TTC	1612
	Trp	Leu	Gly	Glu	Pro	Leu	Lys	Leu	Ser	Cys	His	Phe	Pro	Cys	Lys	Phe	
				475				480						485			
	TAC	TCC	TTT	GAG	AAG	TAC	TGG	TGT	AAG	TGG	AGC	AAC	AGA	GGC	TGC	AGC	1660
	Tyr	Ser	Phe	Glu	Lys	Tyr	Trp	Cys	Lys	Trp	Ser	Asn	Arg	Gly	Cys	Ser	
			490					495					500				
20	GCC	CTG	CCC	ACC	CAG	AAC	GAC	GGC	CCC	AGC	CAG	GCC	TTT	GTG	AGC	TGC	1708
	Ala	Leu	Pro	Thr	Gln	Asn	Asp	Gly	Pro	Ser	Gln	Ala	Phe	Val	Ser	Cys	
		505					510					515					
25	GAC	CAG	AAC	AGC	CAG	GTC	GTC	TCC	CTG	AAC	CTG	GAC	ACA	GTC	ACC	AAG	1756
	Asp	Gln	Asn	Ser	Gln	Val	Val	Ser	Leu	Asn	Leu	Asp	Thr	Val	Thr	Lys	
	520					525					530					535	
30	GAG	GAT	GAA	GGC	TGG	TAC	TGG	TGT	GGA	GTG	AAG	GAA	GGC	CCC	CGA	TAC	1804
	Glu	Asp	Glu	Gly	Trp	Tyr	Trp	Cys	Gly	Val	Lys	Glu	Gly	Pro	Arg	Tyr	
					540				545						550		
35	GGG	GAG	ACG	GCG	GCT	GTC	TAC	GTG	GCA	GTG	GAG	AGC	AGG	GTG	AAG	GGG	1852
	Gly	Glu	Thr	Ala	Ala	Val	Tyr	Val	Ala	Val	Glu	Ser	Arg	Val	Lys	Gly	
				555				560						565			
	TCC	CAA	GGC	GCC	AAG	CAA	GTG	AAA	GCT	GCC	CCT	GCG	GGG	GCG	GCA	ATA	1900
	Ser	Gln	Gly	Ala	Lys	Gln	Val	Lys	Ala	Ala	Pro	Ala	Gly	Ala	Ala	Ile	
			570					575					580				
40	CAG	TCG	AGG	GCC	GGG	GAG	ATC	CAG	AAC	AAA	GCC	CTT	CTG	GAC	CCC	AGC	1948
	Gln	Ser	Arg	Ala	Gly	Glu	Ile	Gln	Asn	Lys	Ala	Leu	Leu	Asp	Pro	Ser	
		585					590					595					
45	TTT	TTC	GCA	AAG	GAA	AGT	GTG	AAG	GAC	GCT	GCT	GGT	GGA	CCC	GGA	GCA	1996
	Phe	Phe	Ala	Lys	Glu	Ser	Val	Lys	Asp	Ala	Ala	Gly	Gly	Pro	Gly	Ala	
	600					605					610					615	
50	CCT	GCA	GAT	CCT	GGC	CGC	CCT	ACA	GGA	TAC	AGC	GGG	AGC	TCC	AAA	GCA	2044
	Pro	Ala	Asp	Pro	Gly	Arg	Pro	Thr	Gly	Tyr	Ser	Gly	Ser	Ser	Lys	Ala	
					620					625					630		
55	CTG	GTC	TCC	ACC	CTG	GTG	CCC	CTG	GCC	CTG	GTC	CTG	GTC	GCA	GGG	GTC	2092
	Leu	Val	Ser	Thr	Leu	Val	Pro	Leu	Ala	Leu	Val	Leu	Val	Ala	Gly	Val	
				635				640						645			
	GTG	GCG	ATC	GGG	GTG	GTC	CGA	GCC	CGG	CAC	AGG	AAG	AAC	GTC	GAC	CGG	2140
	Val	Ala	Ile	Gly	Val	Val	Arg	Ala	Arg	His	Arg	Lys	Asn	Val	Asp	Arg	
			650					655					660				

	ATT TCA ATC AGG AGC TAC CGG ACA GAT ATC AGC ATG TCA GAC TTT GAG	2188
	Ile Ser Ile Arg Ser Tyr Arg Thr Asp Ile Ser Met Ser Asp Phe Glu	
	665 670 675	
5	AAC TCC AGG GAT TTT GAA GGA CGT GAC AAC ATG GGA GCC TCT CCA GAG	2236
	Asn Ser Arg Asp Phe Glu Gly Arg Asp Asn Met Gly Ala Ser Pro Glu	
	680 685 690 695	
10	GCC CAA GAG ACG TCT CTC GGA GGG AAG GAC GAG TTT GCC ACC ACT ACC	2284
	Ala Gln Glu Thr Ser Leu Gly Gly Lys Asp Glu Phe Ala Thr Thr Thr	
	700 705 710	
15	GAG GAC ACC GTG GAG AGC AAA GAA CCC AAG AAG GCA AAG AGG TCG TCC	2332
	Glu Asp Thr Val Glu Ser Lys Glu Pro Lys Lys Ala Lys Arg Ser Ser	
	715 720 725	
20	AAG GAG GAA GCC GAC GAG GCC TTC ACC ACC TTC CTC CTC CAG GCC AAA	2380
	Lys Glu Glu Ala Asp Glu Ala Phe Thr Thr Phe Leu Leu Gln Ala Lys	
	730 735 740	
25	AAC CTG GCC TCC GCC GCA ACC CAG AAC GGC CCG ACA GAA GCC TAG ACGGAG	2431
	Asn Leu Ala Ser Ala Ala Thr Gln Asn Gly Pro Thr Glu Ala	
	745 750 755	
30	CCCTGGGCGC CCCTTCCCTC CGCACGTGGC AATCACGCTC CGAATCACGC TGATCCTCAG	2491
	GGCCCTCAGC TCGGGGGGCT CCACTGCCTG CACTCACACC CCGCCTAGGC TTCTCCTGTC	2551
	TGTCCTCAGA GGGTGTGCTG GTTCCTTCTT GGTGGCATCC AAGCCTGGCT TACTTGTTCC	2611
	TATTGGGGGT GAGGTGGTAC GAGGAGTTCC CACCTGCAGC TTATTCTGAAC GAGAGAACTA	2671
	AAGGTGTGGA GGAGAATTAA GATCGCAGAG GGGCCTCTCA GAAAGAAAAG GAGTGGGTGG	2731
35	GGAGACAACC GCAGAAAGGG GGCCATTTCAG CGCTTCCCTG TCCCCTTATT TGGGGATGTC	2791
	AGTGGAATCC TCCCTTCCAC CCCATCTCTG CACCTCTCCA TCCCCACTCC ATTCCATCTT	2851
40	CTCTTCTTCT TTCCCTCATT AAAAATGTGC ATTTGGTTAC TCACTAGATT CCAGGGACTC	2911
	TGCTAGACAC TGGGATAGGT AGGCCGCAAT CCCAGGCGGC AGCCTTCCGC AAACATCAAG	2971
	GAGCCCCTGG AGCCACAGC ATCTCTTCAC GTGTACACTC ACTGACCTCT GCCTCTGCTG	3031
45	GGAGAAATCA TAAAGGGTCT GCAGCCCTGA GGCCTTAGGG ATTATGTAAC ACAGGCATAC	3091
	ACACAAGGCA CCATCAACAC ATTCTTACCA TTTCACAGGT GAGAAAGCCG AGGTCCTGAG	3151
50	AGGTGGAGAG GTTTGCTCAG AGTCAGCAAG TGAGATGTAC GAGTCTCAAG CTAAAGATTT	3211
	GACACCTGCT GTCCCTACAG GAGGGCCTCC TCTCTCCAGA TGAGACAGCA TTCCATAGGA	3271
	AGGAGAAGAA AAATGTAAAT AAGACTGGTC TTTCACAGGC CCCACATCAG GGAAGATACC	3331
55	CCTTTCCTG TCTGTCACTC ACAGAGACCT AATAGGATAA GAGAATGGTC AACACTCAAA	3391
	CCCCGAATG TGAAGAGTTC TAAGTGAAA GGGAGGAAAA AGGGGGGATT TGATGGTGCC	3451
	AGGGAGGGGC TGATCTCCAA AGAACTAAGG TTTAAGTTTT TTTGTTTTTT TTTTCCTTC	3511

TTCTAAGCTC TGCACTTCAA CTAGCATCTA TGAGCTGGCA CTTGCTAACA AATCAAAAAT 3571  
 GTGAATTAAT TAATAATTAA AGACCATGAT TTCCTCCAAA AAAAAAAAAA AAAAAAAAAA 3630

5

(2) INFORMATION FOR SEQ ID NO: 6:

10 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 757 amino acids  
 (B) TYPE: amino acid  
 (C) STRANDNESS: single  
 (D) TOPOLOGY: linear  
 15 DESCRIPTION: Bovine Polyimmunoglobulin Receptor

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

20 Met Ser Arg Leu Phe Leu Ala Cys Leu Leu Ala Ile Phe Pro Val Val  
 1 5 10 15  
 Ser Met Lys Ser Pro Ile Phe Gly Pro Glu Glu Val Ser Ser Val Glu  
 20 25 30  
 25 Gly Arg Ser Val Ser Ile Lys Cys Tyr Tyr Pro Pro Thr Ser Val Asn  
 35 40 45  
 Arg His Thr Arg Lys Tyr Trp Cys Arg Gln Gly Ala Gln Gly Arg Cys  
 30 50 55 60  
 Thr Thr Leu Ile Ser Ser Glu Gly Tyr Val Ser Asp Asp Tyr Val Gly  
 65 70 75 80  
 35 Arg Ala Asn Leu Thr Asn Phe Pro Glu Ser Gly Thr Phe Val Val Asp  
 85 90 95  
 Ile Ser His Leu Thr His Lys Asp Ser Gly Arg Tyr Lys Cys Gly Leu  
 100 105 110  
 40 Gly Ile Ser Ser Arg Gly Leu Asn Phe Asp Val Ser Leu Glu Val Ser  
 115 120 125  
 Gln Asp Pro Ala Gln Ala Ser His Ala His Val Tyr Thr Ile Asp Leu  
 45 130 135 140  
 Gly Arg Thr Val Thr Ile Asn Cys Pro Phe Thr Arg Ala Asn Ser Glu  
 145 150 155 160  
 50 Lys Arg Lys Ser Leu Cys Lys Lys Thr Ile Gln Asp Cys Phe Gln Val  
 165 170 175  
 Val Asp Ser Thr Gly Tyr Val Ser Asn Ser Tyr Lys Asp Arg Ala His  
 180 185 190  
 55 Ile Ser Ile Leu Gly Thr Asn Thr Leu Val Phe Ser Val Val Ile Asn  
 195 200 205

Arg Val Lys Leu Ser Asp Ala Gly Met Tyr Val Cys Gln Ala Gly Asp  
 210 215 220  
 5 Asp Ala Lys Ala Asp Lys Ile Asn Ile Asp Leu Gln Val Leu Glu Pro  
 225 230 235 240  
 Glu Pro Glu Leu Val Tyr Gly Asp Leu Arg Ser Ser Val Thr Phe Asp  
 245 250 255  
 10 Cys Ser Leu Gly Pro Glu Val Ala Asn Val Pro Lys Phe Leu Cys Gln  
 260 265 270  
 Lys Lys Asn Gly Gly Ala Cys Asn Val Val Ile Asn Thr Leu Gly Lys  
 275 280 285  
 15 Lys Ala Gln Asp Phe Gln Gly Arg Ile Val Ser Val Pro Lys Asp Asn  
 290 295 300  
 20 Gly Val Phe Ser Val His Ile Thr Ser Leu Arg Lys Glu Asp Ala Gly  
 305 310 315 320  
 Arg Tyr Val Cys Gly Ala Gln Pro Glu Gly Glu Pro Gln Asp Gly Trp  
 325 330 335  
 25 Pro Val Gln Ala Trp Gln Leu Phe Val Asn Glu Glu Thr Ala Ile Pro  
 340 345 350  
 Ala Ser Pro Ser Val Val Lys Gly Val Arg Gly Gly Ser Val Thr Val  
 355 360 365  
 30 Ser Cys Pro Tyr Asn Pro Lys Asp Ala Asn Ser Ala Lys Tyr Trp Cys  
 370 375 380  
 35 His Trp Glu Glu Ala Gln Asn Gly Arg Cys Pro Arg Leu Val Glu Ser  
 385 390 395 400  
 Arg Gly Leu Met Lys Glu Gln Tyr Glu Gly Arg Leu Val Leu Leu Thr  
 405 410 415  
 40 Glu Pro Gly Asn Gly Thr Tyr Thr Val Ile Leu Asn Gln Leu Thr Asp  
 420 425 430  
 Gln Asp Ala Gly Phe Tyr Trp Cys Val Thr Asp Gly Asp Thr Arg Trp  
 435 440 445  
 45 Ile Ser Thr Val Glu Leu Lys Val Val Gln Gly Glu Pro Ser Leu Lys  
 450 455 460  
 50 Val Pro Lys Asn Val Thr Ala Trp Leu Gly Glu Pro Leu Lys Leu Ser  
 465 470 475 480  
 Cys His Phe Pro Cys Lys Phe Tyr Ser Phe Glu Lys Tyr Trp Cys Lys  
 485 490 495  
 55 Trp Ser Asn Arg Gly Cys Ser Ala Leu Pro Thr Gln Asn Asp Gly Pro  
 500 505 510  
 Ser Gln Ala Phe Val Ser Cys Asp Gln Asn Ser Gln Val Val Ser Leu  
 515 520 525



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      Asn Leu Asp Thr Val Thr Lys Glu Asp Glu Gly Trp Tyr Trp Cys Gly
      530                               535                               540
5  Val Lys Glu Gly Pro Arg Tyr Gly Glu Thr Ala Ala Val Tyr Val Ala
   545                               550                               555                               560

      Val Glu Ser Arg Val Lys Gly Ser Gln Gly Ala Lys Gln Val Lys Ala
                               565                               570                               575
10 Ala Pro Ala Gly Ala Ala Ile Gln Ser Arg Ala Gly Glu Ile Gln Asn
   580                               585                               590

      Lys Ala Leu Leu Asp Pro Ser Phe Phe Ala Lys Glu Ser Val Lys Asp
15 595                               600                               605

      Ala Ala Gly Gly Pro Gly Ala Pro Ala Asp Pro Gly Arg Pro Thr Gly
   610                               615                               620

20 Tyr Ser Gly Ser Ser Lys Ala Leu Val Ser Thr Leu Val Pro Leu Ala
   625                               630                               635                               640

      Leu Val Leu Val Ala Gly Val Val Ala Ile Gly Val Val Arg Ala Arg
                               645                               650                               655
25 His Arg Lys Asn Val Asp Arg Ile Ser Ile Arg Ser Tyr Arg Thr Asp
   660                               665                               670

      Ile Ser Met Ser Asp Phe Glu Asn Ser Arg Asp Phe Glu Gly Arg Asp
30 675                               680                               685

      Asn Met Gly Ala Ser Pro Glu Ala Gln Glu Thr Ser Leu Gly Gly Lys
   690                               695                               700

35 Asp Glu Phe Ala Thr Thr Thr Glu Asp Thr Val Glu Ser Lys Glu Pro
   705                               710                               715                               720

      Lys Lys Ala Lys Arg Ser Ser Lys Glu Glu Ala Asp Glu Ala Phe Thr
                               725                               730                               735
40 Thr Phe Leu Leu Gln Ala Lys Asn Leu Ala Ser Ala Ala Thr Gln Asn
   740                               745                               750

      Gly Pro Thr Glu Ala
45 755

```

50 (2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

```

55 (A) LENGTH:          3095 base pairs
   (B) TYPE:           nucleic acid
   (C) STRANDEDNESS:   single
   (D) TOPOLOGY:       linear
      DESCRIPTION:      Mouse Polyimmunoglobulin Receptor

```

## (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence  
(B) LOCATION: 85...2400

5

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

	TCACCTGGAG AGAAGGAAGT AGCTAAAACA TTCTCATACA AGAAGCCAAC CTGAGCGGCA	60
10	CAGCCCCCCT GGAAGCCACA AGCA ATG AGG CTC TAC TTG TTC ACG CTC TTG Met Arg Leu Tyr Leu Phe Thr Leu Leu	111
	1 5	
15	GTA ACT GTC TTT TCA GGG GTC TCC ACA AAA AGC CCC ATA TTT GGT CCC Val Thr Val Phe Ser Gly Val Ser Thr Lys Ser Pro Ile Phe Gly Pro	159
	10 15 20 25	
20	CAG GAG GTG AGT AGT ATA GAA GGC GAC TCT GTT TCC ATC ACG TGC TAC Gln Glu Val Ser Ser Ile Glu Gly Asp Ser Val Ser Ile Thr Cys Tyr	207
	30 35 40	
25	TAC CCA GAC ACC TCT GTC AAC CGG CAC ACC CGG AAA TAC TGG TGC CGA Tyr Pro Asp Thr Ser Val Asn Arg His Thr Arg Lys Tyr Trp Cys Arg	255
	45 50 55	
30	CAA GGA GCC AGC GGC ATG TGC ACA ACG CTC ATC TCT TCA AAT GGC TAC Gln Gly Ala Ser Gly Met Cys Thr Thr Leu Ile Ser Ser Asn Gly Tyr	303
	60 65 70	
35	CTC TCC AAG GAG TAT TCA GGC AGA GCC AAC CTC ATC AAC TTC CCA GAG Leu Ser Lys Glu Tyr Ser Gly Arg Ala Asn Leu Ile Asn Phe Pro Glu	351
	75 80 85	
40	AAC AAC ACA TTT GTG ATT AAC ATT GAG CAG CTC ACC CAG GAC GAC ACT Asn Asn Thr Phe Val Ile Asn Ile Glu Gln Leu Thr Gln Asp Asp Thr	399
	90 95 100 105	
45	GGG AGC TAC AAG TGT GGC CTG GGT ACC AGT AAC CGA GGC CTG TCC TTC Gly Ser Tyr Lys Cys Gly Leu Gly Thr Ser Asn Arg Gly Leu Ser Phe	447
	110 115 120	
50	GAT GTC AGC CTG GAG GTC AGC CAG GTT CCT GAG TTG CCG AGT GAC ACC Asp Val Ser Leu Glu Val Ser Gln Val Pro Glu Leu Pro Ser Asp Thr	495
	125 130 135	
55	CAC GTC TAC ACA AAG GAC ATA GGC AGA AAT GTG ACC ATT GAA TGC CCT His Val Tyr Thr Lys Asp Ile Gly Arg Asn Val Thr Ile Glu Cys Pro	543
	140 145 150	
60	TTC AAA AGG GAG AAT GTT CCC AGC AAG AAA TCC CTG TGT AAG AAG ACA Phe Lys Arg Glu Asn Val Pro Ser Lys Lys Ser Leu Cys Lys Lys Thr	591
	155 160 165	
65	AAC CAG TCC TGC GAA CTT GTC ATT GAC TCT ACT GAG AAG GTG AAC CCC Asn Gln Ser Cys Glu Leu Val Ile Asp Ser Thr Glu Lys Val Asn Pro	639
	170 175 180 185	

	AGC	TAT	ATA	GGC	AGA	GCA	AAA	CTT	TTT	ATG	AAA	GGG	ACC	GAC	CTA	ACT	687
	Ser	Tyr	Ile	Gly	Arg	Ala	Lys	Leu	Phe	Met	Lys	Gly	Thr	Asp	Leu	Thr	
					190					195					200		
5	GTA	TTC	TAT	GTC	AAC	ATT	AGT	CAC	CTA	ACG	CAC	AAT	GAT	GCT	GGG	CTG	735
	Val	Phe	Tyr	Val	Asn	Ile	Ser	His	Leu	Thr	His	Asn	Asp	Ala	Gly	Leu	
				205					210					215			
10	TAC	ATC	TGC	CAA	GCT	GGA	GAA	GGT	CCT	AGT	GCT	GAT	AAG	AAG	AAT	GTT	783
	Tyr	Ile	Cys	Gln	Ala	Gly	Glu	Gly	Pro	Ser	Ala	Asp	Lys	Lys	Asn	Val	
			220					225					230				
15	GAC	CTC	CAG	GTG	CTA	GCG	CCT	GAG	CCA	GAG	CTG	CTT	TAT	AAA	GAC	CTG	831
	Asp	Leu	Gln	Val	Leu	Ala	Pro	Glu	Pro	Glu	Leu	Leu	Tyr	Lys	Asp	Leu	
		235					240					245					
20	AGG	TCC	TCA	GTG	ACT	TTT	GAA	TGT	GAC	CTG	GGC	CGT	GAG	GTG	GCA	AAC	879
	Arg	Ser	Ser	Val	Thr	Phe	Glu	Cys	Asp	Leu	Gly	Arg	Glu	Val	Ala	Asn	
		250				255					260					265	
	GAG	GCC	AAA	TAT	CTG	TGC	CGG	ATG	AAT	AAG	GAA	ACC	TGT	GAT	GTG	ATC	927
	Glu	Ala	Lys	Tyr	Leu	Cys	Arg	Met	Asn	Lys	Glu	Thr	Cys	Asp	Val	Ile	
					270					275					280		
25	ATT	AAC	ACC	CTG	GGG	AAG	AGG	GAT	CCA	GAC	TTT	GAG	GGC	AGG	ATC	CTG	975
	Ile	Asn	Thr	Leu	Gly	Lys	Arg	Asp	Pro	Asp	Phe	Glu	Gly	Arg	Ile	Leu	
				285					290					295			
30	ATA	ACC	CCC	AAG	GAT	GAC	AAT	GGC	CGC	TTC	AGT	GTG	TTG	ATC	ACA	GGC	1023
	Ile	Thr	Pro	Lys	Asp	Asp	Asn	Gly	Arg	Phe	Ser	Val	Leu	Ile	Thr	Gly	
			300					305					310				
35	CTG	AGG	AAG	GAG	GAT	GCA	GGG	CAC	TAC	CAG	TGT	GGA	GCC	CAC	AGT	TCT	1071
	Leu	Arg	Lys	Glu	Asp	Ala	Gly	His	Tyr	Gln	Cys	Gly	Ala	His	Ser	Ser	
		315					320					325					
40	GGT	TTG	CCT	CAA	GAA	GGC	TGG	CCC	ATC	CAG	ACT	TGG	CAA	CTC	TTT	GTC	1119
	Gly	Leu	Pro	Gln	Glu	Gly	Trp	Pro	Ile	Gln	Thr	Trp	Gln	Leu	Phe	Val	
		330				335					340					345	
	AAT	GAA	GAG	TCT	ACC	ATT	CCC	AAT	CGT	CGC	TCT	GTT	GTG	AAG	GGA	GTC	1167
	Asn	Glu	Glu	Ser	Thr	Ile	Pro	Asn	Arg	Arg	Ser	Val	Val	Lys	Gly	Val	
					350					355					360		
45	ACA	GGA	GGC	TCT	GTG	GCC	ATC	GCC	TGT	CCC	TAT	AAC	CCC	AAG	GAA	AGC	1215
	Thr	Gly	Gly	Ser	Val	Ala	Ile	Ala	Cys	Pro	Tyr	Asn	Pro	Lys	Glu	Ser	
				365					370					375			
50	AGC	AGC	CTC	AAG	TAC	TGG	TGT	CGC	TGG	GAA	GGG	GAC	GGA	AAT	GGA	CAT	1263
	Ser	Ser	Leu	Lys	Tyr	Trp	Cys	Arg	Trp	Glu	Gly	Asp	Gly	Asn	Gly	His	
			380					385					390				
55	TGC	CCC	GCG	CTT	GTG	GGG	ACC	CAG	GCC	CAG	GTG	CAA	GAA	GAG	TAT	GAA	1311
	Cys	Pro	Ala	Leu	Val	Gly	Thr	Gln	Ala	Gln	Val	Gln	Glu	Glu	Tyr	Glu	
		395					400					405					
	GGC	CGA	CTG	GCA	CTG	TTT	GAT	CAG	CCA	GGC	AAT	GGT	ACT	TAC	ACT	GTC	1359
	Gly	Arg	Leu	Ala	Leu	Phe	Asp	Gln	Pro	Gly	Asn	Gly	Thr	Tyr	Thr	Val	
		410				415					420					425	

5	ATC Ile	CTC Leu	AAC Asn	CAG Gln	CTC Leu	ACC Thr	ACC Thr	GAG Glu	GAT Asp	GCT Ala	GGC Gly	TTC Phe	TAT Tyr	TGG Trp	TGT Cys	CTT Leu	1407
					430					435					440		
	ACC Thr	AAT Asn	GGT Gly	GAC Asp	TCT Ser	CGC Arg	TGG Trp	AGA Arg	ACC Thr	ACA Thr	ATA Ile	GAA Glu	CTC Leu	CAG Gln	GTT Val	GCC Ala	1455
10	GAA Glu	GCT Ala	ACA Thr	AGG Arg	GAG Glu	CCA Pro	AAC Asn	CTT Leu	GAG Glu	GTG Val	ACG Thr	CCA Pro	CAG Gln	AAC Asn	GCA Ala	ACA Thr	1503
					460					465					470		
	GCA Ala	GTA Val	CTA Leu	GGA Gly	GAG Glu	ACC Thr	TTC Phe	ACC Thr	GTT Val	TCC Ser	TGC Cys	CAC His	TAT Tyr	CCG Pro	TGC Cys	AAA Lys	1551
15	TTC Phe	TAC Tyr	TCC Ser	CAG Gln	GAG Glu	AAA Lys	TAC Tyr	TGG Trp	TGC Cys	AAG Lys	TGG Trp	AGC Ser	AAC Asn	AAG Lys	GGT Gly	TGC Cys	1599
					490					495					500		
	CAC His	ATC Ile	CTG Leu	CCA Pro	AGC Ser	CAT His	GAC Asp	GAA Glu	GGT Gly	GCC Ala	CGC Arg	CAA Gln	TCT Ser	TCT Ser	GTG Val	AGC Ser	1647
20	TGC Cys	GAC Asp	CAG Gln	AGC Ser	AGC Ser	CAG Gln	CTG Leu	GTC Val	TCC Ser	ATG Met	ACC Thr	CTG Leu	AAC Asn	CCG Pro	GTC Val	AGT Ser	1695
					525					530					535		
	AAG Lys	GAA Glu	GAT Asp	GAA Glu	GGC Gly	TGG Trp	TAC Tyr	TGG Trp	TGT Cys	GGG Gly	GTA Val	AAG Lys	CAA Gln	GGC Gly	CAG Gln	ACC Thr	1743
25	TAT Tyr	GGA Gly	GAA Glu	ACT Thr	ACC Thr	GCC Ala	ATC Ile	TAT Tyr	ATA Ile	GCA Ala	GTT Val	GAA Glu	GAG Glu	AGG Arg	ACC Thr	AGA Arg	1791
					555					560					565		
	GGG Gly	TCA Ser	TCC Ser	CAT His	GTC Val	AAC Asn	CCA Pro	ACA Thr	GAT Asp	GCA Ala	AAT Asn	GCA Ala	CGT Arg	GCC Ala	AAA Lys	GTC Val	1839
30	GCT Ala	CTG Leu	GAA Glu	GAA Glu	GAG Glu	GTA Val	GTG Val	GAC Asp	TCC Ser	TCC Ser	ATC Ile	AGT Ser	GAA Glu	AAA Lys	GAG Glu	AAC Asn	1887
					590					595					600		
	AAA Lys	GCC Ala	ATT Ile	CCA Pro	AAT Asn	CCC Pro	GGG Gly	CCT Pro	TTT Phe	GCC Ala	AAC Asn	GAA Glu	AGA Arg	GAG Glu	ATA Ile	CAG Gln	1935
35	AAT Asn	GTG Val	AGA Arg	GAC Asp	CAA Gln	GCT Ala	CAG Gln	GAG Glu	AAC Asn	AGA Arg	GCA Ala	TCT Ser	GGG Gly	GAT Asp	GCT Ala	GGC Gly	1983
					620					625					630		
	AGT Ser	GCT Ala	GAT Asp	GGA Gly	CAA Gln	AGC Ser	AGG Arg	AGC Ser	TCC Ser	AGC Ser	TCC Ser	AAA Lys	GTG Val	CTG Leu	TTC Phe	TCC Ser	2031
40					635					640					645		

	ACC CTG GTG CCC CTG GGT CTG GTG CTG GCA GTG GGT GCT ATA GCT GTG	2079
	Thr Leu Val Pro Leu Gly Leu Val Leu Ala Val Gly Ala Ile Ala Val	
	650 655 660 665	
5	TGG GTG GCC AGA GTC CGA CAT CGG AAG AAT GTA GAC CGC ATG TCA ATC	2127
	Trp Val Ala Arg Val Arg His Arg Lys Asn Val Asp Arg Met Ser Ile	
	670 675 680	
10	AGC AGC TAC AGG ACA GAC ATT AGC ATG GCA GAC TTC AAG AAC TCC AGA	2175
	Ser Ser Tyr Arg Thr Asp Ile Ser Met Ala Asp Phe Lys Asn Ser Arg	
	685 690 695	
15	GAT TTG GGA GGC AAT GAC AAC ATG GGG GCC TCT CCA GAC ACA CAG CAA	2223
	Asp Leu Gly Gly Asn Asp Asn Met Gly Ala Ser Pro Asp Thr Gln Gln	
	700 705 710	
20	ACA GTC ATC GAA GGA AAA GAT GAA ATC GTG ACT ACC ACG GAG TGC ACC	2271
	Thr Val Ile Glu Gly Lys Asp Glu Ile Val Thr Thr Thr Glu Cys Thr	
	715 720 725	
	GCT GAG CCA GAA GAA TCC AAG AAA GCA AAA AGG TCA TCC AAG GAG GAA	2319
	Ala Glu Pro Glu Glu Ser Lys Lys Ala Lys Arg Ser Ser Lys Glu Glu	
	730 735 740 745	
25	GCT GAC ATG GCC TAC TCG GCA TTC CTG CTT CAG TCC AGC ACC ATA GCT	2367
	Ala Asp Met Ala Tyr Ser Ala Phe Leu Leu Gln Ser Ser Thr Ile Ala	
	750 755 760	
30	GCA CAG GTC CAC GAT GGT CCC CAG GAA GCC TAG GCAGTGCTGA CCACCCACCC	2420
	Ala Gln Val His Asp Gly Pro Gln Glu Ala	
	765 770	
	TTGCCTGTGA CAATCAACTT GAGAATCACA CTGATCCGCT CGCAGCCCAC ACTCACCCAT	2480
35	CACCTCCGCT CTTCCCTCCT GTCCTCAGAG GTGTGCTGGT TCCTTCCTCG GCCATGGAAG	2540
	CCTGGCCTAG TTACGCCTGT TTAGGAGAGA GTGTGAGGCG TTCTTTTCTC TATGAAGAGA	2600
40	GTGAGGTGGA AATGAGGAGG AGGTGAACCT GAGAGACATC TCTGGAGGAA GAGGGTTGAG	2660
	AATAGGGGCT CGTTTCAGGA GAAAAGGCCA TTTGAATCTT CTTTATAACC ATATGATAGG	2720
	ATGTCAGCGT AACTCTTCTC TCCTCCATCT CTCCTTTCCT ATCCTCTTGA TTCAAACAAC	2780
45	ACATCTGAGA ACTCACTAGG CTTCACTGCC TACTAAATGC TGAGAGCCAG GCCACAATCT	2840
	TTCTATAAAT ATTACTGGAA GAGATGCCAT CTCCTCCAG ATTCTGTCTT TTCATTAAGA	2900
50	TAAGACATCA TTACCAGGCA TACCTCCTGC CTCTGTGCCT CATAGGCATA CACAAGCCAT	2960
	AAGGGCATCA TGATTTTCAG ATGAGAAGAG ATGTTTCTCA AGAGTGCCTA GTGAGATAGA	3020
	CTAGCGTCAA ACCAGATGTG GCAACTCCTG GCTCTTGCC TACGATCTGT CTTCAAGAAA	3080
55	AAAAAAAAA AAAAA	3095

(2) INFORMATION FOR SEQ ID NO: 8:

## (i) SEQUENCE CHARACTERISTICS:

5 (A) LENGTH: 771 amino acids  
 (B) TYPE: amino acid  
 (C) STRANDNESS: single  
 (D) TOPOLOGY: linear  
 10 DESCRIPTION: Mouse Polyimmunoglobulin Receptor

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

Met Arg Leu Tyr Leu Phe Thr Leu Leu Val Thr Val Phe Ser Gly Val  
 1 5 10 15  
 15 Ser Thr Lys Ser Pro Ile Phe Gly Pro Gln Glu Val Ser Ser Ile Glu  
 20 25 30  
 Gly Asp Ser Val Ser Ile Thr Cys Tyr Tyr Pro Asp Thr Ser Val Asn  
 20 35 40 45  
 Arg His Thr Arg Lys Tyr Trp Cys Arg Gln Gly Ala Ser Gly Met Cys  
 50 55 60  
 25 Thr Thr Leu Ile Ser Ser Asn Gly Tyr Leu Ser Lys Glu Tyr Ser Gly  
 65 70 75 80  
 Arg Ala Asn Leu Ile Asn Phe Pro Glu Asn Asn Thr Phe Val Ile Asn  
 85 90 95  
 30 Ile Glu Gln Leu Thr Gln Asp Asp Thr Gly Ser Tyr Lys Cys Gly Leu  
 100 105 110  
 Gly Thr Ser Asn Arg Gly Leu Ser Phe Asp Val Ser Leu Glu Val Ser  
 35 115 120 125  
 Gln Val Pro Glu Leu Pro Ser Asp Thr His Val Tyr Thr Lys Asp Ile  
 130 135 140  
 40 Gly Arg Asn Val Thr Ile Glu Cys Pro Phe Lys Arg Glu Asn Val Pro  
 145 150 155 160  
 Ser Lys Lys Ser Leu Cys Lys Lys Thr Asn Gln Ser Cys Glu Leu Val  
 165 170 175  
 45 Ile Asp Ser Thr Glu Lys Val Asn Pro Ser Tyr Ile Gly Arg Ala Lys  
 180 185 190  
 Leu Phe Met Lys Gly Thr Asp Leu Thr Val Phe Tyr Val Asn Ile Ser  
 195 200 205  
 50 His Leu Thr His Asn Asp Ala Gly Leu Tyr Ile Cys Gln Ala Gly Glu  
 210 215 220  
 55 Gly Pro Ser Ala Asp Lys Lys Asn Val Asp Leu Gln Val Leu Ala Pro  
 225 230 235 240  
 Glu Pro Glu Leu Leu Tyr Lys Asp Leu Arg Ser Ser Val Thr Phe Glu  
 245 250 255

	Cys	Asp	Leu	Gly	Arg	Glu	Val	Ala	Asn	Glu	Ala	Lys	Tyr	Leu	Cys	Arg
				260					265					270		
5	Met	Asn	Lys	Glu	Thr	Cys	Asp	Val	Ile	Ile	Asn	Thr	Leu	Gly	Lys	Arg
			275					280					285			
	Asp	Pro	Asp	Phe	Glu	Gly	Arg	Ile	Leu	Ile	Thr	Pro	Lys	Asp	Asp	Asn
10		290					295					300				
	Gly	Arg	Phe	Ser	Val	Leu	Ile	Thr	Gly	Leu	Arg	Lys	Glu	Asp	Ala	Gly
	305					310					315					320
	His	Tyr	Gln	Cys	Gly	Ala	His	Ser	Ser	Gly	Leu	Pro	Gln	Glu	Gly	Trp
15					325					330					335	
	Pro	Ile	Gln	Thr	Trp	Gln	Leu	Phe	Val	Asn	Glu	Glu	Ser	Thr	Ile	Pro
				340					345					350		
20	Asn	Arg	Arg	Ser	Val	Val	Lys	Gly	Val	Thr	Gly	Gly	Ser	Val	Ala	Ile
			355					360					365			
	Ala	Cys	Pro	Tyr	Asn	Pro	Lys	Glu	Ser	Ser	Ser	Leu	Lys	Tyr	Trp	Cys
25		370					375					380				
	Arg	Trp	Glu	Gly	Asp	Gly	Asn	Gly	His	Cys	Pro	Ala	Leu	Val	Gly	Thr
	385					390				395						400
	Gln	Ala	Gln	Val	Gln	Glu	Glu	Tyr	Glu	Gly	Arg	Leu	Ala	Leu	Phe	Asp
30					405					410					415	
	Gln	Pro	Gly	Asn	Gly	Thr	Tyr	Thr	Val	Ile	Leu	Asn	Gln	Leu	Thr	Thr
				420					425					430		
35	Glu	Asp	Ala	Gly	Phe	Tyr	Trp	Cys	Leu	Thr	Asn	Gly	Asp	Ser	Arg	Trp
			435					440					445			
	Arg	Thr	Thr	Ile	Glu	Leu	Gln	Val	Ala	Glu	Ala	Thr	Arg	Glu	Pro	Asn
40		450					455					460				
	Leu	Glu	Val	Thr	Pro	Gln	Asn	Ala	Thr	Ala	Val	Leu	Gly	Glu	Thr	Phe
	465					470					475					480
	Thr	Val	Ser	Cys	His	Tyr	Pro	Cys	Lys	Phe	Tyr	Ser	Gln	Glu	Lys	Tyr
45					485					490					495	
	Trp	Cys	Lys	Trp	Ser	Asn	Lys	Gly	Cys	His	Ile	Leu	Pro	Ser	His	Asp
				500					505					510		
50	Glu	Gly	Ala	Arg	Gln	Ser	Ser	Val	Ser	Cys	Asp	Gln	Ser	Ser	Gln	Leu
			515					520					525			
	Val	Ser	Met	Thr	Leu	Asn	Pro	Val	Ser	Lys	Glu	Asp	Glu	Gly	Trp	Tyr
		530					535					540				
55	Trp	Cys	Gly	Val	Lys	Gln	Gly	Gln	Thr	Tyr	Gly	Glu	Thr	Thr	Ala	Ile
	545					550					555					560

Tyr Ile Ala Val Glu Glu Arg Thr Arg Gly Ser Ser His Val Asn Pro  
 565 570 575  
 5 Thr Asp Ala Asn Ala Arg Ala Lys Val Ala Leu Glu Glu Glu Val Val  
 580 585 590  
 Asp Ser Ser Ile Ser Glu Lys Glu Asn Lys Ala Ile Pro Asn Pro Gly  
 595 600 605  
 10 Pro Phe Ala Asn Glu Arg Glu Ile Gln Asn Val Arg Asp Gln Ala Gln  
 610 615 620  
 Glu Asn Arg Ala Ser Gly Asp Ala Gly Ser Ala Asp Gly Gln Ser Arg  
 625 630 635 640  
 15 Ser Ser Ser Ser Lys Val Leu Phe Ser Thr Leu Val Pro Leu Gly Leu  
 645 650 655  
 Val Leu Ala Val Gly Ala Ile Ala Val Trp Val Ala Arg Val Arg His  
 660 665 670  
 20 Arg Lys Asn Val Asp Arg Met Ser Ile Ser Ser Tyr Arg Thr Asp Ile  
 675 680 685  
 Ser Met Ala Asp Phe Lys Asn Ser Arg Asp Leu Gly Gly Asn Asp Asn  
 690 695 700  
 Met Gly Ala Ser Pro Asp Thr Gln Gln Thr Val Ile Glu Gly Lys Asp  
 705 710 715 720  
 30 Glu Ile Val Thr Thr Thr Glu Cys Thr Ala Glu Pro Glu Glu Ser Lys  
 725 730 735  
 Lys Ala Lys Arg Ser Ser Lys Glu Glu Ala Asp Met Ala Tyr Ser Ala  
 740 745 750  
 35 Phe Leu Leu Gln Ser Ser Thr Ile Ala Ala Gln Val His Asp Gly Pro  
 755 760 765  
 40 Gln Glu Ala  
 770

45 (2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

50 (A) LENGTH: 3269 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear  
 DESCRIPTION: Rat Polyimmunoglobulin Receptor

55 (ix) FEATURE:

(A) NAME/KEY: Coding Sequence  
 (B) LOCATION: 74....2383



## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

	GGCAACGAAG GTACCATGGA TCTTATACAA GAAGTGAACC AACATGCCGC AACCTCCTTG	60
5	GAAGCCACAA GCG ATG AGG CTC TCC TTG TTC GCC CTC TTG GTA ACT GTC Met Arg Leu Ser Leu Phe Ala Leu Leu Val Thr Val 1 5 10	109
10	TTC TCA GGG GTC TCC ACA CAA AGC CCC ATA TTT GGT CCC CAG GAT GTG Phe Ser Gly Val Ser Thr Gln Ser Pro Ile Phe Gly Pro Gln Asp Val 15 20 25	157
15	AGT AGT ATT GAA GGT AAC TCG GTC TCC ATC ACG TGC TAC TAC CCA GAC Ser Ser Ile Glu Gly Asn Ser Val Ser Ile Thr Cys Tyr Tyr Pro Asp 30 35 40	205
20	ACC TCT GTC AAC CGG CAC ACC CGG AAA TAC TGG TGC CGA CAA GGA GCC Thr Ser Val Asn Arg His Thr Arg Lys Tyr Trp Cys Arg Gln Gly Ala 45 50 55 60	253
	AAC GGC TAC TGC GCA ACC CTC ATC TCT TCA AAT GGC TAC CTC TCG AAG Asn Gly Tyr Cys Ala Thr Leu Ile Ser Ser Asn Gly Tyr Leu Ser Lys 65 70 75	301
25	GAG TAT TCA GGC AGA GCC AGC CTC ATC AAC TTC CCA GAG AAT AGC ACA Glu Tyr Ser Gly Arg Ala Ser Leu Ile Asn Phe Pro Glu Asn Ser Thr 80 85 90	349
30	TTT GTG ATT AAC ATT GCA CAT CTC ACC CAG GAG GAC ACT GGG AGC TAC Phe Val Ile Asn Ile Ala His Leu Thr Gln Glu Asp Thr Gly Ser Tyr 95 100 105	397
35	AAG TGT GGT CTG GGT ACC ACT AAC CGA GGC CTG TTT TTC GAT GTC AGC Lys Cys Gly Leu Gly Thr Thr Asn Arg Gly Leu Phe Phe Asp Val Ser 110 115 120	445
40	CTG GAG GTC AGC CAG GTT CCT GAG TTC CCA AAT GAC ACC CAT GTC TAC Leu Glu Val Ser Gln Val Pro Glu Phe Pro Asn Asp Thr His Val Tyr 125 130 135 140	493
	ACA AAG GAC ATA GGC AGA ACT GTG ACC ATC GAA TGC CGT TTC AAA GAG Thr Lys Asp Ile Gly Arg Thr Val Thr Ile Glu Cys Arg Phe Lys Glu 145 150 155	541
45	GGG AAT GCT CAT AGC AAG AAA TCC CTG TGT AAG AAG AGA GGA GAG GCC Gly Asn Ala His Ser Lys Lys Ser Leu Cys Lys Lys Arg Gly Glu Ala 160 165 170	589
50	TGC GAA GTT GTC ATC GAC TCT ACT GAG TAC GTG GAC CCC AGC TAT AAG Cys Glu Val Val Ile Asp Ser Thr Glu Tyr Val Asp Pro Ser Tyr Lys 175 180 185	637
55	GAC AGA GCA ATC CTT TTT ATG AAA GGG ACC AGC CGC GAT ATA TTC TAT Asp Arg Ala Ile Leu Phe Met Lys Gly Thr Ser Arg Asp Ile Phe Tyr 190 195 200	685
	GTC AAC ATT AGC CAC CTA ATA CCC AGT GAT GCT GGA CTG TAT GTT TGC Val Asn Ile Ser His Leu Ile Pro Ser Asp Ala Gly Leu Tyr Val Cys 205 210 215 220	733

	CAA	GCT	GGA	GAA	GGC	CCC	AGT	GCT	GAT	AAA	AAT	AAT	GCT	GAC	CTC	CAG	781
	Gln	Ala	Gly	Glu	Gly	Pro	Ser	Ala	Asp	Lys	Asn	Asn	Ala	Asp	Leu	Gln	
					225					230					235		
5	GTG	CTA	GAG	CCT	GAG	CCA	GAG	CTG	CTT	TAT	AAA	GAC	CTG	AGG	TCC	TCA	829
	Val	Leu	Glu	Pro	Glu	Pro	Glu	Leu	Leu	Tyr	Lys	Asp	Leu	Arg	Ser	Ser	
				240				245					250				
10	GTG	ACT	TTT	GAA	TGT	GAC	CTG	GGC	CGT	GAA	GTG	GCA	AAT	GAT	GCC	AAA	877
	Val	Thr	Phe	Glu	Cys	Asp	Leu	Gly	Arg	Glu	Val	Ala	Asn	Asp	Ala	Lys	
			255				260					265					
15	TAT	CTG	TGT	CGG	AAG	AAC	AAG	GAA	ACC	TGT	GAT	GTC	ATC	ATC	AAC	ACC	925
	Tyr	Leu	Cys	Arg	Lys	Asn	Lys	Glu	Thr	Cys	Asp	Val	Ile	Ile	Asn	Thr	
		270					275					280					
20	CTG	GGG	AAG	AGA	GAT	CCA	GCC	TTT	GAA	GGC	AGG	ATC	CTG	CTA	ACC	CCC	973
	Leu	Gly	Lys	Arg	Asp	Pro	Ala	Phe	Glu	Gly	Arg	Ile	Leu	Leu	Thr	Pro	
	285					290				295						300	
25	AGG	GAT	GAC	AAT	GGC	CGC	TTC	AGT	GTG	TTG	ATC	ACA	GGC	CTG	AGG	AAG	1021
	Arg	Asp	Asp	Asn	Gly	Arg	Phe	Ser	Val	Leu	Ile	Thr	Gly	Leu	Arg	Lys	
				305					310						315		
30	GAG	GAT	GCA	GGG	CAC	TAC	CAG	TGT	GGA	GCG	CAC	AGT	TCT	GGT	TTG	CCT	1069
	Glu	Asp	Ala	Gly	His	Tyr	Gln	Cys	Gly	Ala	His	Ser	Ser	Gly	Leu	Pro	
				320				325					330				
35	CAA	GAA	GGC	TGG	CCC	GTC	CAG	GCT	TGG	CAA	CTC	TTT	GTC	AAT	GAA	GAG	1117
	Gln	Glu	Gly	Trp	Pro	Val	Gln	Ala	Trp	Gln	Leu	Phe	Val	Asn	Glu	Glu	
			335				340					345					
40	TCC	ACG	ATT	CCC	AAT	AGT	CGC	TCT	GTT	GTG	AAG	GGT	GTC	ACA	GGA	GGC	1165
	Ser	Thr	Ile	Pro	Asn	Ser	Arg	Ser	Val	Val	Lys	Gly	Val	Thr	Gly	Gly	
		350				355						360					
45	TCT	GTG	GCC	ATC	GTC	TGT	CCC	TAT	AAC	CCC	AAG	GAA	AGC	AGC	AGC	CTC	1213
	Ser	Val	Ala	Ile	Val	Cys	Pro	Tyr	Asn	Pro	Lys	Glu	Ser	Ser	Ser	Leu	
	365				370					375						380	
50	AAG	TAC	TGG	TGT	CAC	TGG	GAA	GCC	GAC	GAG	AAT	GGA	CGC	TGC	CCG	GTG	1261
	Lys	Tyr	Trp	Cys	His	Trp	Glu	Ala	Asp	Glu	Asn	Gly	Arg	Cys	Pro	Val	
				385					390					395			
55	CTC	GTG	GGG	ACC	CAG	GCC	CTG	GTG	CAA	GAA	GGA	TAT	GAA	GGC	CGA	CTG	1309
	Leu	Val	Gly	Thr	Gln	Ala	Leu	Val	Gln	Glu	Gly	Tyr	Glu	Gly	Arg	Leu	
				400				405					410				
60	GCA	CTG	TTC	GAT	CAG	CCG	GGC	AGT	GGC	GCC	TAC	ACT	GTC	ATC	CTC	AAC	1357
	Ala	Leu	Phe	Asp	Gln	Pro	Gly	Ser	Gly	Ala	Tyr	Thr	Val	Ile	Leu	Asn	
			415				420					425					
65	CAG	CTC	ACC	ACC	CAG	GAT	TCT	GGC	TTC	TAC	TGG	TGT	CTT	ACC	GAT	GGT	1405
	Gln	Leu	Thr	Thr	Gln	Asp	Ser	Gly	Phe	Tyr	Trp	Cys	Leu	Thr	Asp	Gly	
		430				435						440					

	GAC	TCT	CGC	TGG	AGA	ACC	ACG	ATA	GAA	CTG	CAG	GTT	GCT	GAA	GCT	ACA	1453
	Asp	Ser	Arg	Trp	Arg	Thr	Thr	Ile	Glu	Leu	Gln	Val	Ala	Glu	Ala	Thr	
	445					450				455						460	
5	AAG	AAG	CCA	GAC	CTT	GAG	GTG	ACA	CCA	CAG	AAC	GCG	ACC	GCG	GTG	ATA	1501
	Lys	Lys	Pro	Asp	Leu	Glu	Val	Thr	Pro	Gln	Asn	Ala	Thr	Ala	Val	Ile	
				465					470						475		
10	GGA	GAG	ACC	TTC	ACA	ATC	TCC	TGC	CAC	TAT	CCG	TGC	AAA	TTC	TAC	TCC	1549
	Gly	Glu	Thr	Phe	Thr	Ile	Ser	Cys	His	Tyr	Pro	Cys	Lys	Phe	Tyr	Ser	
				480				485						490			
15	CAG	GAG	AAA	TAC	TGG	TGC	AAG	TGG	AGC	AAC	GAC	GGC	TGC	CAC	ATC	CTG	1597
	Gln	Glu	Lys	Tyr	Trp	Cys	Lys	Trp	Ser	Asn	Asp	Gly	Cys	His	Ile	Leu	
			495				500					505					
20	CCG	AGC	CAT	GAT	GAA	GGT	GCC	CGC	CAG	TCC	TCT	GTG	AGC	TGT	GAC	CAG	1645
	Pro	Ser	His	Asp	Glu	Gly	Ala	Arg	Gln	Ser	Ser	Val	Ser	Cys	Asp	Gln	
		510					515					520					
	AGC	AGC	CAG	ATC	GTC	TCC	ATG	ACC	CTG	AAC	CCG	GTC	AAA	AAG	GAA	GAT	1693
	Ser	Ser	Gln	Ile	Val	Ser	Met	Thr	Leu	Asn	Pro	Val	Lys	Lys	Glu	Asp	
	525					530				535						540	
25	GAA	GGC	TGG	TAC	TGG	TGT	GGG	GTA	AAA	GAA	GGT	CAG	GTC	TAT	GGA	GAA	1741
	Glu	Gly	Trp	Tyr	Trp	Cys	Gly	Val	Lys	Glu	Gly	Gln	Val	Tyr	Gly	Glu	
					545					550					555		
30	ACT	ACA	GCC	ATC	TAT	GTA	GCA	GTT	GAA	GAG	AGG	ACC	AGA	GGG	TCA	CCC	1789
	Thr	Thr	Ala	Ile	Tyr	Val	Ala	Val	Glu	Glu	Arg	Thr	Arg	Gly	Ser	Pro	
				560					565					570			
35	CAC	ATC	AAC	CCG	ACA	GAT	GCA	AAC	GCA	CGT	GCA	AAA	GAT	GCT	CCA	GAG	1837
	His	Ile	Asn	Pro	Thr	Asp	Ala	Asn	Ala	Arg	Ala	Lys	Asp	Ala	Pro	Glu	
			575					580					585				
40	GAA	GAG	GCA	ATG	GAA	TCC	TCT	GTC	AGG	GAG	GAT	GAA	AAC	AAG	GCC	AAT	1885
	Glu	Glu	Ala	Met	Glu	Ser	Ser	Val	Arg	Glu	Asp	Glu	Asn	Lys	Ala	Asn	
		590					595					600					
	CTG	GAC	CCC	AGG	CTT	TTT	GCA	GAC	GAA	AGA	GAG	ATA	CAG	AAT	GCG	GGA	1933
	Leu	Asp	Pro	Arg	Leu	Phe	Ala	Asp	Glu	Arg	Glu	Ile	Gln	Asn	Ala	Gly	
	605				610					615						620	
45	GAC	CAA	GCT	CAG	GAG	AAC	AGA	GCA	TCT	GGG	AAT	GCT	GGC	AGT	GCT	GGT	1981
	Asp	Gln	Ala	Gln	Glu	Asn	Arg	Ala	Ser	Gly	Asn	Ala	Gly	Ser	Ala	Gly	
					625					630					635		
50	GGA	CAA	AGC	GGG	AGC	TCC	AAA	GTC	CTA	TTC	TCC	ACC	CTG	GTG	CCC	CTG	2029
	Gly	Gln	Ser	Gly	Ser	Ser	Lys	Val	Leu	Phe	Ser	Thr	Leu	Val	Pro	Leu	
				640					645					650			
55	GGT	TTG	GTG	CTG	GCA	GTG	GGT	GCT	GTG	GCT	GTG	TGG	GTG	GCC	AGA	GTC	2077
	Gly	Leu	Val	Leu	Ala	Val	Gly	Ala	Val	Ala	Val	Trp	Val	Ala	Arg	Val	
			655				660						665				
	CGA	CAT	CGG	AAG	AAT	GTA	GAC	CGC	ATG	TCA	ATC	AGC	AGC	TAC	AGG	ACA	2125
	Arg	His	Arg	Lys	Asn	Val	Asp	Arg	Met	Ser	Ile	Ser	Ser	Tyr	Arg	Thr	
		670					675					680					

5	GAC ATT AGC ATG GGA GAC TTC AGG AAC TCC AGG GAT TTG GGA GGC AAT Asp Ile Ser Met Gly Asp Phe Arg Asn Ser Arg Asp Leu Gly Gly Asn 685 690 695 700	2173
10	GAC AAC ATG GGC GCC ACT CCA GAC ACA CAA GAA ACA GTC CTC GAA GGA Asp Asn Met Gly Ala Thr Pro Asp Thr Gln Glu Thr Val Leu Glu Gly 705 710 715	2221
15	AAA GAT GAA ATA GAG ACT ACC ACC GAG TGT ACC ACC GAG CCA GAG GAA Lys Asp Glu Ile Glu Thr Thr Thr Glu Cys Thr Thr Glu Pro Glu Glu 720 725 730	2269
20	TCC AAG AAA GCA AAA AGG TCA TCC AAG GAG GAA GCT GAC ATG GCC TAC Ser Lys Lys Ala Lys Arg Ser Ser Lys Glu Glu Ala Asp Met Ala Tyr 735 740 745	2317
25	TCA GCA TTC CTG TTT CAG TCC AGC ACA ATA GCT GCG CAG GTC CAT GAT Ser Ala Phe Leu Phe Gln Ser Ser Thr Ile Ala Ala Gln Val His Asp 750 755 760	2365
30	GGT CCC CAG GAA GCC TAG GCAGTGCTGA CCACCTACCC CTGCCTGTGA CAATCAACT Gly Pro Gln Glu Ala 765	2422
35	TGAGAATCAC ATTGATCCAC TCGCAGCCCA CCCTCGCCCA TCACCCAGGC TCTTCCCTCC	2482
40	TGTTCTCAGA GGTGTGCTGG TTCCTCCCTC AGTCGTGGAA GCCTGGCCTA CTTATGCCTG	2542
45	TTTAGGAGAG AGCGTGAGGA GTTCTTTTTG CTGTTAAAGA GTAAGGTGGA AATGAGTTGA	2602
50	GCCCAAGAGG TGTCTCTGAG AGACGAGGGT TCAGAGCAGG GGCTCATTTT AGGAGGAAGA	2662
55	GCCATTTGAA GCCTCTTTAT ACACATATGC TAGGATGTCA GGATAGCTCT TCTCCTCCAT	2722
60	CTCTCCTTTC TTCTCTTCTT GATTGAGACA ACAGATCCGA AAACCTCACTA GGCTTCCGGT	2782
65	GTCTACTAAA TGCTGAGAGT CAGGCCACAG CCTTCTATA AACATCACTG GAAGAGACAC	2842
70	CACCTCGTCC CAGATTCTGT CTTTTCCCTA AGCTATCAAT CATTACCGGG GATTCCCTTT	2902
75	GCCTCTGCAC CTCATAGGCA ACAAAAGAAA CATAAGTCCT GCAGTCTAAG GCATACCCAA	2962
80	GCCATAAGGG CACCACGAGA CTCAGATGAG AAGAGATTTT TCTCCAGAGT ACTCAGTGAG	3022
85	ATAGACTAGT GTCAAGCCAG ATGGGGCAAC TCCTGGCTCT TGGCCTGGGA CTTGTCTTCA	3082
90	AGATCTCTGC TCTTATTAGA GAAAGAACTT TAGCATGAGG AAAAGTAAGA GAAAACAAGT	3142
95	TACATGGGCA TGGTGGTGTG CTCCTGCAAT CCCAATATTA AGAGGTAAA AAATAGGACC	3202
100	AGAAGTTTAA AGTAATCCTT GGCTACCTAG TGAGTGTAAG GCCAGCCTGG AATCAATAAG	3262
105	AGTTGGT	3269

## (2) INFORMATION FOR SEQ ID NO: 10:

## (i) SEQUENCE CHARACTERISTICS:

5 (A) LENGTH: 770 amino acids  
 (B) TYPE: amino acid  
 (C) STRANDNESS: single  
 (D) TOPOLOGY: linear  
 10 DESCRIPTION: Rat Polyimmunoglobulin Receptor

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

Met Arg Leu Ser Leu Phe Ala Leu Leu Val Thr Val Phe Ser Gly Val  
 1 5 10 15  
 15 Ser Thr Gln Ser Pro Ile Phe Gly Pro Gln Asp Val Ser Ser Ile Glu  
 20 25 30  
 Gly Asn Ser Val Ser Ile Thr Cys Tyr Tyr Pro Asp Thr Ser Val Asn  
 20 35 40 45  
 Arg His Thr Arg Lys Tyr Trp Cys Arg Gln Gly Ala Asn Gly Tyr Cys  
 50 55 60  
 25 Ala Thr Leu Ile Ser Ser Asn Gly Tyr Leu Ser Lys Glu Tyr Ser Gly  
 65 70 75 80  
 Arg Ala Ser Leu Ile Asn Phe Pro Glu Asn Ser Thr Phe Val Ile Asn  
 85 90 95  
 30 Ile Ala His Leu Thr Gln Glu Asp Thr Gly Ser Tyr Lys Cys Gly Leu  
 100 105 110  
 Gly Thr Thr Asn Arg Gly Leu Phe Phe Asp Val Ser Leu Glu Val Ser  
 115 120 125  
 35 Gln Val Pro Glu Phe Pro Asn Asp Thr His Val Tyr Thr Lys Asp Ile  
 130 135 140  
 40 Gly Arg Thr Val Thr Ile Glu Cys Arg Phe Lys Glu Gly Asn Ala His  
 145 150 155 160  
 Ser Lys Lys Ser Leu Cys Lys Lys Arg Gly Glu Ala Cys Glu Val Val  
 165 170 175  
 45 Ile Asp Ser Thr Glu Tyr Val Asp Pro Ser Tyr Lys Asp Arg Ala Ile  
 180 185 190  
 Leu Phe Met Lys Gly Thr Ser Arg Asp Ile Phe Tyr Val Asn Ile Ser  
 195 200 205  
 50 His Leu Ile Pro Ser Asp Ala Gly Leu Tyr Val Cys Gln Ala Gly Glu  
 210 215 220  
 55 Gly Pro Ser Ala Asp Lys Asn Asn Ala Asp Leu Gln Val Leu Glu Pro  
 225 230 235 240  
 Glu Pro Glu Leu Leu Tyr Lys Asp Leu Arg Ser Ser Val Thr Phe Glu  
 245 250 255

Cys Asp Leu Gly Arg Glu Val Ala Asn Asp Ala Lys Tyr Leu Cys Arg  
 260 265 270  
 5 Lys Asn Lys Glu Thr Cys Asp Val Ile Ile Asn Thr Leu Gly Lys Arg  
 275 280 285  
 Asp Pro Ala Phe Glu Gly Arg Ile Leu Leu Thr Pro Arg Asp Asp Asn  
 290 295 300  
 10 Gly Arg Phe Ser Val Leu Ile Thr Gly Leu Arg Lys Glu Asp Ala Gly  
 305 310 315 320  
 15 His Tyr Gln Cys Gly Ala His Ser Ser Gly Leu Pro Gln Glu Gly Trp  
 325 330 335  
 Pro Val Gln Ala Trp Gln Leu Phe Val Asn Glu Glu Ser Thr Ile Pro  
 340 345 350  
 20 Asn Ser Arg Ser Val Val Lys Gly Val Thr Gly Gly Ser Val Ala Ile  
 355 360 365  
 Val Cys Pro Tyr Asn Pro Lys Glu Ser Ser Ser Leu Lys Tyr Trp Cys  
 370 375 380  
 25 His Trp Glu Ala Asp Glu Asn Gly Arg Cys Pro Val Leu Val Gly Thr  
 385 390 395 400  
 30 Gln Ala Leu Val Gln Glu Gly Tyr Glu Gly Arg Leu Ala Leu Phe Asp  
 405 410 415  
 Gln Pro Gly Ser Gly Ala Tyr Thr Val Ile Leu Asn Gln Leu Thr Thr  
 420 425 430  
 35 Gln Asp Ser Gly Phe Tyr Trp Cys Leu Thr Asp Gly Asp Ser Arg Trp  
 435 440 445  
 Arg Thr Thr Ile Glu Leu Gln Val Ala Glu Ala Thr Lys Lys Pro Asp  
 450 455 460  
 Leu Glu Val Thr Pro Gln Asn Ala Thr Ala Val Ile Gly Glu Thr Phe  
 465 470 475 480  
 45 Thr Ile Ser Cys His Tyr Pro Cys Lys Phe Tyr Ser Gln Glu Lys Tyr  
 485 490 495  
 Trp Cys Lys Trp Ser Asn Asp Gly Cys His Ile Leu Pro Ser His Asp  
 500 505 510  
 50 Glu Gly Ala Arg Gln Ser Ser Val Ser Cys Asp Gln Ser Ser Gln Ile  
 515 520 525  
 Val Ser Met Thr Leu Asn Pro Val Lys Lys Glu Asp Glu Gly Trp Tyr  
 530 535 540  
 55 Trp Cys Gly Val Lys Glu Gly Gln Val Tyr Gly Glu Thr Thr Ala Ile  
 545 550 555 560

(2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 322 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

DESCRIPTION: Guy's 13 Kappa

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 8....320

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

5	CTCGAGC	GAC	ATT	GTG	ATG	ACC	CAG	TCT	CCA	GCA	ATC	ATG	TCT	GCA	TCT	49
	Asp	Ile	Val	Met	Thr	Gln	Ser	Pro	Ala	Ile	Met	Ser	Ala	Ser		
	1				5					10						
10	CCA	GGG	GAG	AAG	GTC	ACC	ATA	ACC	TGC	AGT	GCC	AGC	TCA	AGT	GTA	AGT
	Pro	Gly	Glu	Lys	Val	Thr	Ile	Thr	Cys	Ser	Ala	Ser	Ser	Ser	Val	Ser
	15				20					25					30	
15	TAC	ATG	CAC	TGG	TTC	CAG	CAG	AAG	CCA	GGC	ACT	TCT	CCC	AAA	CTC	TGG
	Tyr	Met	His	Trp	Phe	Gln	Gln	Lys	Pro	Gly	Thr	Ser	Pro	Lys	Leu	Trp
				35					40					45		
20	CTT	TAT	AGC	ACA	TCC	AAC	CTG	GCT	TCT	GGA	GTC	CCT	GCT	CGC	TTC	AGT
	Leu	Tyr	Ser	Thr	Ser	Asn	Leu	Ala	Ser	Gly	Val	Pro	Ala	Arg	Phe	Ser
			50					55					60			
25	GGC	AGT	GGA	TCT	GGG	ACC	TCT	TAC	TCT	CTC	ACA	ATC	AGC	CGA	ATG	GAG
	Gly	Ser	Gly	Ser	Gly	Thr	Ser	Tyr	Ser	Leu	Thr	Ile	Ser	Arg	Met	Glu
		65				70						75				
30	GCT	GAA	GAT	GCT	GCC	ACT	TAT	TAC	TGC	CAT	CAA	AGG	ACT	AGT	TAC	CCG
	Ala	Glu	Asp	Ala	Ala	Thr	Tyr	Tyr	Cys	His	Gln	Arg	Thr	Ser	Tyr	Pro
		80				85					90					
35	TAC	ACG	TTC	GGA	GGG	GGG	ACC	AAG	CTG	GAA	A	TA				
	Tyr	Thr	Phe	Gly	Gly	Gly	Thr	Lys	Leu	Glu	Ile					
	95				100					105						

## (2) INFORMATION FOR SEQ ID NO: 12:

## (i) SEQUENCE CHARACTERISTICS:

35	(A)	LENGTH:	105 amino acids
	(B)	TYPE:	amino acid
	(C)	STRANDNESS:	single
	(D)	TOPOLOGY:	linear
40		DESCRIPTION:	Guy's 13 Kappa

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

45	Asp	Ile	Val	Met	Thr	Gln	Ser	Pro	Ala	Ile	Met	Ser	Ala	Ser	Pro	Gly
	1				5					10					15	
	Glu	Lys	Val	Thr	Ile	Thr	Cys	Ser	Ala	Ser	Ser	Ser	Val	Ser	Tyr	Met
			20						25					30		
50	His	Trp	Phe	Gln	Gln	Lys	Pro	Gly	Thr	Ser	Pro	Lys	Leu	Trp	Leu	Tyr
			35					40					45			
	Ser	Thr	Ser	Asn	Leu	Ala	Ser	Gly	Val	Pro	Ala	Arg	Phe	Ser	Gly	Ser
		50					55					60				
55	Gly	Ser	Gly	Thr	Ser	Tyr	Ser	Leu	Thr	Ile	Ser	Arg	Met	Glu	Ala	Glu
		65				70					75				80	



Asp Ala Ala Thr Tyr Tyr Cys His Gln Arg Thr Ser Tyr Pro Tyr Thr  
85 90 95

5 Phe Gly Gly Gly Thr Lys Leu Glu Ile  
100 105

10 (2) INFORMATION FOR SEQ ID NO: 13:

(i) SEQUENCE CHARACTERISTICS:

15 (A) LENGTH: 402 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear  
DESCRIPTION: Guy's 13 Gamma 1

20 (ix) FEATURE:

(A) NAME/KEY: Coding Sequence  
(B) LOCATION: 7...402

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

CTCGAG ATG GAA TGG ACC TGG GTT TTT CTC TTC CTC CTG TCA GGA ACT	48
Met Glu Trp Thr Trp Val Phe Leu Phe Leu Leu Ser Gly Thr	
1 5 10	
GCA GGC GTC CAC TCT GGG GTC CAG CTT CAG CAG TCA GGA CCT GAC CTG	96
Ala Gly Val His Ser Gly Val Gln Leu Gln Gln Ser Gly Pro Asp Leu	
15 20 25 30	
GTG AAA CCT GGG GCC TCA GTG AAG ATA TCC TGC AAG GCT TCT GGA TAC	144
Val Lys Pro Gly Ala Ser Val Lys Ile Ser Cys Lys Ala Ser Gly Tyr	
35 40 45	
ACA TTC ACT GAC TAC AAC ATA CAC TGG GTG AAG CAG AGC CGT GGA AAG	192
Thr Phe Thr Asp Tyr Asn Ile His Trp Val Lys Gln Ser Arg Gly Lys	
50 55 60	
AGC CTT GAG TGG ATT GGA TAT ATT TAT CCT TAC AAT GGT AAT ACT TAC	240
Ser Leu Glu Trp Ile Gly Tyr Ile Tyr Pro Tyr Asn Gly Asn Thr Tyr	
65 70 75	
TAC AAC CAG AAG TTC AAG AAC AAG GCC ACA TTG ACT GTA GAC AAT TCC	288
Tyr Asn Gln Lys Phe Lys Asn Lys Ala Thr Leu Thr Val Asp Asn Ser	
80 85 90	
TCC ACC TCA GCC TAC ATG GAG CTC CGC AGC CTG ACA TCT GAG GAC TCT	336
Ser Thr Ser Ala Tyr Met Glu Leu Arg Ser Leu Thr Ser Glu Asp Ser	
95 100 105 110	
GCA GTC TAT TAC TGT GCA ACC TAC TTT GAC TAC TGG GGC CAA GGC ACC	384
Ala Val Tyr Tyr Cys Ala Thr Tyr Phe Asp Tyr Trp Gly Gln Gly Thr	
115 120 125	

ACT CTC ACA GTC TCC TCA  
Thr Leu Thr Val Ser Ser  
130

402

5

(2) INFORMATION FOR SEQ ID NO: 14:

10 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 132 amino acids  
(B) TYPE: amino acid  
(C) STRANDNESS: single  
15 (D) TOPOLOGY: linear  
DESCRIPTION: Guy's 13 Gamma 1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

20 Met Glu Trp Thr Trp Val Phe Leu Phe Leu Leu Ser Gly Thr Ala Gly  
1 5 10 15  
Val His Ser Gly Val Gln Leu Gln Gln Ser Gly Pro Asp Leu Val Lys  
20 25 30  
25 Pro Gly Ala Ser Val Lys Ile Ser Cys Lys Ala Ser Gly Tyr Thr Phe  
35 40 45  
30 Thr Asp Tyr Asn Ile His Trp Val Lys Gln Ser Arg Gly Lys Ser Leu  
50 55 60  
Glu Trp Ile Gly Tyr Ile Tyr Pro Tyr Asn Gly Asn Thr Tyr Tyr Asn  
65 70 75 80  
35 Gln Lys Phe Lys Asn Lys Ala Thr Leu Thr Val Asp Asn Ser Ser Thr  
85 90 95  
Ser Ala Tyr Met Glu Leu Arg Ser Leu Thr Ser Glu Asp Ser Ala Val  
100 105 110  
40 Tyr Tyr Cys Ala Thr Tyr Phe Asp Tyr Trp Gly Gln Gly Thr Thr Leu  
115 120 125  
45 Thr Val Ser Ser  
130

50

(2) INFORMATION FOR SEQ ID NO: 15:

(i) SEQUENCE CHARACTERISTICS:

55 (A) LENGTH: 31 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

ACCAGATCTA TGGAATGGAC CTGGGTTTTT C 31

5

10 (2) INFORMATION FOR SEQ ID NO: 16:

(i) SEQUENCE CHARACTERISTICS:

15 (A) LENGTH: 30 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

20 CCCAAGCTTG GTTTTGGAGA TGGTTTTCTC 30

25

(2) INFORMATION FOR SEQ ID NO: 17:

(i) SEQUENCE CHARACTERISTICS:

30 (A) LENGTH: 31 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

35

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

GATAAGCTTG GTCCTACTCC TCCTCCTCCT A 31

40

(2) INFORMATION FOR SEQ ID NO: 18:

(i) SEQUENCE CHARACTERISTICS:

45 (A) LENGTH: 30 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

AATCTCGAGT CAGTAGCAGA TGCCATCTCC 30

55

(2) INFORMATION FOR SEQ ID NO: 19:

(i) SEQUENCE CHARACTERISTICS:

5

(A) LENGTH: 30 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

GGAAAGCTTT GTACATATGC AAGGCTTACA

30

CLAIMS

We claim:

- 5           95.    An immunoglobulin comprising a protection protein in association with an immunoglobulin derived heavy chain having at least a portion of an antigen binding domain.
96.    The immunoglobulin of claim 95 further  
10 comprising an immunoglobulin derived light chain having at least a portion of an antigen binding domain associated with said immunoglobulin derived heavy chain.
97.    The immunoglobulin of claim 95 further  
15 comprising a second immunoglobulin derived heavy chain having at least a portion of an antigen binding domain associated with said protection protein.
98.    The immunoglobulin of claim 97 further  
20 comprising at least one immunoglobulin derived light chain having at least a portion of an antigen binding domain bound to said second immunoglobulin derived heavy chain.
99.    The immunoglobulin of claim 95 further  
25 comprising an immunoglobulin J chain bound to said immunoglobulin derived heavy chain and optionally to a second immunoglobulin derived heavy chain.
100.   The immunoglobulin of claim 95 that is a therapeutic immunoglobulin.
101.   The immunoglobulin of claim 100 wherein said  
30 therapeutic immunoglobulin binds to mucosal pathogen antigens.
102.   The immunoglobulin of claim 101 that is capable of preventing dental caries.

103. The immunoglobulin of claim 95 wherein said antigen binding domain is capable of binding an antigen from S. mutans serotypes c, e and f or S.sobrinus serotypes d and g.

5        104. The immunoglobulin of claim 95 wherein said protection protein has an amino acid sequence which substantially corresponds to at least a portion of the amino acid residues selected from the group consisting of 1 to 627 and 1 to 606 of the rabbit polyimmunoglobulin  
10 receptor and wherein said protection protein does not have an amino acid residue sequence corresponding to amino acid residues 628-755 of the rabbit polyimmunoglobulin receptor.

15        105. The immunoglobulin of claim 104 wherein said protection protein has an amino acid sequence which does not contain amino acid residues corresponding to amino acid residues 628 to 775 of the rabbit polyimmunoglobulin receptor and which does contain amino acid residues which correspond to one or more of the following amino acid  
20 segments:

- a) amino acids corresponding to amino acid residues 21-43 of the rabbit polyimmunoglobulin receptor;
- b) amino acids corresponding to amino acid residues 1 - 118 of the rabbit polyimmunoglobulin receptor;
- 25        c) amino acids corresponding to amino acid residues 119 - 223 of the rabbit polyimmunoglobulin receptor;
- d) amino acids corresponding to amino acid residues 224 - 332 of the rabbit polyimmunoglobulin receptor;
- e) amino acids corresponding to amino acid residues 333 - 441 of the rabbit polyimmunoglobulin receptor;
- 30        f) amino acids corresponding to amino acid residues 442 - 552 of the rabbit polyimmunoglobulin receptor;

g) amino acids corresponding to amino acid residues 553 - 606 or 553 - 627 of the rabbit poly-immunoglobulin receptor.

106. The immunoglobulin of claim 95 wherein said  
5 protection protein has an amino acid sequence which does not contain amino acid residues of a polyimmunoglobulin receptor of a species which are analogous to amino acid residues 628 to 755 of the rabbit polyimmunoglobulin receptor and which does contain amino acid residues from  
10 a polyimmunoglobulin receptor of a species which are analogous to one or more of the following amino acid segments:

- a) amino acids corresponding to amino acid residues 21 - 43 of the rabbit polyimmunoglobulin receptor;
- 15 b) amino acids corresponding to amino acid residues 1 - 118 of the rabbit polyimmunoglobulin receptor;
- c) amino acids corresponding to amino acid residues 119 - 223 of the rabbit polyimmunoglobulin receptor;
- d) amino acids corresponding to amino acid residues  
20 224 - 332 of the rabbit polyimmunoglobulin receptor;
- e) amino acids corresponding to amino acid residues 333 - 441 of the rabbit polyimmunoglobulin receptor;
- f) amino acids corresponding to amino acid residues 442 - 552 of the rabbit polyimmunoglobulin  
25 receptor;
- g) amino acids corresponding to amino acid residues 553 - 606 or 553 - 627 of the rabbit polyimmunoglobulin receptor.

107. The immunoglobulin of claim 106 wherein said  
30 species is human.

108. The immunoglobulin of claim 95 wherein said protection protein includes the amino acid sequence of at least one of the domains selected from the group consist-

ing of the following portions of the rabbit polyimmuno-  
globulin receptor: domain I, domain II, domain III,  
domain IV, domain V, and amino acid residues 553 to 627  
of domain VI; and does not have an amino acid sequence  
5 corresponding to amino acid residues 628-755 of the  
rabbit polyimmunoglobulin receptor.

109. The immunoglobulin of claim 95 wherein said  
protection protein does not have any amino acid sequence  
which corresponds to or is analogous to amino acid resi-  
10 dues 628-755 of the rabbit polyimmunoglobulin receptor  
and which does include:

a) at least one domain which is from the  
polyimmunoglobulin receptor of a first animal and which  
is analogous to at least a portion of the following amino  
15 acid segments of the rabbit polyimmunoglobulin receptor:  
domain I, domain II, domain III, domain IV, domain V, and  
amino acid residues 553 to 627 of domain VI;

b) at least one domain which is from the  
polyimmunoglobulin receptor of a second animal and which  
20 corresponds to or is analogous to the following amino  
acid residue segments of the rabbit polyimmunoglobulin  
receptor: domain I, domain II, domain III, domain IV,  
domain V, and amino acid residues 553 to 627 of domain  
VI.

25 110. The immunoglobulin of claim 95 wherein said  
protection protein does not have any amino acid sequence  
which corresponds to or is analogous to amino acid resi-  
dues 628-755 of the rabbit polyimmunoglobulin receptor  
and which does include:

30 a) at least one amino acid segment which is from  
the polyimmunoglobulin receptor of a first animal and  
which is analogous to at least a portion of the following  
amino acid residue segments of the rabbit



polyimmunoglobulin receptor: domain I, domain II, domain III, domain IV, domain V, and amino acid residues 553 to 627 of domain VI;

b) at least one amino acid segment which is from  
5 the polyimmunoglobulin receptor of a second animal and which corresponds to or is analogous to the following amino acid residue segments of the rabbit polyimmunoglobulin receptor: domain I, domain II, domain III, domain IV, domain V, and amino acid residues 553 to  
10 627 of domain VI.

111. The immunoglobulin of claim 110 wherein said first animal is a mammal and said second animal is a rabbit.

112. The immunoglobulin of claim 110 wherein said  
15 first animal is a human and said second animal is a rabbit.

113. The immunoglobulin of claim 95 wherein said immunoglobulin derived heavy chain contains at least a portion of an IgM or IgA heavy chain of any subtype.

20 114. The immunoglobulin of claim 95 wherein said immunoglobulin derived heavy chain is comprised of immunoglobulin domains from two different isotopes of immunoglobulin.

115. The immunoglobulin of claim 115 wherein said  
25 immunoglobulin domains are selected from the group consisting of:

- a) the C<sub>H</sub>1 of a mouse IgG1 and the C<sub>H</sub>2 and C<sub>H</sub>3 of mouse IgA; and
- b) the C<sub>H</sub>1 and C<sub>H</sub>2 of a mouse IgG1 and the C<sub>H</sub>2 and  
30 C<sub>H</sub>3 of mouse IgA;

116. The immunoglobulin of claim 95 wherein said antigen binding domain substantially corresponds to the Guy's 13 heavy chain variable region.

117. The immunoglobulin of claim 96 wherein said antigen binding domain substantially corresponds to the Guy's 13 light chain variable region.

118. A composition comprising the immunoglobulin of  
5 any of claims 95-117 and at least one plant macromolecule.

119. The composition of claim 118 wherein said plant macromolecule is derived from a dicotyledonous, monocotyledonous, solanaceous, alfalfa or tobacco plant.

10 120. The composition of claim 118 wherein said plant macromolecule is selected from the group consisting of ribulose biphosphate carboxylase, light harvesting complex, pigments, secondary metabolites and chlorophyll.

121. The composition of claim 118 wherein said  
15 immunoglobulin is present in a concentration of between 0.001% and 99% mass excluding water.

122. The composition of claim 119 wherein said plant macromolecules are present in a concentration of between 1% and 99% mass excluding water.

20 123. A method of producing the immunoglobulin of any of claims 95-117 comprising the steps of:

(a) introducing into a plant cell an expression vector containing a nucleotide sequence encoding a protection protein operably linked to a transcriptional  
25 promoter; and

(b) introducing into said plant cell an expression vector containing a nucleotide sequence encoding an immunoglobulin derived heavy chain having at least a portion of an antigen binding domain operably linked to a  
30 transcriptional promoter.

124. The method of claim 123 further comprising the step of:

(c) introducing into said plant cell an expression vector containing a nucleotide sequence encoding an immunoglobulin derived light chain having at least a portion of an antigen binding domain operably linked to a transcriptional promoter.

125. The method of claim 123 further comprising the step of introducing into said plant cell an expression vector containing a nucleotide sequence encoding an immunoglobulin J chain operably linked to a transcriptional promoter.

126. The method of claim 123 wherein said immunoglobulin derived heavy chain is immunoglobulin alpha chain and said immunoglobulin derived light chain is an immunoglobulin kappa or lambda chain.

127. The method of claim 123 wherein said immunoglobulin derived heavy chain is comprised of portions of immunoglobulin alpha chain and immunoglobulin gamma chain.

128. The method of claim 123 wherein said plant cells are part of a plant.

129. The method of claim 123 further comprising growing said plant cells into a regenerated plant.

130. The method of claims 128 or 129 wherein said plant is a dicotyledonous, monocotyledonous, solanaceous, leguminous, alfalfa or tobacco plant.

131. The method of claim 123 wherein said immunoglobulin derived heavy chain is a chimeric immunoglobulin heavy chain.

132. A method of producing a therapeutic immunoglobulin composition containing plant macromolecules, said method comprising the step of shearing under pressure plants or parts thereof to produce a pulp containing a therapeutic immunoglobulin and plant macro-

molecule mixture, said immunoglobulin comprising a protection protein, and wherein said immunoglobulin is encoded by at least one nucleic acid sequence that has been introduced into the cells of said plants.

5        133. The method of claim 132 further comprising the step of separating said solid plant derived material from said liquid.

10        134. The method of claim 132 wherein said portion of said plant is a leaf, stem, root, tuber, fruit or entire plant.

135. The method of claim 132 wherein said shearing is accomplished by a mechanical device which releases liquid from the apoplast or symplast of said plant.

15        136. The method of claim 133 wherein said separation is by centrifugation, settling, flocculation or filtration.

137. A method for producing an assembled immunoglobulin molecule having heavy, light and J chains and a protection protein comprising the steps of:

20        a) introducing into a eukaryotic cell nucleotide sequences operably linked for expression encoding:

- 25            i) an immunoglobulin derived heavy chain having at least a portion of an antigen binding domain,
- ii) an immunoglobulin derived light chain having at least a portion of an antigen binding domain,
- iii) an immunoglobulin J chain, and
- iv) a protection protein; and

30        b) maintaining said cell under conditions allowing production and assembly of said immunoglobulin derived heavy and light chains, said immunoglobulin J

chain and said protection protein into an immunoglobulin molecule.

138. A method for producing an assembled immunoglobulin molecule having heavy, light and J chains and a protection protein by maintaining under conditions allowing protein production and immunoglobulin assembly, a eukaryotic cell containing nucleotide sequences operably linked for expression encoding:

- i) an immunoglobulin derived heavy chain having at least a portion of an antigen binding domain,
- ii) an immunoglobulin derived light chain having at least a portion of an antigen binding domain,
- iii) an immunoglobulin J chain, and
- iv) a protection protein.

139. The method of claims 137 or 138 wherein said eukaryotic cell is a plant cell.

140. A method of making an immunoglobulin resistant to environmental conditions comprising the steps of :

a) operably linking a nucleotide sequence encoding at least a portion of the antigen binding domain derived from an immunoglobulin heavy chain to a nucleotide sequence encoding at least one domain derived from an immunoglobulin alpha heavy chain to form a nucleotide sequence encoding a chimeric immunoglobulin heavy chain;

b) expressing said nucleotide sequence encoding said chimeric immunoglobulin heavy chain to produce said chimeric immunoglobulin heavy chain in a plant cell which also contains at least one other molecule selected from the group consisting of: a protection protein, an immunoglobulin derived light chain having at least a

portion of an antigen binding domain and an immunoglobulin J chain; and  
thereby allowing the chimeric immunoglobulin heavy chain to assemble with said at least one other molecule to form  
5 said immunoglobulin resistant to said environmental conditions.

141. The method of claim 140 wherein said other molecule is a protection protein and said plant cell also contains an immunoglobulin derived light chain having at  
10 least a portion of an antigen binding domain and an immunoglobulin J chain.

142. A process for producing an immunoglobulin resistant to environmental conditions by maintaining under conditions allowing protein production and  
15 immunoglobulin assembly a plant cell containing:

a) a nucleotide sequence encoding a chimeric immunoglobulin heavy chain in which a nucleotide sequence encoding at least a portion of an antigen binding domain derived from heavy chain is operably linked to a  
20 nucleotide sequence encoding at least one domain derived from an immunoglobulin alpha heavy chain; and

b) at least one other molecule selected from the group consisting of: a protection protein, an immunoglobulin derived light chain having at least a portion of  
25 an antigen binding domain and an immunoglobulin J chain; thereby allowing the chimeric immunoglobulin heavy chain to assemble with said at least one other molecule to form said immunoglobulin resistant to said environmental conditions.

30 143. The immunoglobulin of claim 95 wherein said chimeric immunoglobulin heavy chain contains an immunoglobulin domain from one of the following immunoglobulin

heavy chains: IgG, IgA, IgM, IgE, IgD; and also contains a protection protein-binding domain from IgA or IgM.

144. The immunoglobulin of claim 143 wherein said immunoglobulin heavy chains are human, rodent, rabbit,  
5 bovine, ovine, caprine, fowl, canine, feline or primate immunoglobulin heavy chains.

145. The immunoglobulin of claim 143 wherein said protection protein-binding domain is from the IgA of a human, rodent, rabbit, bovine, ovine, canine, feline or  
10 primate.

146. The immunoglobulin of claim 143 wherein said chimeric immunoglobulin heavy chain is comprised of immunoglobulin chains of mouse IgG1 and said protection protein-binding domain is from mouse IgA or IgM.

15 147. The immunoglobulin of claim 143 wherein said chimeric immunoglobulin heavy chain is comprised of immunoglobulin domains of a human IgG, IgM, IgD or IgE and said protection protein-binding domain is from a human IgA or IgM.